

# An Introduction to *GenomeInfoDb*

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## 1 Introduction

---

The *GenomeInfoDb* provides an interface to access seqlevelsStyles (such as UCSC, NCBI, Ensembl) and their supported mappings for organisms. For instance, for Homo sapiens, seqlevelsStyle "UCSC" maps to "chr1", "chr2", ..., "chrX", "chrY". The section below introduces these functions with examples.

## 2 Functionality for all existing organisms

### 2.1 genomeStyles

The `genomeStyles` lists out for each organism, the seqlevelsStyles and their mappings.

```
seqmap <- genomeStyles()
head(seqmap,n=2)

## $Arabidopsis_thaliana
##   circular auto sex NCBI TAIR9 Ensembl
## 1  FALSE  TRUE FALSE   1  Chr1      1
## 2  FALSE  TRUE FALSE   2  Chr2      2
## 3  FALSE  TRUE FALSE   3  Chr3      3
## 4  FALSE  TRUE FALSE   4  Chr4      4
## 5  FALSE  TRUE FALSE   5  Chr5      5
## 6   TRUE FALSE FALSE  MT  ChrM      Mt
## 7   TRUE FALSE  TRUE Pltd ChrC      Pt
##
## $Caenorhabditis_elegans
##   circular auto sex NCBI UCSC Ensembl
## 1  FALSE  TRUE FALSE   I  chrI      I
## 2  FALSE  TRUE FALSE  II  chrII     II
## 3  FALSE  TRUE FALSE III chrIII     III
## 4  FALSE  TRUE FALSE  IV  chrIV     IV
## 5  FALSE  TRUE FALSE   V  chrV      V
## 6  FALSE FALSE  TRUE   X  chrX      X
## 7   TRUE  TRUE FALSE  MT  chrM     MtDNA
```

Organism's supported by GenomeInfoDb can be found by :

```
names(genomeStyles())

## [1] "Arabidopsis_thaliana"      "Caenorhabditis_elegans"
## [3] "Canis_familiaris"         "Cyanidioschyzon_merolae"
## [5] "Drosophila_melanogaster"   "Gossypium_hirsutum"
## [7] "Homo_sapiens"              "Mus_musculus"
## [9] "Oryza_sativa"              "Populus_trichocarpa"
## [11] "Rattus_norvegicus"         "Saccharomyces_cerevisiae"
## [13] "Zea_mays"
```

If one knows the organism one is interested in, then we can directly access the information for the given organism along. Each function accepts an argument called species which as "genus species", the default is "Homo sapiens". In the following example we list out only the first five entries returned by the code snippet.

```
head(genomeStyles("Homo_sapiens"),5)

##   circular auto sex NCBI UCSC dbSNP Ensembl
## 1  FALSE  TRUE FALSE   1 chr1   ch1      1
## 2  FALSE  TRUE FALSE   2 chr2   ch2      2
## 3  FALSE  TRUE FALSE   3 chr3   ch3      3
## 4  FALSE  TRUE FALSE   4 chr4   ch4      4
## 5  FALSE  TRUE FALSE   5 chr5   ch5      5
```

We can also check if a given style is supported by GenomeInfoDb for a given species. For example, if we want to know if "UCSC" mapping is supported for "Homo sapiens" we can ask :

```
"UCSC" %in% names(genomeStyles("Homo_sapiens"))  
## [1] TRUE
```

## 2.2 extractSeqlevels

We can also extract the desired seqlevelsStyle from a given organism using the `extractSeqlevels`

```
extractSeqlevels(species="Arabidopsis_thaliana", style="NCBI")  
## [1] "1" "2" "3" "4" "5" "MT" "Pltd"
```

## 2.3 extractSeqlevelsByGroup

We can also extract the desired seqlevelsStyle from a given organism based on a group (Group - 'auto' denotes autosomes, 'circular' denotes circular chromosomes and 'sex' denotes sex chromosomes; the default is all chromosomes are returned).

```
extractSeqlevelsByGroup(species="Arabidopsis_thaliana", style="NCBI",  
                        group="auto")  
## [1] "1" "2" "3" "4" "5"
```

## 2.4 seqlevelsStyle

We can find the seqname Style for a given character vector by using the `seqlevelsStyle`

```
seqlevelsStyle(paste0("chr", c(1:30)))  
## [1] "UCSC"  
seqlevelsStyle(c("2L", "2R", "X", "Xhet"))  
## [1] "NCBI"
```

## 2.5 seqlevelsInGroup

We can also subset a given character vector containing seqnames using the `seqlevelsInGroup`. We currently support 3 groups: 'auto' for autosomes, 'sex' for allosomes/sex chromosomes and circular for 'circular' chromosomes. The user can also provide the style and species they are working with. In the following examples, we extract the sex, auto and circular chromosomes for Homo sapiens :

```
newchr <- paste0("chr", c(1:22, "X", "Y", "M", "1_gl000192_random", "4_ctg9_hap1"))  
seqlevelsInGroup(newchr, group="sex")
```

```
## [1] "chrX" "chrY"

seqlevelsInGroup(newchr, group="auto")

## [1] "chr1" "chr2" "chr3" "chr4" "chr5" "chr6" "chr7" "chr8" "chr9"
## [10] "chr10" "chr11" "chr12" "chr13" "chr14" "chr15" "chr16" "chr17" "chr18"
## [19] "chr19" "chr20" "chr21" "chr22"

seqlevelsInGroup(newchr, group="circular")

## [1] "chrM"

seqlevelsInGroup(newchr, group="sex", "Homo_sapiens", "UCSC")

## [1] "chrX" "chrY"
```

if we have a vector containing seqnames and we want to verify the species and style for them, we can use:

```
seqnames <- c("chr1", "chr9", "chr2", "chr3", "chr10")
all(seqnames %in% extractSeqlevels("Homo_sapiens", "UCSC"))

## [1] TRUE
```

## 2.6 mapSeqlevels

Returns a matrix with 1 column per supplied sequence name and 1 row per sequence renaming map compatible with the specified style. If `best.only` is `TRUE` (the default), only the "best" renaming maps (i.e. the rows with less NAs) are returned.

```
mapSeqlevels(c("chrII", "chrIII", "chrM"), "NCBI")

## chrII chrIII chrM
## "II" "III" "MT"
```

We also have several seqlevel utility functions. Let us construct a basic `GRanges` and show how these functions can be used.

```
gr <- GRanges(paste0("ch", 1:35), IRanges(1:35, width=5))
gr

## GRanges object with 35 ranges and 0 metadata columns:
##      seqnames      ranges strand
##      <Rle> <IRanges> <Rle>
##      [1]      ch1       1-5      *
##      [2]      ch2       2-6      *
##      [3]      ch3       3-7      *
##      [4]      ch4       4-8      *
##      [5]      ch5       5-9      *
##      ...      ...       ...      ...
##      [31]     ch31      31-35      *
##      [32]     ch32      32-36      *
##      [33]     ch33      33-37      *
##      [34]     ch34      34-38      *
##      [35]     ch35      35-39      *
##      -----
```

```
## seqinfo: 35 sequences from an unspecified genome; no seqlengths
```

As you can see, we have "ch" instead of "chr" for chromosome names. We can use `renameSeqlevels` to change the "ch" to "chr"

## 2.7 renameSeqlevels

As the first argument - it takes the object whose seqlevels we need to change, and as the second argument it takes a named vector which has the changes.

```
newnames <- paste0("chr",1:35)
names(newnames) <- paste0("ch",1:35)
head(newnames)

##      ch1      ch2      ch3      ch4      ch5      ch6
## "chr1" "chr2" "chr3" "chr4" "chr5" "chr6"

gr <- renameSeqlevels(gr,newnames)
gr

## GRanges object with 35 ranges and 0 metadata columns:
##      seqnames      ranges strand
##      <Rle> <IRanges> <Rle>
##      [1]      chr1         1-5      *
##      [2]      chr2         2-6      *
##      [3]      chr3         3-7      *
##      [4]      chr4         4-8      *
##      [5]      chr5         5-9      *
##      ...      ...         ...      ...
##      [31]     chr31        31-35      *
##      [32]     chr32        32-36      *
##      [33]     chr33        33-37      *
##      [34]     chr34        34-38      *
##      [35]     chr35        35-39      *
##      -----
##      seqinfo: 35 sequences from an unspecified genome; no seqlengths
```

Humans have just 22 primary chromosomes - but here we have some extra seqlevels which we want to remove - there are several ways we can achieve this:

## 2.8 dropSeqlevels

Here the second argument is the seqlevels that you want to drop. Because these seqlevels are in use (i.e. have ranges on them), the ranges on these sequences need to be removed before the seqlevels can be dropped. We call this *pruning*. The `pruning.mode` argument controls how to prune `gr`. Unlike for list-like objects (e.g. `GRangesList`) for which pruning can be done in various ways, pruning a `GRanges` object is straightforward and achieved by specifying `pruning.mode="coarse"`.

```
dropSeqlevels(gr, paste0("chr",23:35), pruning.mode="coarse")

## GRanges object with 22 ranges and 0 metadata columns:
##      seqnames      ranges strand
```

```
##          <Rle> <IRanges> <Rle>
## [1]      chr1      1-5      *
## [2]      chr2      2-6      *
## [3]      chr3      3-7      *
## [4]      chr4      4-8      *
## [5]      chr5      5-9      *
## ...      ...      ...      ...
## [18]     chr18     18-22     *
## [19]     chr19     19-23     *
## [20]     chr20     20-24     *
## [21]     chr21     21-25     *
## [22]     chr22     22-26     *
## -----
## seqinfo: 22 sequences from an unspecified genome; no seqlengths
```

## 2.9 keepSeqlevels

Here the second argument is the seqlevels that you want to keep.

```
keepSeqlevels(gr, paste0("chr",1:22), pruning.mode="coarse")

## GRanges object with 22 ranges and 0 metadata columns:
##      seqnames      ranges strand
##      <Rle> <IRanges> <Rle>
## [1]      chr1      1-5      *
## [2]      chr2      2-6      *
## [3]      chr3      3-7      *
## [4]      chr4      4-8      *
## [5]      chr5      5-9      *
## ...      ...      ...      ...
## [18]     chr18     18-22     *
## [19]     chr19     19-23     *
## [20]     chr20     20-24     *
## [21]     chr21     21-25     *
## [22]     chr22     22-26     *
## -----
## seqinfo: 22 sequences from an unspecified genome; no seqlengths
```

## 2.10 keepStandardChromosomes

This function internally uses the pre-defined tables inside GenomeInfoDb to find the correct seqlevels according to the sequence style of the object.

```
keepStandardChromosomes(gr, pruning.mode="coarse")

## GRanges object with 35 ranges and 0 metadata columns:
##      seqnames      ranges strand
##      <Rle> <IRanges> <Rle>
## [1]      chr1      1-5      *
## [2]      chr2      2-6      *
## [3]      chr3      3-7      *
```

```
##      [4]      chr4      4-8      *
##      [5]      chr5      5-9      *
##      ...      ...      ...      ...
##     [31]     chr31     31-35     *
##     [32]     chr32     32-36     *
##     [33]     chr33     33-37     *
##     [34]     chr34     34-38     *
##     [35]     chr35     35-39     *
## -----
## seqinfo: 35 sequences from an unspecified genome; no seqlengths
```

One can also specify the optional species argument to be more precise.

```
plantgr <- GRanges(c(1:5,"MT","Pltd"), IRanges(1:7,width=5))
keepStandardChromosomes(plantgr, species="Arabidopsis thaliana",
                        pruning.mode="coarse")

## GRanges object with 7 ranges and 0 metadata columns:
##      seqnames      ranges strand
##      <Rle> <IRanges> <Rle>
##     [1]      1      1-5      *
##     [2]      2      2-6      *
##     [3]      3      3-7      *
##     [4]      4      4-8      *
##     [5]      5      5-9      *
##     [6]      MT      6-10     *
##     [7]     Pltd      7-11     *
## -----
## seqinfo: 7 sequences from an unspecified genome; no seqlengths
```

### 3 Seqinfo objects

```
## Note that all the arguments (except 'genome') must have the
## same length. 'genome' can be of length 1, whatever the lengths
## of the other arguments are.
x <- Seqinfo(seqnames=c("chr1", "chr2", "chr3", "chrM"),
             seqlengths=c(100, 200, NA, 15),
             isCircular=c(NA, FALSE, FALSE, TRUE),
             genome="toy")

length(x)

## [1] 4

seqnames(x)

## [1] "chr1" "chr2" "chr3" "chrM"

names(x)

## [1] "chr1" "chr2" "chr3" "chrM"

seqlevels(x)
```

```
## [1] "chr1" "chr2" "chr3" "chrM"

seqlengths(x)

## chr1 chr2 chr3 chrM
## 100 200 NA 15

isCircular(x)

## chr1 chr2 chr3 chrM
## NA FALSE FALSE TRUE

genome(x)

## chr1 chr2 chr3 chrM
## "toy" "toy" "toy" "toy"

x[c("chrY", "chr3", "chr1")] # subset by names

## Seqinfo object with 3 sequences from 2 genomes (NA, toy):
## seqnames seqlengths isCircular genome
## chrY NA NA <NA>
## chr3 NA FALSE toy
## chr1 100 NA toy

## Rename, drop, add and/or reorder the sequence levels:
xx <- x
seqlevels(xx) <- sub("chr", "ch", seqlevels(xx)) # rename
xx

## Seqinfo object with 4 sequences (1 circular) from toy genome:
## seqnames seqlengths isCircular genome
## ch1 100 NA toy
## ch2 200 FALSE toy
## ch3 NA FALSE toy
## chM 15 TRUE toy

seqlevels(xx) <- rev(seqlevels(xx)) # reorder
xx

## Seqinfo object with 4 sequences (1 circular) from toy genome:
## seqnames seqlengths isCircular genome
## chM 15 TRUE toy
## ch3 NA FALSE toy
## ch2 200 FALSE toy
## ch1 100 NA toy

seqlevels(xx) <- c("ch1", "ch2", "chY") # drop/add/reorder
xx

## Seqinfo object with 3 sequences from 2 genomes (toy, NA):
## seqnames seqlengths isCircular genome
## ch1 100 NA toy
## ch2 200 FALSE toy
## chY NA NA <NA>

seqlevels(xx) <- c(chY="Y", ch1="1", "22") # rename/reorder/drop/add
xx
```



```
## Seqinfo object with 3 sequences from 2 genomes (NA, toy):
##   seqnames seqlengths isCircular genome
##   Y          NA          NA    <NA>
##   1          100         NA     toy
##   22         NA          NA    <NA>

y <- Seqinfo(seqnames=c("chr3", "chr4", "chrM"),
              seqlengths=c(300, NA, 15))

y

## Seqinfo object with 3 sequences from an unspecified genome:
##   seqnames seqlengths isCircular genome
##   chr3      300         NA    <NA>
##   chr4      NA          NA    <NA>
##   chrM      15          NA    <NA>

merge(x, y) # rows for chr3 and chrM are merged

## Warning in .merge_two_Seqinfo_objects(x, y): Each of the 2 combined objects
has sequence levels not in the other:
##   - in 'x': chr1, chr2
##   - in 'y': chr4
## Make sure to always combine/compare objects based on the same reference
## genome (use suppressWarnings() to suppress this warning).

## Seqinfo object with 5 sequences (1 circular) from 2 genomes (toy, NA):
##   seqnames seqlengths isCircular genome
##   chr1      100         NA     toy
##   chr2      200        FALSE    toy
##   chr3      300        FALSE    toy
##   chrM      15         TRUE     toy
##   chr4      NA          NA    <NA>

suppressWarnings(merge(x, y))

## Seqinfo object with 5 sequences (1 circular) from 2 genomes (toy, NA):
##   seqnames seqlengths isCircular genome
##   chr1      100         NA     toy
##   chr2      200        FALSE    toy
##   chr3      300        FALSE    toy
##   chrM      15         TRUE     toy
##   chr4      NA          NA    <NA>

## Note that, strictly speaking, merging 2 Seqinfo objects is not
## a commutative operation, i.e., in general 'z1 <- merge(x, y)'
## is not identical to 'z2 <- merge(y, x)'. However 'z1' and 'z2'
## are guaranteed to contain the same information (i.e. the same
## rows, but typically not in the same order):
suppressWarnings(merge(y, x))

## Seqinfo object with 5 sequences (1 circular) from 2 genomes (toy, NA):
##   seqnames seqlengths isCircular genome
##   chr3      300        FALSE    toy
##   chr4      NA          NA    <NA>
##   chrM      15         TRUE     toy
```

```
## chr1      100      NA    toy
## chr2      200     FALSE  toy

## This contradicts what 'x' says about circularity of chr3 and chrM:
isCircular(y)[c("chr3", "chrM")] <- c(TRUE, FALSE)
y

## Seqinfo object with 3 sequences (1 circular) from an unspecified genome:
## seqnames seqlengths isCircular genome
## chr3      300      TRUE  <NA>
## chr4      NA      NA    <NA>
## chrM      15     FALSE  <NA>

if (interactive()) {
  merge(x, y) # raises an error
}
```

## 4 Examples

### 4.1 converting seqlevel styles (eg:UCSC to NCBI)

A quick example using *Drosophila Melanogaster*. The txdb object contains seqlevels in UCSC style, we want to convert them to NCBI

```
txdb <- TxDb.Dmelanogaster.UCSC.dm3.ensGene
seqlevels(txdb)

## [1] "chr2L"      "chr2R"      "chr3L"      "chr3R"      "chr4"      "chrX"
## [7] "chrU"       "chrM"       "chr2LHet"   "chr2RHet"   "chr3LHet"   "chr3RHet"
## [13] "chrXHet"    "chrYHet"    "chrUextra"

genomeStyles("Drosophila melanogaster")

##   circular sex auto NCBI      UCSC      Ensembl
## 1  FALSE FALSE TRUE  2L      chr2L      2L
## 2  FALSE FALSE TRUE  2R      chr2R      2R
## 3  FALSE FALSE TRUE  3L      chr3L      3L
## 4  FALSE FALSE TRUE  3R      chr3R      3R
## 5  FALSE FALSE TRUE   4      chr4       4
## 6  FALSE TRUE  FALSE  X      chrX       X
## 7  FALSE TRUE  FALSE  Y      chrY       Y
## 8   TRUE FALSE FALSE  MT      chrM dmel_mitochondrion_genome
## 9  FALSE FALSE FALSE 2LHet  chr2LHet  2LHet
## 10 FALSE FALSE FALSE 2RHet  chr2RHet  2RHet
## 11 FALSE FALSE FALSE 3LHet  chr3LHet  3LHet
## 12 FALSE FALSE FALSE 3RHet  chr3RHet  3RHet
## 13 FALSE FALSE FALSE Xhet   chrXHet  XHet
## 14 FALSE FALSE FALSE Yhet   chrYHet  YHet
## 15 FALSE FALSE FALSE  Un     chrU     U
## 16 FALSE FALSE FALSE <NA> chrUextra Uextra

mapSeqlevels(seqlevels(txdb), "NCBI")
```

```
##      chr2L      chr2R      chr3L      chr3R      chr4      chrX      chrU
##      "2L"      "2R"      "3L"      "3R"      "4"      "X"      "Un"
##      chrM chr2LHet chr2Rhet chr3LHet chr3Rhet chrXHet chrYHet
##      "MT"  "2LHet"  "2Rhet"  "3LHet"  "3Rhet"  "Xhet"  "Yhet"
## chrUextra
##      NA
```

## 4.2 converting styles and removing unwanted seqlevels

Suppose we read in a Bam file or a BED file and the resulting GRanges have a lot of seqlevels which are not required by your analysis or you want to rename the seqlevels from the current style to your own style (eg:UCSC to NCBI), we can use the functionality provided by GenomeInfoDb to do that.

Let us say that we have extracted the seqlevels of the Seqinfo object(say GRanges from a BED file) in a variable called "sequence".

```
sequence <- seqlevels(x)

## sequence is in UCSC format and we want NCBI style
newStyle <- mapSeqlevels(sequence,"NCBI")
newStyle <- newStyle[complete.cases(newStyle)] # removing NA cases.

## rename the seqlevels
x <- renameSeqlevels(x,newStyle)

## keep only the seqlevels you want (say autosomes)
auto <- extractSeqlevelsByGroup(species="Homo sapiens", style="NCBI",
                                group="auto")
x <- keepSeqlevels(x,auto)
```

## 5 Session Information

Here is the output of `sessionInfo` on the system on which this document was compiled:

```
toLatex(sessionInfo())
```

- R version 4.5.2 (2025-10-31), x86\_64-pc-linux-gnu
- Locale: LC\_CTYPE=en\_US.UTF-8, LC\_NUMERIC=C, LC\_TIME=en\_US.UTF-8, LC\_COLLATE=C, LC\_MONETARY=en\_US.UTF-8, LC\_MESSAGES=en\_US.UTF-8, LC\_PAPER=en\_US.UTF-8, LC\_NAME=C, LC\_ADDRESS=C, LC\_TELEPHONE=C, LC\_MEASUREMENT=en\_US.UTF-8, LC\_IDENTIFICATION=C
- Time zone: Etc/UTC
- TZcode source: system (glibc)
- Running under: Ubuntu 24.04.3 LTS
- Matrix products: default
- BLAS: /usr/lib/x86\_64-linux-gnu/openblas-pthread/libblas.so.3

- LAPACK:  
/usr/lib/x86\_64-linux-gnu/openblas-pthread/libopenblas-p-r0.3.26.so ;  
LAPACK version 3.12.0
- Base packages: base, datasets, grDevices, graphics, methods, stats, stats4, utils
- Other packages: AnnotationDbi 1.72.0, Biobase 2.70.0, BiocGenerics 0.56.0,  
BiocStyle 2.38.0, GenomeInfoDb 1.46.2, GenomicFeatures 1.62.0,  
GenomicRanges 1.62.1, IRanges 2.44.0, S4Vectors 0.48.0, Seqinfo 1.0.0,  
TxDb.Dmelanogaster.UCSC.dm3.ensGene 3.2.2, generics 0.1.4
- Loaded via a namespace (and not attached): BiocIO 1.20.0, BiocManager 1.30.27,  
BiocParallel 1.44.0, Biostrings 2.78.0, DBI 1.2.3, DelayedArray 0.36.0,  
GenomicAlignments 1.46.0, KEGGREST 1.50.0, Matrix 1.7-4, MatrixGenerics 1.22.0,  
R6 2.6.1, RCurl 1.98-1.17, RSQLite 2.4.5, Rsamtools 2.26.0, S4Arrays 1.10.1,  
SparseArray 1.10.8, SummarizedExperiment 1.40.0, UCSC.utils 1.6.1, XML 3.99-0.20,  
XVector 0.50.0, abind 1.4-8, bit 4.6.0, bit64 4.6.0-1, bitops 1.0-9, blob 1.3.0,  
bslib 0.10.0, buildtools 1.0.0, cachem 1.1.0, cigarillo 1.0.0, cli 3.6.5, codetools 0.2-20,  
compiler 4.5.2, crayon 1.5.3, curl 7.0.0, digest 0.6.39, evaluate 1.0.5, fastmap 1.2.0,  
grid 4.5.2, highr 0.11, htmltools 0.5.9, httr 1.4.7, jquerylib 0.1.4, jsonlite 2.0.0,  
knitr 1.51, lattice 0.22-7, lifecycle 1.0.5, maketools 1.3.2, matrixStats 1.5.0,  
memoise 2.0.1, parallel 4.5.2, pkgconfig 2.0.3, png 0.1-8, restfulr 0.0.16, rjson 0.2.23,  
rlang 1.1.7, rmarkdown 2.30, rtracklayer 1.70.1, sass 0.4.10, sys 3.4.3, tools 4.5.2,  
vctrs 0.7.1, xfun 0.56, yaml 2.3.12