# Package 'vissE'

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    Description This package enables the interpretation and analysis of results from a gene set enrichment analysis using network-based and text-mining approaches. Most enrichment analyses re-
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ment analysis using network-based and text-mining approaches. Most enrichment analyses result in large lists of significant gene sets that are difficult to interpret. Tools in this package help build a similarity-based network of significant gene sets from a gene set enrichment analysis that can then be investigated for their biological function using text-mining approaches.

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#### **Description**

This package enables the interpretation and analysis of results from a gene set enrichment analysis using network-based and text-mining approaches. Most enrichment analyses result in large lists of significant gene sets that are difficult to interpret. Tools in this package help build a similarity-based network of significant gene sets from a gene set enrichment analysis that can then be investigated for their biological function using text-mining approaches.

# **Details**

This package supports four workflows to enhance gene set enrichment analysis:

- Clustering results from a gene set enrichment analysis (e.g. using limma::fry, singscore or GSEA). The functions required for this analysis are computeMsigOverlap, computeMsigNetwork and plotMsigNetwork.
- 2. Interpreting gene set clusters (identified in the first analysis) by performing text-mining of gene set names and descriptions. The main function required to perform text-mining of gene sets is plotMsigWordcloud. Other functions can be used to access intermmediate results.
- 3. Visualise gene-level statistics for gene set clusters identified in the first analysis to link back gene set clusters to the genes of interest. This can be done using the plotGeneStats function.
- 4. Identifying gene sets similar to a list of genes identified from a DE analysis using set overlap measures. This can be done using the characteriseGeneset function.

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#### See Also

Useful links:

- https://davislaboratory.github.io/vissE
- Report bugs at https://github.com/DavisLaboratory/vissE/issues

bhuvad\_theme

Custom theme

# Description

Custom theme

## Usage

```
bhuvad_theme(rl = 1.1)
```

# Arguments

rl

a numeric, scaling factor to apply to text sizes

## Value

a ggplot2 theme

#### **Examples**

```
p1 = ggplot2::ggplot()
p1 + bhuvad_theme()
```

characteriseGeneset

Functionally characterise a list of genes

# Description

This function can be used to perform a network-based enrichment analysis of a list of genes. The list of genes are characterised based on their similarity with gene sets from the MSigDB. A network of similar gene sets is retrieved using this function.

## Usage

```
characteriseGeneset(
   gs,
   thresh = 0.2,
   measure = c("ovlapcoef", "jaccard"),
   gscolcs = c("h", "c2", "c5"),
   org = c("auto", "hs", "mm")
)
```

#### **Arguments**

a GeneSet object, representing the list of genes that need to be characterised. gs thresh a numeric, specifying the threshold to discard pairs of gene sets.

a character, specifying the similarity measure to use: ari for the Adjusted Rand measure

Index, jaccard for the Jaccard Index and ovlapcoef for the Overlap Coeffi-

a character, listing the MSigDB collections to use as a background (defaults to h, gscolcs

c2, and c5). Collection types can be retrieved using msigdb::listCollections().

a character, specifying the organism to use. This can either be "auto" (default), org

"hs" or "mm".

#### Value

an igraph object, containing gene sets that are similar to the query set. The network contains relationships between results of the query too.

#### **Examples**

```
library(GSEABase)
data(hgsc)
#create a geneset using one of the Hallmark gene sets
mySet <- GeneSet(</pre>
  geneIds(hgsc[[2]]),
  setName = 'MySet',
  geneIdType = SymbolIdentifier()
#characterise the custom gene set
ig <- characteriseGeneset(mySet)</pre>
plotMsigNetwork(ig)
```

computeMsigNetwork

Compute a network using computed gene set overlap

## **Description**

Computes an igraph object using information on gene sets and gene sets computed using the computeMsigOverlap() function.

# Usage

```
computeMsigNetwork(genesetOverlap, msigGsc)
```

## **Arguments**

genesetOverlap a data.frame, containing results of an overlap analysis computed using the computeMsigOverlap()

function.

a GeneSetCollection object, containing gene sets used to compute overlap. msigGsc

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#### Value

```
an igraph object
```

## **Examples**

```
data(hgsc)
ovlap <- computeMsigOverlap(hgsc)
ig <- computeMsigNetwork(ovlap, hgsc)</pre>
```

computeMsigOverlap

Compute gene set overlap

#### **Description**

Compute overlap between gene sets from a GeneSetCollection using the Jaccard index or the overlap coefficient. These values can then be used to compute a network of gene set overlaps.

## Usage

```
computeMsigOverlap(
  msigGsc1,
  msigGsc2 = NULL,
  thresh = 0.25,
  measure = c("ari", "jaccard", "ovlapcoef")
)
```

# Arguments

msigGsc1 a GeneSetCollection object.

msigGsc2 a GeneSetCollection object or NULL if pairwise overlaps are to be computed.

thresh a numeric, specifying the threshold to discard pairs of gene sets.

measure a character, specifying the similarity measure to use: ari for the Adjusted Rand

Index, jaccard for the Jaccard Index and ovlapcoef for the Overlap Coeffi-

cient.

#### Value

a data.frame, containing the overlap structure of gene sets represented as a network in the simple interaction format (SIF).

```
data(hgsc)
ovlap <- computeMsigOverlap(hgsc)</pre>
```

computeMsigWordFreq Compute word frequencies for a single MSigDB collection

# Description

Compute word frequencies for a single MSigDB collection

# Usage

```
computeMsigWordFreq(
  msigGsc,
  weight = NULL,
  measure = c("tfidf", "tf"),
  version = msigdb::getMsigdbVersions(),
  org = c("auto", "hs", "mm"),
  rmwords = getMsigExclusionList(),
  idf = NULL
)
```

# **Arguments**

msigGsc	a GeneSetCollection object, containing gene sets from the MSigDB. The GSEABase::getBroadSets(function can be used to parse XML files downloaded from MSigDB.
weight	a named numeric vector, containing weights to apply to each gene-set. This can be -log10(FDR), -log10(p-value) or an enrichment score (ideally unsigned).
measure	a character, specifying how frequencies should be computed. "tf" uses term frequencies and "tfidf" (default) applies inverse document frequency weights to term frequencies.
version	a character, specifying the version of msigdb to use (see msigdb::getMsigdbVersions()).
org	a character, specifying the organism to use. This can either be "auto" (default), "hs" or "mm".
rmwords	a character vector, containing an exclusion list of words to discard from the analysis.
idf	a list of named numeric vectors, specifying inverse document frequencies to use to penalise terms from gene-set names and short descriptions. This should be a vector of length 2 with names "Name" and "Short". Numeric vectors should contain weights and names should represent the term. Precomputed versions can be retrieved using the msigdb::getMsigdbIDF().

# Value

a list, containing two data.frames summarising the results of the frequency analysis on gene set names and short descriptions.

```
data(hgsc)
freq <- computeMsigWordFreq(hgsc, measure = 'tfidf')</pre>
```

findMsigClusters 7

findMsigClusters	Identify gene-set cl	lusters from a gene-so	et overlap network
------------------	----------------------	------------------------	--------------------

## **Description**

This function identifies gene-set clusters from a gene-set overlap network produced using vissE. Various graph clustering algorithms from the igraph package can be used for clustering. Gene-set clusters identified are then sorted based on their size and a given statistic of interest (absolute of the statistic is maximised per cluster).

## Usage

```
findMsigClusters(
   ig,
   genesetStat = NULL,
   minSize = 2,
   alg = igraph::cluster_walktrap,
   algparams = list()
)
```

## **Arguments**

ig	an igraph object, containing a network of gene set overlaps computed using computeMsigNetwork().
genesetStat	a named numeric, containing statistics for each gene-set that are to be used in cluster prioritisation. If NULL, clusters are prioritised based on their size (number of gene-sets in them).
minSize	a numeric, stating the minimum size a cluster can be (default is 2).
alg	a function, from the igraph package that should be used to perform graph-clustering (default is igraph::cluster_walktrap). The function should produce a communities object.
algparams	a list, specifying additional parameters that are to be passed to the graph clustering algorithm.

#### **Details**

Gene-sets clusters are identified using graph clustering and are prioritised based on a combination of cluster size and optionally, a statistic of interest (e.g., enrichment scores). A product-of-ranks approach is used to prioritise clusters when gene-set statistics are available. In this approach, clusters are ranked based on their cluster size (largest to smallest) and on the median absolute statistic of gene-sets within it (largest to smallest). The product of these ranks is computed and clusters are ranked based on these product-of-rank statistic (smallest to largest).

When prioritising using cluster size and gene-set statistics, if statistics for some gene-sets in the network are missing, only the size is used in cluster prioritisation.

## Value

a list, containing gene-sets that belong to each cluster. Items in the list are organised based on prioritisation.

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## **Examples**

```
data(hgsc)
ovlap <- computeMsigOverlap(hgsc, thresh = 0.25)
ig <- computeMsigNetwork(ovlap, hgsc)
findMsigClusters(ig)</pre>
```

#### **Description**

List of words to discard when performing text mining MSigDB gene set names and short descriptions.

## Usage

```
getMsigExclusionList(custom = c())
```

#### **Arguments**

custom

a character vector, containing list of words to add onto existing exclusion list.

#### Value

a character vector, containing words to be excluded from the text mining analysis.

## **Examples**

```
getMsigExclusionList('remove')
```

hgsc

The Hallmark collection from the MSigDB

## **Description**

The molecular signatures database (MSigDB) is a collection of over 25000 gene expression signatures. Signatures in v7.2 are divided into 9 categories. The Hallmarks collection contains gene expression signatures representing molecular processes that are hallmarks in cancer development and progression.

# Usage

hgsc

## **Format**

A GeneSetCollection object with 50 GeneSet objects representing the 50 Hallmark gene expression signatures.

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#### References

Subramanian, A., Tamayo, P., Mootha, V. K., Mukherjee, S., Ebert, B. L., Gillette, M. A., ... & Mesirov, J. P. (2005). Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. Proceedings of the National Academy of Sciences, 102(43), 15545-15550.

Liberzon, A., Subramanian, A., Pinchback, R., Thorvaldsdóttir, H., Tamayo, P., & Mesirov, J. P. (2011). Molecular signatures database (MSigDB) 3.0. Bioinformatics, 27(12), 1739-1740.

Liberzon, A., Birger, C., Thorvaldsdóttir, H., Ghandi, M., Mesirov, J. P., & Tamayo, P. (2015). The molecular signatures database hallmark gene set collection. Cell systems, 1(6), 417-425.

plotGeneStats

Plot gene statistics for clusters of gene sets

## **Description**

This function plots gene statistics against gene frequencies for any given cluster of gene sets. The plot can be used to identify genes that are over-represented in a cluster of gene-sets (identified based on gene-set overlaps) and have a strong statistic (e.g. log fold-chage or p-value).

## Usage

```
plotGeneStats(
   geneStat,
   msigGsc,
   groups,
   statName = "Gene-level statistic",
   topN = 5
)
```

#### **Arguments**

msigGsc

geneStat a named numeric, containing the statistic to be displayed. The vector must be

named with either gene Symbols or Entrez IDs depending on annotations in

a GeneSetCollection object, containing gene sets from the MSigDB. The GSEABase::getBroadSets(

msigGsc.

function can be used to parse XML files downloaded from MSigDB.

groups a named list, of character vectors or numeric indices specifying node groupings.

Each element of the list represent a group and contains a character vector with

node names.

statName a character, specifying the name of the statistic.

topN a numeric, specifying the number of genes to label. The top genes are those with

the largest count and statistic.

## Value

a ggplot object, plotting the gene-level statistic against gene frequencies in the cluster of gene sets.

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#### **Examples**

```
library(GSEABase)

data(hgsc)
groups <- list('g1' = names(hgsc)[1:25], 'g2' = names(hgsc)[26:50])

#create statistics
allgenes = unique(unlist(geneIds(hgsc)))
gstats = rnorm(length(allgenes))
names(gstats) = allgenes

#plot
plotGeneStats(gstats, hgsc, groups)</pre>
```

plotMsigNetwork

Plot a gene set overlap network

#### **Description**

Plots a network of gene set overlap with overlap computed using the computeMsigOverlap() and a graph created using computeMsigNetwork().

#### Usage

```
plotMsigNetwork(
   ig,
   markGroups = NULL,
   genesetStat = NULL,
   nodeSF = 1,
   edgeSF = 1,
   lytFunc = "graphopt",
   lytParams = list(),
   rmUnmarkedGroups = FALSE,
   maxGrp = 12
)
```

## **Arguments**

ig an igraph object, containing a network of gene set overlaps computed using

computeMsigNetwork().

markGroups a named list, of character vectors. Each element of the list represent a group and

contains a character vector with node names. Up to 12 groups can be visualised

in the plot.

genesetStat a named numeric, statistic to project onto the nodes. These could be p-values,

log fold-changes or gene set score from a singscore-based analysis.

nodeSF a numeric, indicating the scaling factor to apply to node sizes.

edgeSF a numeric, indicating the scaling factor to apply to edge widths.

lytFunc a character, specifying the layout to use (see ggraph::create\_layout()).

lytParams a named list, containing additional parameters needed for the layout (see ggraph::create\_layout()

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rmUnmarkedGroups

a logical, indicating whether unmarked groups should be removed from the network (TRUE) or retained (FALSE - default).

maxGrp

a numeric, specifying the maximum number of groups to plot.

## Value

```
a ggplot2 object
```

# **Examples**

```
data(hgsc)
ovlap <- computeMsigOverlap(hgsc, thresh = 0.15)
ig <- computeMsigNetwork(ovlap, hgsc)
groups <- list(
   'g1' = c("HALLMARK_HYPOXIA", "HALLMARK_GLYCOLYSIS"),
   'g2' = c("HALLMARK_INTERFERON_GAMMA_RESPONSE")
)
plotMsigNetwork(ig, markGroups = groups)</pre>
```

plotMsigPPI

Plot PPI network for gene-set clusters identified using vissE

## **Description**

This function plots the protein-protein interaction (PPI) network for a gene-set cluster identified using vissE. The international molecular exchange (IMEx) PPI is used to obtain PPIs for genes present in a gene-set cluster.

## Usage

```
plotMsigPPI(
    ppidf,
    msigGsc,
    groups,
    geneStat = NULL,
    statName = "Gene-level statistic",
    threshConfidence = 0,
    threshFrequency = 0.25,
    threshStatistic = 0,
    threshUseAbsolute = TRUE,
    topN = 5,
    nodeSF = 1,
    edgeSF = 1,
    lytFunc = "graphopt",
    lytParams = list()
)
```

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#### **Arguments**

ppidf a data.frame, containing a protein-protein interaction from the IMEx database.

This can be retrieved from the msigdb::getIMEX() function.

 $msigGsc \hspace{1cm} a \hspace{1cm} Gene Set Collection \hspace{1cm} object, containing \hspace{1cm} gene \hspace{1cm} sets \hspace{1cm} from \hspace{1cm} the \hspace{1cm} MSigDB. \hspace{1cm} The \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase) \hspace{1cm} a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase) \hspace{1cm} a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase) \hspace{1cm} a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase) \hspace{1cm} a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase) \hspace{1cm} a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase) \hspace{1cm} a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase) \hspace{1cm} a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase) \hspace{1cm} a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase) \hspace{1cm} a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase) \hspace{1cm} a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase) \hspace{1cm} a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase) \hspace{1cm} a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase) \hspace{1cm} a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase) \hspace{1cm} a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase) \hspace{1cm} a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase) \hspace{1cm} a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase) \hspace{1cm} a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase) \hspace{1cm} a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} (a \hspace{1cm} GSEABase:: getBroadSets \hspace{1cm} ($ 

function can be used to parse XML files downloaded from MSigDB.

groups a named list, of character vectors or numeric indices specifying node groupings.

Each element of the list represent a group and contains a character vector with

node names.

geneStat a named numeric, containing the statistic to be displayed. The vector must be

named with either gene Symbols or Entrez IDs depending on annotations in

msigGsc.

statName a character, specifying the name of the statistic.

threshConfidence

a numeric, specifying the confidence threshold to apply to determine high confidence interactions. This should be a value between 0 and 1 (default is 0).

threshFrequency

a numeric, specifying the frequency threshold to apply to determine more frequent genes in the gene-set cluster. The frequecy of a gene is computed as the proportion of gene-sets to which the gene belongs. This should be a value be-

tween 0 and 1 (default is 0.25).

threshStatistic

a numeric, specifying the threshold to apply to gene-level statistics (e.g. a log fold-change). This should be a value between 0 and 1 (default is 0).

threshUseAbsolute

a logical, indicating whether the threshStatistic threshold should be applied to absolute values (default TRUE). This can be used to threshold on statistics

such as the log fold-chage from a differential expression analysis.

topN a numeric, specifying the number of genes to label. The top genes are those with

the largest count and statistic.

nodeSF a numeric, indicating the scaling factor to apply to node sizes.

edgeSF a numeric, indicating the scaling factor to apply to edge widths.

lytFunc a character, specifying the layout to use (see ggraph::create\_layout()).

lytParams a named list, containing additional parameters needed for the layout (see ggraph::create\_layout()

#### Value

a ggplot object with the protein-protein interaction networks plot for each gene-set cluster.

```
data(hgsc)
grps = list('early' = 'HALLMARK_ESTROGEN_RESPONSE_EARLY', 'late' = 'HALLMARK_ESTROGEN_RESPONSE_LATE')
ppi = msigdb::getIMEX(org = 'hs', inferred = TRUE)
plotMsigPPI(ppi, hgsc, grps)
```

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plotMsigWordcloud

Compute and plot word frequencies for multiple MSigDB collections

## Description

Given a gene set collection, this function computes the word frequency of gene set names from the Molecular Signatures Database (MSigDB) collection (split by \_). Word frequencies are also computed using short descriptions attached with each gene set object.

# Usage

```
plotMsigWordcloud(
   msigGsc,
   groups,
   weight = NULL,
   measure = c("tfidf", "tf"),
   version = msigdb::getMsigdbVersions(),
   org = c("auto", "hs", "mm"),
   rmwords = getMsigExclusionList(),
   type = c("Name", "Short"),
   idf = NULL
)
```

#### **Arguments**

msigGsc	a GeneSetCollection object, containing gene sets from the MSigDB. The GSEABase::getBroadSets(function can be used to parse XML files downloaded from MSigDB.
groups	a named list, of character vectors or numeric indices specifying node groupings. Each element of the list represent a group and contains a character vector with node names.
weight	a named numeric vector, containing weights to apply to each gene-set. This can be -log10(FDR), -log10(p-value) or an enrichment score (ideally unsigned).
measure	a character, specifying how frequencies should be computed. "tf" uses term frequencies and "tfidf" (default) applies inverse document frequency weights to term frequencies.
version	a character, specifying the version of msigdb to use (see msigdb::getMsigdbVersions()).
org	a character, specifying the organism to use. This can either be "auto" (default), "hs" or "mm".
rmwords	a character vector, containing an exclusion list of words to discard from the analysis.
type	a character, specifying the source of text mining. Either gene set names (Name) or descriptions (Short) can be used.
idf	a list of named numeric vectors, specifying inverse document frequencies to use to penalise terms from gene-set names and short descriptions. This should be a vector of length 2 with names "Name" and "Short". Numeric vectors should

contain weights and names should represent the term. Precomputed versions

can be retrieved using the msigdb::getMsigdbIDF().

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# Value

a ggplot object.

```
data("hgsc")
groups <- list('g1' = names(hgsc)[1:25], 'g2' = names(hgsc)[26:50])
plotMsigWordcloud(hgsc, groups, rmwords = getMsigExclusionList())</pre>
```

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