

# Package ‘Seqtometry’

May 23, 2026

**Title** Signature scoring for single cell analysis

**Version** 1.1.0

**URL** <https://github.com/HawigerLab/Seqtometry>

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## Description

This package provides functions used in Seqtometry (Kousnetsov et al. 2024), a method for analyzing single cell (scRNA-seq or scATAC-seq) data via signature (gene set) enrichment scores. The Seqtometry scores may be useful for annotating or characterizing cells, either in a flow cytometry like workflow (where scores are standalone features used for progressive partitioning as described in the Seqtometry publication) or in a cluster-based workflow (as features of clusters). The exported impute function (a port of Python's MAGIC-impute, van Dijk et al. 2018), may also be useful for single cell analysis on its own.

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Seqtometry-package      *Signature scoring for single cell analysis*

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## Description

This package provides functions used in Seqtometry (Kousnetsov et al. 2024), a method for analyzing single cell (scRNA-seq or scATAC-seq) data via signature (gene set) enrichment scores. The Seqtometry scores may be useful for annotating or characterizing cells, either in a flow cytometry like workflow (where scores are standalone features used for progressive partitioning as demonstrated in the Seqtometry publication) or in a cluster-based workflow (as features of clusters). The exported impute function (a port of Python’s MAGIC-impute, van Dijk et al. 2018), may also be useful for single cell analysis on its own.

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.apply\_diff\_op            *Perform data diffusion*

---

### Description

Apply diffusion operator to data in order to perform imputation.

### Usage

```
.apply_diff_op(gex, pcs, aff, dft, t_max, tol, exact_solver)
```

### Arguments

gex	<b>matrix or Matrix</b> Gene expression matrix
pcs	<b>matrix</b> Principal components matrix
aff	<b>dgCMatrix</b> Markov affinity matrix
dft	<b>NULL or integer(1)</b> Diffusion time
t_max	<b>integer(1)</b> Maximum diffusion time
tol	<b>numeric(1)</b> Tolerance for Procrustes disparity
exact_solver	<b>logical(1)</b> Perform imputation in gene space

### Value

**list(matrix, integer(1))** Imputed matrix and diffusion time used

---

.calc\_diff\_op            *Compute diffusion operator*

---

### Description

Calculate graph diffusion operator (Markov affinity matrix).

### Usage

```
.calc_diff_op(pcs, knn, ka, dist_metric)
```

### Arguments

pcs	<b>matrix</b> Principal components matrix (used for kNN search)
knn	<b>integer(1)</b> Number of nearest neighbors to search for
ka	<b>integer(1)</b> Number of nearest neighbors to use for adaptive kernel
dist_metric	<b>character(1)</b> Type of metric to use for distance calculations during kNN search

### Value

**dgCMatrix** Markov affinity matrix

---

`.calc_pca`*PCA wrapper*

---

**Description**

Calculate leading principal components (via truncated singular value decomposition).

**Usage**

```
.calc_pca(gex, npc, scale)
```

**Arguments**

<code>gex</code>	<b>matrix or Matrix</b> Gene expression matrix (without any zero variance genes)
<code>npc</code>	<b>numeric(1)</b> Number of leading principal components to compute
<code>scale</code>	<b>logical(1)</b> Whether to scale genes to unit variance

**Value**

**list** PC loading/rotation matrices as well as centering/scaling vectors

---

`.check_params`*Parameter validation*

---

**Description**

Checks that all parameters used in `impute` are valid.

**Usage**

```
.check_params(args)
```

**Arguments**

<code>args</code>	<b>list</b> The arguments to the <code>magic_impute</code> function
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**Value**

NULL (but stops execution for invalid parameters)

---

.gene\_indices                      *Finds row indices of signature genes*

---

### Description

For converting from character to integer based indexing

### Usage

```
.gene_indices(mat, gss)
```

### Arguments

mat	<b>matrix-like</b> Gene expression data (genes x cells)
gss	<b>named list of character</b> Signature genes (with same nomenclature system as mat)

### Value

**integer** 0-based indices (for passing to Rcpp function) of signature genes

---

.invert\_pca                      *PCA inversion*

---

### Description

Reverses operations done for PCA: back-rotation, unscaling, and uncentering.

### Usage

```
.invert_pca(pcs, rot, ctr, sdv, low_mem)
```

### Arguments

pcs	<b>matrix</b> The principal components (scaled left singular vectors)
rot	<b>matrix</b> The rotation matrix (right singular vectors)
ctr	<b>integer</b> The centering vector
sdv	<b>integer or NULL</b> The scaling vector (or NULL if no scaling was applied)
low_mem	<b>logical(1)</b> Whether to use delayed operations to reduce memory usage

### Value

**matrix or DelayedMatrix**  $\text{rot} \% \% \text{t}(\text{pcs}) * \text{sdv} + \text{ctr}$

---

<code>.minmax_scale</code>	<i>Minmax transform</i>
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**Description**

Scales input vector to unit range

**Usage**

```
.minmax_scale(x)
```

**Arguments**

`x`                    **numeric** Values to be scaled

**Value**

**numeric** Minmax transformed values

---

<code>.normalize</code>	<i>LogCP10K transform</i>
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---

**Description**

Simple normalization method for scRNA-seq data.

**Usage**

```
.normalize(gex)
```

**Arguments**

`gex`                    **matrix or Matrix** Gene expression matrix (cells x genes)

**Value**

**matrix or Matrix** Transformed (normalized) matrix

---

.procrustes                      *Procrustes disparity*

---

### Description

Calculates symmetric Procrustes distance (adapted from MATLAB procrustes).

### Usage

```
.procrustes(x, y)
```

### Arguments

x	<b>matrix</b>
y	<b>matrix</b>

### Value

**numeric(1)** Procrustes disparity between input matrices

---

impute                              *MAGIC imputation (van Dijk et al. 2018)*

---

### Description

Calculates a graph diffusion operator for the given input matrix and applies it to produce an imputed matrix.

### Usage

```
impute(  
  gex,  
  transpose = TRUE,  
  do_norm = FALSE,  
  pca = NULL,  
  npc = 100L,  
  scale = TRUE,  
  knn = 16L,  
  ka = 6L,  
  dist_metric = "euclidean",  
  dft = NULL,  
  t_max = 16L,  
  tol = 0.001,  
  exact_solver = TRUE,  
  conserve_memory = FALSE,  
  env_ret = FALSE,  
  verbose = FALSE  
)
```

**Arguments**

<code>gex</code>	<b>matrix or Matrix</b> Gene expression values (that has passed quality control).
<code>transpose</code>	<b>logical(1)</b> Whether to transpose <code>gex</code> (make it cells x genes) prior to downstream operations.
<code>do_norm</code>	<b>logical(1)</b> Whether to perform LogCP10K normalization on <code>gex</code> .
<code>pca</code>	<b>matrix (cells x PCs) or NULL</b> Precomputed principal component matrix (or NULL to derive it from <code>gex</code> ).
<code>npc</code>	<b>integer(1)</b> Number of principal components (min = 1) to calculate.
<code>scale</code>	<b>logical(1)</b> Whether to scale columns of input matrix to unit variance prior to PCA.
<code>knn</code>	<b>integer(1)</b> Number of nearest neighbors (min = 2) to consider during distance calculation.
<code>ka</code>	<b>integer(1)</b> Number of nearest neighbors (min = 2, max <= knn) to use for the adaptive kernel.
<code>dist_metric</code>	<b>character(1)</b> Type of metric to use for distance calculations during kNN search.
<code>dft</code>	<b>NULL or integer(1)</b> Automatic (NULL) or user-defined (integer) diffusion time (min = 1, max = 16).
<code>t_max</code>	<b>integer(1)</b> Maximum diffusion time to test when using automatic diffusion time (min = 1, max = 16).
<code>tol</code>	<b>numeric(1)</b> Threshold for Procrustes disparity (min = 0, max = 1) between successive diffusion times.
<code>exact_solver</code>	<b>logical(1)</b> Whether to perform imputation in gene space (TRUE) or PCA space (FALSE).
<code>conserve_memory</code>	<b>logical(1)</b> Whether to avoid allocating a large dense matrix when <code>exact_solver</code> = FALSE.
<code>env_ret</code>	<b>logical(1)</b> Return all variables in the environment (TRUE) or just the imputed matrix (FALSE).
<code>verbose</code>	<b>logical(1)</b> Whether to print messages at different major parts of the algorithm.

**Value**

**matrix-like or list** If `env_ret` = FALSE, then just the imputed matrix. Otherwise the function environment as a list containing all parameters (possibly modified) as well as

- `imp` **matrix or DelayedMatrix** Imputed matrix.
- `aff` **dgCMatrix** Markov affinity matrix (graph diffusion operator).
- `pca` **list** Possibly computed (if `pca` was NULL), yielding a four element list, where:
  - `x` **matrix (cells x PCs)** The principal components matrix (scaled left singular vectors).
  - `v` **matrix (genes x PCs)** The rotation matrix (right singular vectors).
  - `center` **integer (cells)** The centering vector.
  - `scale` **integer (cells) or NULL** The scaling vector (or NULL if no scaling was applied).

## Examples

```

box::use(
  TENxPBMCData[TENxPBMCData],
  SingleCellExperiment[rowData, logcounts],
  scuttle[quickPerCellQC, logNormCounts],
  scater[runUMAP, plotReducedDim],
  patchwork[wrap_plots])

# PBMC data, basic processing pipeline
dat <- TENxPBMCData(dataset = "pbmc3k")
dimnames(dat) <- list(
  rowData(dat)[["Symbol_TENx"]],
  dat[["Barcode"]])
dat <- dat |>
  quickPerCellQC() |>
  logNormCounts() |>
  runUMAP()

# MAGIC imputation
imp <- logcounts(dat) |>
  as("dgCMatrx") |>
  impute()

# Visualize unimputed versus imputed expression
# on UMAP plots for a gene of interest (GOI)
goi <- "CD19"
dat[["Imputed_GOI"]] <- imp[goi, ]
p1 <- plotReducedDim(dat, "UMAP", color_by = goi)
p2 <- plotReducedDim(dat, "UMAP", color_by = "Imputed_GOI")
wrap_plots(p1, p2, ncol = 2)

```

---

score

*Seqtometry scoring (Kousnetsov et al. 2024)*

---

## Description

Computes signature scores (a weighted KS-like statistic) for single cell expression data

## Usage

```
score(mat, signatures, minmax = TRUE)
```

## Arguments

mat	<b>matrix, Matrix, or DelayedMatrix</b> Gene expression data (genes x cells)
signatures	<b>named list of character</b> Signature genes (with same nomenclature system used in mat)
minmax	<b>logical(1)</b> Whether to perform minmax transform on scoring results (default: TRUE)

**Value**

**data.table** Single cell scores (cells x signatures) for each signature, where cell barcodes are stored in the "id" column

**Examples**

```
box::use(
  TENxPBMCData[TENxPBMCData],
  SingleCellExperiment[rowData, logcounts],
  scuttle[quickPerCellQC, logNormCounts],
  scater[runUMAP, plotReducedDim],
  patchwork[wrap_plots])

# PBMC data, basic processing pipeline
dat <- TENxPBMCData(dataset = "pbmc3k")
dimnames(dat) <- list(
  rowData(dat)[["Symbol_TENx"]],
  dat[["Barcode"]])
dat <- dat |>
  quickPerCellQC() |>
  logNormCounts() |>
  runUMAP()

# MAGIC imputation
imp <- logcounts(dat) |>
  as("dgCMatrx") |>
  impute()

# Score with a B cell signature (gene set)
options(future.globals.maxSize = 1024^3)
b_cell_sig <- list("B_cell" = c("CD19", "MS4A1", "CD79A", "CD79B"))
dat[["B cell signature"]] <- Seqtometry::score(imp, b_cell_sig)[["B_cell"]]

# Visualize a hallmark B cell gene versus a B cell signature score
p1 <- plotReducedDim(dat, "UMAP", color_by = "CD19")
p2 <- plotReducedDim(dat, "UMAP", color_by = "B cell signature")
wrap_plots(p1, p2, ncol = 2)
```

wks

*Helper function for performing a weighted Kolmogorov-Smirnov-like procedure.*

**Description**

Helper function for performing a weighted Kolmogorov-Smirnov-like procedure.

**Usage**

```
wks(gex, gss, mus, sds)
```

**Arguments**

gex	numeric: normalized gene expression values
gss	list: indices of genes in each gene set
mus	numeric: means of all genes
sds	numeric: standard deviations of all genes

**Value**

Modified Kuiper statistic (sum of minimal and maximal deviations during running sum)

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