

Package ‘HiCool’

July 26, 2025

Version 1.9.1

Date 2022-11-04

Title HiCool

Description HiCool provides an R interface to process and normalize Hi-C paired-end fastq reads into .(m)cool files.
.(m)cool is a compact, indexed HDF5 file format specifically tailored for efficiently storing HiC-based data.
On top of processing fastq reads, HiCool provides a convenient reporting function to generate shareable reports summarizing Hi-C experiments and including quality controls.

License MIT + file LICENSE

URL <https://github.com/js2264/HiCool>

BugReports <https://github.com/js2264/HiCool/issues>

Depends R (>= 4.2), HiCExperiment

Imports BiocIO, S4Vectors, GenomicRanges, IRanges, InteractionSet, vroom, basilisk.utils, basilisk, reticulate, rmarkdown, rmdformats, plotly, dplyr, stringr, sessioninfo, utils

Suggests HiContacts, HiContactsData, AnnotationHub, BiocFileCache, BiocStyle, testthat, knitr, rmarkdown

biocViews HiC, DNA3DStructure, DataImport

Encoding UTF-8

VignetteBuilder knitr

LazyData false

Roxygen list(markdown = TRUE)

RoxygenNote 7.3.2

Config/testthat/edition 3

StagedInstall no

git_url <https://git.bioconductor.org/packages/HiCool>

git_branch devel

git_last_commit 1422bc6

git_last_commit_date 2025-07-17

Repository Bioconductor 3.22

Date/Publication 2025-07-25
Author Jacques Serizay [aut, cre]
Maintainer Jacques Serizay <jacquesserizay@gmail.com>

Contents

getLoops	2
HiCool	3
HiCReport	5
Index	6

getLoops	<i>Finding loops in contact map</i>
----------	-------------------------------------

Description

Find loops using chromosight

Usage

```
getLoops(  
  x,  
  resolution = NULL,  
  output_prefix = file.path("chromosight", "chromo"),  
  norm = "auto",  
  max.dist = "auto",  
  min.dist = "auto",  
  min.separation = "auto",  
  n.mads = 5L,  
  pearson = "auto",  
  nreads = "no",  
  ncores = 1L  
)
```

Arguments

x	A HiCExperiment object
resolution	Which resolution to use to search loops
output_prefix	Prefix to chromosight output (default: "chromosight/chromo")
norm	Normalization parameter for chromosight
min.dist, max.dist	Min and max distance to use to filter for significant loops
min.separation	Minimum separation between anchors of potential loops
n.mads	Number of MADs to use to filter relevant bins to search for loops
pearson	Minimum Pearson correlation score to use to filter for significant loops
nreads	Number of reads to subsample to before searching for loops
ncores	Number of cores for chromosight

Value

A HiCExperiment object with a new "loops" topologicalFeatures storing significant interactions identified by chromosight, and an additional chromosight_args metadata entry.

Examples

```
contacts_yeast <- contacts_yeast()
contacts_yeast <- getLoops(contacts_yeast)
metadata(contacts_yeast)$chromosight_args
topologicalFeatures(contacts_yeast, 'loops')
```

HiCool

*Processing Hi-C paired-end fastq files in R***Description**

HiCool::HiCool() automatically processes paired-end HiC sequencing files by performing the following steps:

1. Automatically setting up an appropriate conda environment using basilisk;
2. Mapping the reads to the provided genome reference using hicstuff and filtering of irrelevant pairs;
3. Filtering the resulting pairs file to remove unwanted chromosomes (e.g. chrM);
4. Binning the filtered pairs into a cool file at a chosen resolution;
5. Generating a multi-resolution mcool file;
6. Normalizing matrices at each resolution by iterative correction using cooler.

The filtering strategy used by hicstuff is described in Cournac et al., BMC Genomics 2012.

Usage

```
HiCool(
  r1 = "~/repos/tinyMapper/tests/testHiC_R1.fq.gz",
  r2 = "~/repos/tinyMapper/tests/testHiC_R2.fq.gz",
  genome = "R64-1-1",
  restriction = "DpnII,HinfI",
  binning = NULL,
  iterative = TRUE,
  balancing_args = " --min-nnz 10 --mad-max 5 ",
  threads = 1L,
  exclude_chr = "Mito|chrM|MT",
  output = "HiCool",
  keep_bam = FALSE,
  build_report = TRUE,
  scratch = tempdir()
)

importHiCoolFolder(output, hash, resolution = NULL)

getHiCoolArgs(log)

getHicStats(log)
```

Arguments

<code>r1</code>	Path to fastq file (R1 read)
<code>r2</code>	Path to fastq file (R2 read)
<code>genome</code>	Genome used to map the reads on, provided either as a fasta file (in which case the bowtie2 index will be automatically generated), or as a prefix to a bowtie2 index (e.g. <code>mm10</code> for <code>mm10.*.bt2</code> files). Genome can also be a unique ID for the following references: <code>hg38</code> , <code>mm10</code> , <code>dm6</code> , <code>R64-1-1</code> , <code>GRZc10</code> , <code>WBcel235</code> , <code>Galgal4</code> .
<code>restriction</code>	Restriction enzyme(s) used in HiC (Default: <code>"DpnII,HinfI"</code>)
<code>binning</code>	First resolution used to bin the final mcool file (Default: <code>10000</code> for <code>hg38</code> and <code>mm10</code> , <code>1000</code> for <code>dm6</code> , <code>R64-1-1</code> , ...)
<code>iterative</code>	Should the read mapping be performed iteratively? (Default: <code>TRUE</code>)
<code>balancing_args</code>	Balancing arguments for cooler. See cooler documentation here for a list of all available balancing arguments. These defaults match those used by the 4DN consortium.
<code>threads</code>	Number of CPUs used for parallelization. (Default: <code>1</code>)
<code>exclude_chr</code>	Chromosomes excluded from the final .mcool file. This will not affect the pairs file. (Default: <code>"MitochrMIMT"</code>)
<code>output</code>	Output folder used by HiCool.
<code>keep_bam</code>	Should the bam files be kept? (Default: <code>FALSE</code>)
<code>build_report</code>	Should an automated report be computed? (Default: <code>TRUE</code>)
<code>scratch</code>	Path to temporary directory where processing will take place. (Default: <code>tempdir()</code>)
<code>hash</code>	Unique 6-letter ID used to identify files from a specific HiCool processing run.
<code>resolution</code>	Resolution used to import the mcool file
<code>log</code>	Path to log file generated by hicstuff/hicool

Value

A CoolFile object with prefilled pairsFile and metadata slots.

HiCool utils

- `importHiCoolFolder(folder, hash)` automatically finds the different processed files associated with a specific `HiCool::HiCool()` processing hash ID.
- `getHiCoolArgs()` parses the log file generated by `HiCool::HiCool()` during processing to recover which arguments were used.
- `getHicStats()` parses the log file generated by `HiCool::HiCool()` during processing to recover pre-computed stats about pair numbers, filtering thresholds, etc.

Examples

```
r1 <- HiContactsData::HiContactsData(sample = 'yeast_wt', format = 'fastq_R1')
r2 <- HiContactsData::HiContactsData(sample = 'yeast_wt', format = 'fastq_R2')
hcf <- HiCool(r1, r2, genome = 'R64-1-1', output = './HiCool/')
hcf
getHiCoolArgs(metadata(hcf)$log)
getHicStats(metadata(hcf)$log)
readLines(metadata(hcf)$log)
```

HiCReport	<i>HiC processing report</i>
-----------	------------------------------

Description

HiC processing report

Usage

```
HiCReport(x, output = NULL)
```

Arguments

x	an CoolFile object, generated from <code>HiCool::HiCool()</code> or <code>HiCool::importHiCoolFolder()</code> , or directly from calling <code>HiCExperiment::CoolFile()</code> .
output	Path to save output HTML file.

Value

String to the generated HTML report file

Examples

```
mcool_path <- HiContactsData::HiContactsData('yeast_wt', 'mcool')
pairs_path <- HiContactsData::HiContactsData('yeast_wt', 'pairs.gz')
log_path <- HiContactsData::HiContactsData(sample = 'yeast_wt', format = 'HiCool_log')
cf <- CoolFile(mcool_path, pairs = pairs_path, metadata = list(log = log_path))
HiCReport(cf)
```

Index

`getHiCoolArgs (HiCool)`, [3](#)
`getHicStats (HiCool)`, [3](#)
`getLoops`, [2](#)

`HiCool`, [3](#)
`HiCReport`, [5](#)

`importHiCoolFolder (HiCool)`, [3](#)