

Metadata and Annotation with Bioconductor



What is “Metadata” ?

„Data about data“

Helps in understanding the data under study.

**For example,
annotation of genomic regions (exons, transcription start
sites, regulatory regions)**

**annotation of gene products (transcript and protein IDs, gene
ontology annotation, protein classifications, associated
phenotypes)**

The label „meta-“ is in the eye of the beholder.

We will also want to integrate other ‚primary‘ datasets.

Managing Annotation

Static Annotation

- Bioconductor packages containing annotation information that are installed locally on a computer
- standardized and well-documented structure
- support reproducible analyses
- no need for network connection

Dynamic Annotation

- stored in a database accessed via the internet
- more frequent updates → possibly different result when repeating analyses
- more information
- higher diversity of data formats and conventions

BioMart is usually employed as “dynamic”, but could also be made “static” when installed locally on your computer

Metadata - examples

- **EntrezGene**
is a catalog of genetic loci that connects curated sequence information to official nomenclature. It replaced LocusLink.
- **RefSeq**
is a non-redundant set of transcripts and proteins of known genes for many species, including human, mouse and rat.
- **Enzyme Commission (EC)**
numbers are assigned to different enzymes and linked to genes through EntrezGene.

Metadata - examples

- **Gene Ontology (GO)**
is a structured vocabulary of terms describing gene products according to molecular function, biological process, or cellular component
- **PubMed**
is a service of the U.S. National Library of Medicine. PubMed provides a rich resource of data and tools for papers in journals related to medicine and health. While large, it is not comprehensive, not all papers have been abstracted, and only abstracts are searched.
- **Microarray Data Archives**
The NCBI coordinates the Gene Expression Omnibus (GEO); TIGR provides the Resourcerer database, and the EBI runs ArrayExpress.
(GeoQuery and ArrayExpress packages)

Metadata - examples

- **OMIM**

Online Mendelian Inheritance in Man is a catalog of human genes and genetic disorders.

- **KEGG**

Kyoto Encyclopedia of Genes and Genomes; a collection of data resources including a rich collection of pathway data.

- **IntAct**

Protein Interaction data, mainly derived from experiments.

- **Pfam**

Pfam is a large collection of multiple sequence alignments and hidden Markov models covering many common protein domains and families.

Bioconductor Annotation Packages

```
> ls("package:org.Hs.eg.db")  
[1] "org.Hs.eg"                                "org.Hs.egACCNUM"  
[3] "org.Hs.egACCNUM2EG"                      "org.Hs.egALIAS2EG"  
[5] "org.Hs.egCHR"                            "org.Hs.egCHRLLENGTHS"  
[7] "org.Hs.egCHRLLOC"                        "org.Hs.egCHRLOCEND"  
[9] "org.Hs.eg_dbconn"                         "org.Hs.eg_dbfile"  
[11] "org.Hs.eg_dbInfo"                         "org.Hs.eg_dbschema"  
[13] "org.Hs.egENSEMBL"                        "org.Hs.egENSEMBL2EG"  
[15] "org.Hs.egENSEMBLPROT"                     "org.Hs.egENSEMBLPROT2EG"  
[17] "org.Hs.egENSEMBLTRANS"                    "org.Hs.egENSEMBLTRANS2EG"  
[19] "org.Hs.egENZYME"                          "org.Hs.egENZYME2EG"  
[21] "org.Hs.egGENENAME"                        "org.Hs.egGO"  
[23] "org.Hs.egGO2ALLEGS"                       "org.Hs.egGO2EG"  
[25] "org.Hs.egMAP"                            "org.Hs.egMAP2EG"  
[27] "org.Hs.egMAPCOUNTS"                       "org.Hs.egOMIM"  
[29] "org.Hs.egOMIM2EG"                         "org.Hs.egORGANISM"  
[31] "org.Hs.egPATH"                           "org.Hs.egPATH2EG"  
[33] "org.Hs.egPFAM"                           "org.Hs.egPMID"  
[35] "org.Hs.egPMID2EG"                         "org.Hs.egPROSITE"  
[37] "org.Hs.egREFSEQ"                          "org.Hs.egREFSEQ2EG"  
[39] "org.Hs.egSYMBOL"                          "org.Hs.egSYMBOL2EG"  
[41] "org.Hs.egUNIGENE"                         "org.Hs.egUNIGENE2EG"  
[43] "org.Hs.egUNIPROT"
```

Each of these objects represents a (SQL-like) database table

```
> head(toTable(org.Hs.egOMIM))  
gene_id omim_id  
1      1 138670  
2      2 103950  
3      2 104300  
4      9 108345  
5     10 243400  
6     10 612182
```

gene_id is the Entrez Gene ID and is used as a key to link the information together

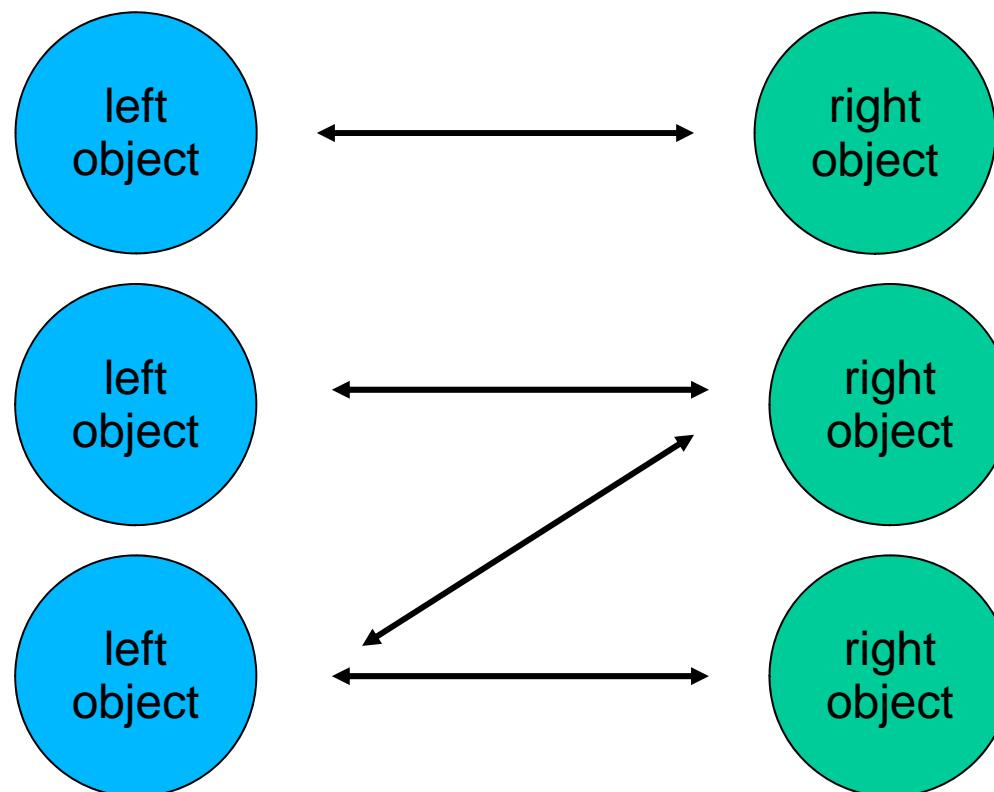
```
> head(toTable(org.Hs.egPFAM))  
gene_id      ipi_id  PfamId  
1    IPI00022895  PF00047  
2    IPI00644018  PF00047  
3    IPI00646799  PF00047  
4    IPI00478003  PF00207  
5    IPI00478003  PF01835  
6    IPI00478003  PF07677
```

Direct SQL querying of these tables is possible, but there are also some R functions for efficient querying and joining.

Bimap objects

Data in the annotation packages are stored in an SQLite database

Conceptually, the tables are Bimaps (bi-directional maps), i.e. n:m mappings between two sets („left“ and „right“ set)



Lkeys, Rkeys: Get left and right keys of a Bimap object

```
> head(Lkeys(org.Hs.egOMIM))  
[1] "1" "10" "100"  
[3] "1000" "10000" "100008586"
```

gene_id



```
> head(Rkeys(org.Hs.egOMIM))  
[1] "100300" "100640" "100650"  
[3] "100660" "100670" "100678"
```

omim_id



Querying across multiple tables

```
> # Find all PfamIDs associated with OMIM code `125853`  
> # DIABETES MELLITUS, NONINSULIN-DEPENDENT; NIDDM  
  
> t1 = toTable(revmap(org.Hs.egOMIM)[ "125853" ] )  
  
> t2 = toTable(org.Hs.egPFAM[t1$gene_id])  
  
> tt = merge(t1, t2)  
  
> head(tt)  
  gene_id omim_id      ipi_id PfamId  
1  10644  125853 IPI00179713 PF00076  
2  10644  125853 IPI00179713 PF00013  
3  10644  125853 IPI00180983 PF00076  
4  10644  125853 IPI00922167 PF00013  
5  10644  125853 IPI00790840      <NA>  
6  10644  125853 IPI00180983 PF00013
```

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http://www.biomart.org/ biomart

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BioMart Project

BioMart is a query-oriented data management system developed jointly by the [Ontario Institute for Cancer Research \(OICR\)](#) and the [European Bioinformatics Institute \(EBI\)](#).

The system can be used with any type of data and is particularly suited for providing 'data mining' like searches of complex descriptive data. BioMart comes with an 'out of the box' website that can be installed, configured and customised according to user requirements. Further access is provided by graphical and text based applications or programmatically using web services or API written in Perl and Java. BioMart has built-in support for query optimisation and data federation and in addition can be configured to work as a DAS 1.5 Annotation server. The process of converting a data source into BioMart format is fully automated by the tools included in the package. Currently supported RDBMS platforms are MySQL, Oracle and Postgres.

BioMart is completely Open Source, licensed under the LGPL, and freely available to anyone without restrictions.

Powered by BioMart software:

• BioMart Central Portal	• Dictybase	• Rat Genome Database	• Pancreatic Expression Database
• Ensembl	• Wormbase	• DroSpeGe	• Reactome
• Ensembl Bacteria	• Gramene	• ArrayExpress	• EU Rat Mart
• Ensembl Metazoa	• Europhenome	• DW	• Paramecium DB
• Ensembl Protists	• UniProt	• Eurexpress	• HGNC
		• HapMap	• VectorBase
			• HTGT

Third party software with BioMart Plugin:

[Bioclipse](#) [biomaRt-BioConductor](#) [Cytoscape](#) [Galaxy](#) [Taverna](#) [WebLab](#)

Completato



Joint project between EMBL-EBI and the Sanger Institute

Produces and maintains genome databases for vertebrates and other eukaryotic species.

<http://www.ensembl.org>

Help & Documentation

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Find a Species

Ensembl Species

Alpaca <i>Vicugna pacos</i>	Fugu <i>Takifugu rubripes</i>	Orangutan <i>Pongo pygmaeus</i>
Anole Lizard <i>Anolis carolinensis</i>	Gorilla <i>Gorilla gorilla</i>	Pig (preview - assembly only) <i>Sus scrofa</i>
Anopheles <i>Anopheles gambiae</i>	Guinea Pig <i>Cavia porcellus</i>	Pika <i>Ochotona princeps</i>
Armadillo <i>Dasypus novemcinctus</i>	Hedgehog <i>Erinaceus europaeus</i>	Platypus <i>Ornithorhynchus anatinus</i>
Bushbaby <i>Otolemur garnettii</i>	Horse <i>Equus caballus</i>	Rabbit <i>Oryctolagus cuniculus</i>
Caenorhabditis elegans	Human <i>Homo sapiens</i>	Rat <i>Rattus norvegicus</i>
Ciona intestinalis	Hyrax <i>Procavia capensis</i>	Saccharomyces cerevisiae
Ciona savignyi	Kangaroo rat <i>Dipodomys ordii</i>	Shrew <i>Sorex araneus</i>
Cat <i>Felis catus</i>	Lamprey (preview - assembly only) <i>Petromyzon marinus</i>	Sloth <i>Choloepus hoffmanni</i>
Chicken <i>Gallus gallus</i>	Lesser hedgehog tenrec <i>Echinops telfairi</i>	Squirrel <i>Spermophilus tridecemlineatus</i>
Chimpanzee <i>Pan troglodytes</i>	Macaque <i>Macaca mulatta</i>	Stickleback <i>Gasterosteus aculeatus</i>
Cow <i>Bos taurus</i>	Medaka <i>Oryzias latipes</i>	Tarsier <i>Tarsius syrichta</i>
Dog <i>Canis familiaris</i>	Megabat <i>Pteropus vampyrus</i>	Tetraodon <i>Tetraodon nigroviridis</i>
Dolphin <i>Tursiops truncatus</i>	Microbat <i>Myotis lucifugus</i>	Tree Shrew <i>Tupaia belangeri</i>
Elephant	Mouse	Xenopus tropicalis

Ensembl martview

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http://www.ensembl.org/biomart/martview/418c7629f86deb18ca08b99a1555c57d/418c7629f86deb18ca08b99a1555c57d ensembl

Google BioMart http://www.en...b99a1555c57d

e!Ensembl Home

New Count Results

Dataset 10506 / 37435 Genes
Homo sapiens genes (NCBI36)

Filters GNF anatomical system data : liver

Attributes Ensembl Gene ID, Ensembl Transcript ID, Associated Gene Name, Chromosome Name, Gene Start (bp), Gene End (bp), Strand

Dataset [None Selected]

Export all results to File TSV Unique results only Go

Email notification to

View 20 rows as TSV Unique results only

Ensembl Gene ID	Ensembl Transcript ID	Associated Gene Name	Chromosome Name	Gene Start (bp)	Gene End (bp)	Strand
ENSG00000198308	ENST00000356319	AL773572.1	21	41473220	41480139	-1
ENSG00000185871	ENST00000340131	AL928970.15-2	9	5841	9833	-1
ENSG00000198567	ENST00000360625	AP000552.1-1	22	19975342	19978982	1
ENSG00000173394	ENST00000312288	AC010129.3	Y	5265788	5266981	-1
ENSG00000215467	ENST00000358609	AL121886.22	20	41714577	41715348	1
ENSG00000131982	ENST00000254302	UBE2L7P_14	54765688	54774632		1
ENSG00000166492	ENST00000321602	AC123788.7-2	11	3386461	3400303	-1
ENSG00000214975	ENST00000244636	AL160400.20-1	6	25084626	25085188	1
ENSG00000215168	ENST00000311910	AL139276.17	X	65192154	65193155	-1
ENSG00000215089	ENST00000341395	AL121869.19-1	X	91601274	91602481	1
ENSG00000214869	ENST00000340195	AC004079.1-1	7	27029882	27031053	1
ENSG00000146556	ENST00000326632	AL627309.15-1	1	4274	19669	-1
ENSG00000174977	ENST00000308863	AC026271.6-1	17	18494394	18496737	1
ENSG00000189212	ENST00000359917	AC005400.1	7	35087426	35192299	-1
ENSG00000080947	ENST00000263511	CROCL2_1	16666518	16691691		-1
ENSG00000186301	ENST00000334429	AL137798.8-1	1	16843841	16849428	1
ENSG00000149345	ENST00000278654	AL121755.23	20	5221034	5221477	-1
ENSG00000170827	ENST00000310120	AL162417.23-1	9	134947760	134950256	1
ENSG00000179408	ENST00000319751	AP000689.1-1	21	36424539	36426078	1
ENSG00000205485	ENST00000348899	AC004980.5-1	7	76016614	76018755	1

VEGA

The Vertebrate Genome Annotation (VEGA) database is a central repository for high quality, frequently updated, manual annotation of vertebrate finished genome sequence.



Current release:

- Human
- Mouse
- Zebrafish
- Dog

<http://vega.sanger.ac.uk>

WormBase



WormBase is the repository of mapping, sequencing and phenotypic information for *C. elegans* (and some other nematodes).

<http://www.wormbase.org>

WormMart

BioMart (MartView) - Mozilla Firefox

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http://www.wormbase.org/BioMart/martview

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Home Genome Blast / Blat WormMart Batch Sequences Markers Genetic Maps Subm...

Find: Any Gene

WormBase The Biology and Genome of C. elegans

Select the dataset for this query

Schema:

Database:

Dataset:

Using MartView

After choosing a DATASET above, select some FILTERS on the next page and then which data you want to EXPORT from the OUTPUT page. At any stage the COUNT button will calculate the number of entries you can expect in the final output.

MartView can generate a number of different types of output, including sequence and tabulated list data. Multiple output formats, including HTML, text and Microsoft Excel, are also supported.

worm: mart
refresh Help Desk

Summary

- ▶ start
 - Not yet initialised
- ▶ filter
 - Not yet initialised
- ▶ output
 - Not yet initialised

GrameneMart



Gramene: A Comparative Mapping Resource for Grains

Gramene is a curated, open-source, Web-accessible data resource for comparative genome analysis in the grasses.

<http://www.gramene.org>

Gramene BioMart Genome Browser (MartView) - Mozilla Firefox

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http://www.gramene.org/Multi/martview gramene

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A Comparative Mapping Resource

GRAMENE e!

Search for: Database: All Search Feedback

Genome Browser BLAST CMap Markers Protein Ontology Gene QTL Literature Species Resources About Gramene Site Map

Select the dataset for this query

Dataset:

Using MartView

After choosing a dataset, click the START button to begin your query. You will be directed to the FILTER page where you can specify which data to include in the final output. Then, click the OUTPUT button to see the results. Finally, click the EXPORT button to download your data in various formats.

MartView can generate a number of different types of output, including sequence and tabulated list data. Multiple output formats, including HTML, text and Microsoft Excel, are also supported.

bioMart
refresh Help Desk

Summary

- ▶ start
Not yet initialised
- ▶ filter
Not yet initialised
- ▶ output
Not yet initialised

Other databases with BioMart interfaces

- dbSNP (via Ensembl)
- HapMap
- Sequence Mart: Ensembl genome sequences



BioMart user interfaces

MartShell

MartShell is a command line BioMart user interface based on a structured query language: Mart Query Language (MQL)

```
arek@localhost:~$ ./martshell.sh
Starting Interactive MartShell

MartShell: An Interactive User Interface to BioMart databases based on Mart Query Language (MQL)
type 'help' for a list of available commands, or type 'help command' to get help for a particular command.

MartShell> list marts;

ArrayExpress
Ensembl_28
MSD_3
SNP_28
UniProt_13
Vega_28

MartShell> use ArrayExpress.AE1;
MartShell> get experiment_accession, experiment_type ;
E-MEXP-2      compound_treatment_design,time_series_design
E-MEXP-1      time_series_design,compound_treatment_design
E-TOXM-1      compound treatment design,dose response design
E-MEXP-32     disease_state_design
E-MEXP-88     cellular_modification_design
E-MEXP-25     disease_state_design
MartShell>
```

BioMart user interfaces

Martview Web based user interface for BioMart, provides functionality for remote users to query all databases hosted by the EBI's public BioMart server.

MartExplorer

Perl and Java **libraries**

biomaRt interface to R/Bioconductor

The biomaRt package

Developed by Steffen Durinck (started Feb 2005)

Two main sets of functions:

- 1. Tailored towards Ensembl, shortcuts for FAQs (frequently asked queries): `getGene`, `getGO`, `getOMIM...`**
- 2. Generic queries, modeled after MQL (Mart query language), can be used with any BioMart dataset, usually require some pre- and postprocessing in R.**

Getting started

```
> library(biomart)
> listMarts()

$biomart
[1] "dicty"    "ensembl"   "snp"      "vega"      "uniprot"   "msd"      "wormbase"

$version
[1] "DICTYBASE (NORTHWESTERN)"  "ENSEMBL 38 (SANGER)"
[3] "SNP 38 (SANGER)"           "VEGA 38 (SANGER)"
[5] "UNIPROT 4-5 (EBI)"         "MSD 4 (EBI)"
[7] "WORMBASE CURRENT (CSHL)"

$host
[1] "www.dictybase.org" "www.biomart.org"     "www.biomart.org"
[4] "www.biomart.org"   "www.biomart.org"     "www.biomart.org"
[7] "www.biomart.org"

$path
[1] ""                      "/biomart/martservice" "/biomart/martservice"
[4] "/biomart/martservice"  "/biomart/martservice" "/biomart/martservice"
[7] "/biomart/martservice"
```

Gene annotation

The function `getGene` allows you to get gene annotation for many types of identifiers

Supported identifiers are:

- **RefSeq**
- **Entrez-Gene**
- **EMBL**
- **HUGO**
- **Ensembl**
- **Affymetrix Genechip Probeset ID**

getGene

```
> mart <- useMart("ensembl", dataset = "hsapiens_gene_ensembl")
> myProbes <- c("210708_x_at", "202763_at", "211464_x_at")
> z <- getGene(id = myProbes, array = "affy_hg_u133_plus_2", mart = mart)
```

	ID	symbol
1	202763_at	CASP3
2	210708_x_at	CASP10
7	211464_x_at	CASP6

description

```
1 Caspase-3 precursor (EC 3.4.22.-) (CASP-3) (Apopain) ...
2 Caspase-10 precursor (EC 3.4.22.-) (CASP-10) (ICE-like apoptotic pro...
7 Caspase-6 precursor (EC 3.4.22.-) (CASP-6) (Apoptotic protease Mch-2)...
```

	chromosome	band	strand	chromosome_start	chromosome_end	ensembl_gene_id
1	4	q35.1	-1	185785845	185807623	ENSG00000164305
2	2	q33.1	1	201756100	201802372	ENSG00000003400
7	4	q25	-1	110829234	110844078	ENSG00000138794

	ensembl_transcript_id
1	ENST00000308394
2	ENST00000272879
7	ENST00000265164

getGene

returns a dataframe

- **Gene symbol**
- **Description**
- **Chromosome name**
- **Band**
- **Start position**
- **End position**
- **BioMartID**

Other functions

- **getGO: GO id, GO term, evidence code**
- **getOMIM (Online Mendelian Inheritance in Man, a catalogue of human genes and genetic disorders): OMIM id, Disease, BioMart id**
- **getINTERPRO (an integrated resource of protein families, domains and functional sites): Interpro id, description**
- **getSequence**
- **getSNP**
- **getHomolog**

getSequence

```
> seq <- getSequence(species="hsapiens", chromosome = 19, start =  
18357968, end = 18360987, mart = mart)
```

chromosome

```
[1] "19"
```

start

```
[1] 18357968
```

end

```
[1] 18360987
```

sequence

```
"AGTCCCAGCTCAGAGCCGAAACCTGCACAGCCATGCCCGGGCAAGAACTCAGGACGGTGAATGGCTCTCAG  
ATGCTCCTGGTGTGCTGGTGCCTCGTGGCTGCCGCATGGGGCGCCCTGTCTCTGGCCGAGGCGAGCCGC  
GCAAGTTCCCGGGACCCTCAGAGTTGCACTCCGAAGACTCCAGATTCCGAGAGTTGCGGAAACGCTACGAG  
GACCTGCTAACCAAGGCTGCCGGCCAACCAGAGCTGGGAAGATTGAACACCGACCTCGTCCCAGGCCCCCTGCA  
GTCCGGATACTCACGCCAGAAGGTAAGTGAATCTTAGAGATCCCCTCCCACCCCCCAAGCAGCCCCCATAT  
CTAATCAGGGATTCCCTCATCTTGAAAAGCCAGACCTACCTCGTATCTCTGGGCCGCCCTCCCGAGGGG  
CTCCCCGAGGCCTCCGCCTCACCGGGCTCTGTTCCGGCTGTCCCCGACGGCGTCAAGGTCGTGGACGTG  
ACACGACCGCTGCCGGCGTCAGCTCAGCCTTGCAAGACCCCAGGCGCCCGCTGCACCTGCGACTGTCGCG  
CCGCCGTCGAGTCGGACCAAATGCTGGCAGAATCTCGTCCGCACGGCCCCAGCTGGAGTTGCACCTGCG  
CCGCAAGCCGCCAGGGGGCGCCGCAGAGCGCGTGCAGCAACGGGGACCACTGTCCGCTCGGGCCCGT  
TGCTGCCGTCTGCACACGGTCCCGCGCTGGAAGACCTGGGCTGGGCCGATTGGGTGCTGTCGCCACGG  
GAGGTGCAAGTGACCATGTGCATCGGCGCGTGCAGCCAGTTCCGGGGCAACATG....
```

SNPs

- Single Nucleotide Polymorphisms (SNPs) are one type of common DNA sequence variations between individuals.

e.g.

AAGGCTAA
ATGGCTAA

- biomaRt uses the SNP mart of Ensembl which is obtained from dbSNP

getSNP

```
> getSNP(chromosome = 8, start = 148350, end = 148612, mart = mart)
```

	tsc	refsnnp_id	allele	chrom_start	chrom_strand
1	TSC1723456	rs3969741	C/A	148394	1
2	TSC1421398	rs4046274	C/A	148394	1
3	TSC1421399	rs4046275	A/G	148411	1
4		rs13291	C/T	148462	1
5	TSC1421400	rs4046276	C/T	148462	1
6		rs4483971	C/T	148462	1
7		rs17355217	C/T	148462	1
8		rs12019378	T/G	148471	1
9	TSC1421401	rs4046277	G/A	148499	1
10		rs11136408	G/A	148525	1
11	TSC1421402	rs4046278	G/A	148533	1
12		rs17419210	C/T	148533	-1
13		rs28735600	G/A	148533	1
14	TSC1737607	rs3965587	C/T	148535	1
15		rs4378731	G/A	148601	1

Homology mapping

The `getHomolog` function enables mapping of many types of identifiers from one species to the same or another type of identifier in another species.

getHomolog

```
> from.mart = useMart("ensembl", dataset = "hsapiens_gene_ensembl")  
  
> to.mart = useMart("ensembl", dataset = "mmusculus_gene_ensembl")  
  
> getHomolog(id = 2, from.type = "entrezgene", to.type = "refseq",  
+           from.mart = from.mart, to.mart = to.mart)
```

	v1	v2	v3
1	ENSMUSG00000030111	ENSMUST00000032203	NM_175628
2	ENSMUSG00000059908	ENSMUST00000032228	NM_008645
3	ENSMUSG00000030131	ENSMUST00000081777	NM_008646
4	ENSMUSG00000071204	ENSMUST00000078431	NM_001013775
5	ENSMUSG00000030113	ENSMUST00000032206	
6	ENSMUSG00000030359	ENSMUST00000032510	NM_007376

getFeature

Select all RefSeq id's involved in diabetes mellitus:

```
>getFeature( OMIM="diabetes mellitus",
  type="refseq",
  species="hsapiens",
  mart=mart)
```

The generic interface: the getBM function

Selecting a dataset: the useDataset function

```
> library(biomaRt)

> mart <- useMart("ensembl")

> listDatasets(mart)

      dataset      version
1 rnorvegicus_gene_ensembl    RGSC3.4
2 scerevisiae_gene_ensembl     SGD1
3 celegans_gene_ensembl      CEL150
4 cintestinalis_gene_ensembl    JGI2
5 ptroglodytes_gene_ensembl   CHIMP1A
6 frubripes_gene_ensembl      FUGU4
7 agambiae_gene_ensembl       AgamP3
8 hsapiens_gene_ensembl       NCBI36
9 ggallus_gene_ensembl        WASHUC1
10 xtropicalis_gene_ensembl   JGI4.1
11 drerio_gene_ensembl        ZFISH5
....(more)...

> mart <- useDataset(dataset = "hsapiens_gene_ensembl", mart = mart)
```

getBM

```
> getBM(attributes = c("affy_hg_u95av2", "hgnc_symbol"),
  filter = "affy_hg_u95av2",
  values = c("1939_at", "1000_at"),
  mart = mart)
```

affy_hg_u95av2 hgnc_symbol

```
1    1000_at      MAPK3
3    1939_at      TP53
```

mart - an object describing the database connection and the dataset

attributes - the name of the data you want to obtain

filter - the name of the data by which you want to filter from the dataset

values - values to filter on

Locally installed BioMarts

- Main use case currently is to use biomaRt to query public BioMart servers over the internet
- But you can also install BioMart server locally, populated with a copy of a public dataset (particular version), or populated with your own data
- Versioning is supported by naming convention

Read more

**Mapping Identifiers for the
Integration of Genomic
Datasets with the
R/Bioconductor package
biomaRt. S. Durinck, E.
Birney, P. Spellmann, W.
Huber, Nature Protocols
2009 (to appear)**

