

# Online queries to BioMart web services through biomaRt

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# This workshop

- What is BioMart?
- Example BioMart databases
- Overview of the biomaRt package
  - Simple biomaRt functions
  - Generic biomaRt functions

Hands - on

# BioMart



- Generic data management system aimed at complex interlinked datasets
- Collaboration between EBI and CSHL
- Originally developed for the Ensembl project but has now been generalized

<http://www.biomart.org>



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# Examples of BioMart databases

# Ensembl

- Joint project between EMBL - EBI and the Sanger Institute
- Produces and maintains automatic annotation on selected eukaryotic genomes
- <http://www.ensembl.org>

- Run a BLAST search
- Search Ensembl
- Data mining [BioMart]
- Upload your own data
- Download data

## Docs and downloads

- Information
- What's New
- About Ensembl
- Ensembl data
- Software

### Mammals

#### *Homo sapiens*

[NCBI 35]  
[browse](#) | [what's new](#) |  
[Vega](#)

#### *Pan troglodytes*

[CHIMP1]  
[browse](#) | [what's new](#)

#### *Mus musculus*

[NCBI m34]  
[browse](#) | [what's new](#) |  
[Vega](#)

#### *Rattus norvegicus*

[RGSC 3.4]  
[browse](#) | [what's new](#)

#### *Canis familiaris*

[CanFam1.0]  
[browse](#) | [what's new](#) |  
[Vega](#)

#### *Bos taurus* [Btau 1.0] - **NEW!**

[browse](#) | [what's new](#)

### Other chordates

#### *Gallus gallus*

[WASHUC1]  
[browse](#) | [what's new](#)

#### *Xenopus tropicalis*

[JGI 3]  
[browse](#) | [what's new](#)

#### *Danio rerio* [WTSI Zv5]

[browse](#) | [what's new](#) |  
[Vega](#)

#### *Takifugu rubripes*

[Fugu 2.0]  
[browse](#) | [what's new](#)

#### *Tetraodon nigroviridis*

[TETRAODON 7]  
[browse](#) | [what's new](#)

#### *Ciona intestinalis*

[JGI 1.95]  
[browse](#) | [what's new](#)

### Other eukaryotes

#### *Drosophila melanogaster*

[BGDP 4]  
[browse](#) | [what's new](#)

#### *Anopheles gambiae*

[MOZ 2]  
[browse](#) | [what's new](#)

#### *Apis mellifera*

[Amel 2.0]  
[browse](#) | [what's new](#)

#### *Caenorhabditis elegans* [WS140]

[browse](#) | [what's new](#)

#### *Saccharomyces cerevisiae* [SGD]

[browse](#) | [what's new](#)



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# Ensembl MartView

Ensembl EnsMart Genome Browser (MartView) - Mozilla Firefox

File Edit View Go Bookmarks Tools Help

http://www.ensembl.org/Multi/martview

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e! project Ensembl MartView

welcome trust sanger institute EBI

Home ▶ EnsMart ▶ TextSearch ▶ BlastSearch ▶ MartSearch ▶ Download ▶

Select the dataset for this query

Database: Ensembl 31

Dataset: Homo sapiens genes (NCBI35)

Using MartView

After choosing a DATASET above, select some FILTERS on the next page and then which data you want to EXPORT from the OUTPUT page. At any stage the COUNT button will calculate the number of entries you can expect in the final output.

MartView can generate a number of different types of output, including sequence and tabulated list data. Multiple output formats, including HTML, text and Microsoft Excel, are also supported.

bioMart

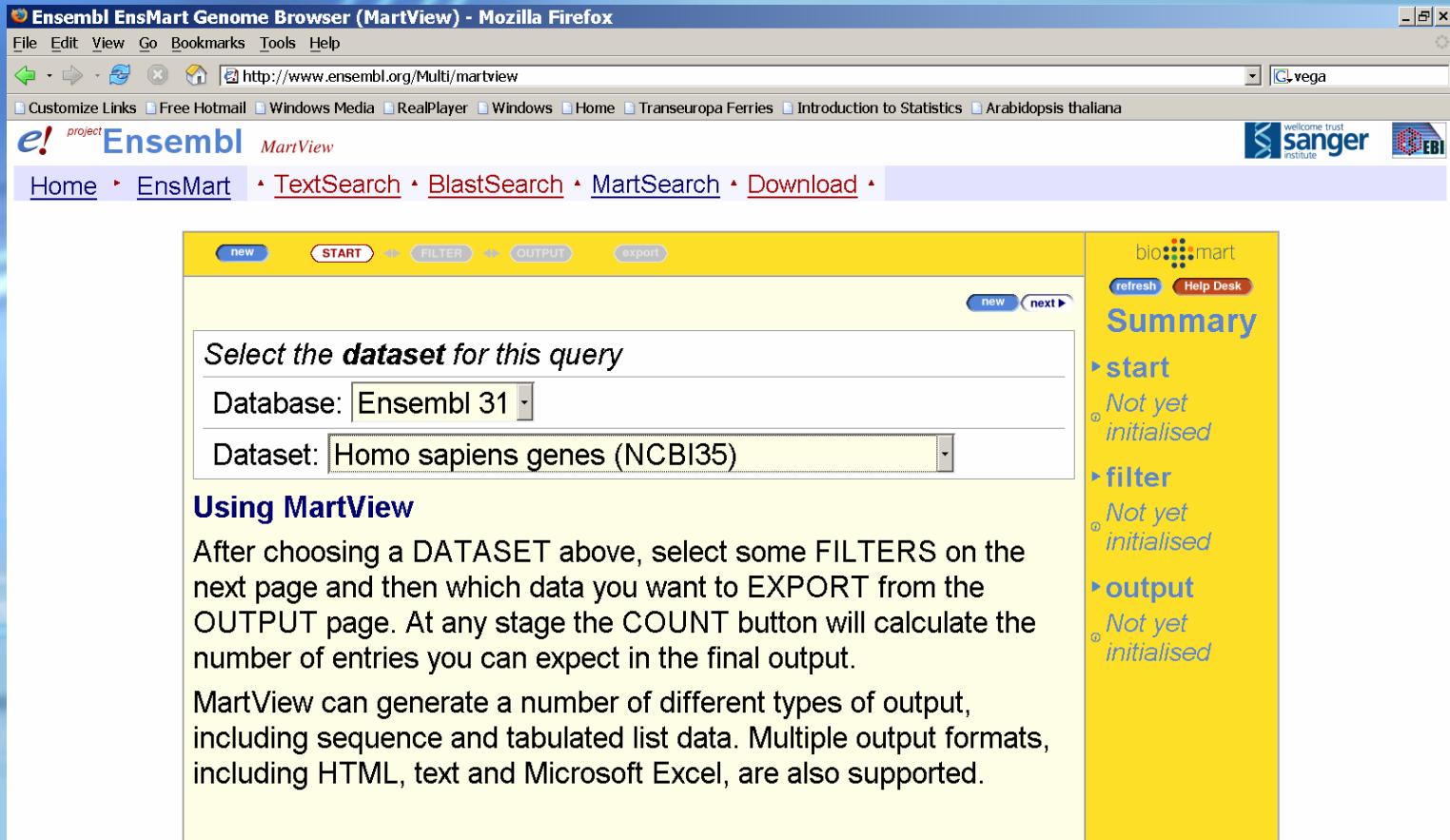
refresh Help Desk

Summary

▶ start  
Not yet initialised

▶ filter  
Not yet initialised

▶ output  
Not yet initialised



# Wormbase

- Repository of mapping, sequencing and phenotypic information on *C. elegans* (and some other nematodes)
- <http://www.wormbase.org>



# WormMart

**BioMart (MartView) - Mozilla Firefox**  
File Edit View Go Bookmarks Tools Help  
<http://www.wormbase.org/BioMart/martview>

Customize Links Free Hotmail Windows Media RealPlayer Windows Home Transeuropa Ferries Introduction to Statistics Arabidopsis thaliana

**Home Genome Blast / Blat WormMart Batch Sequences Markers Genetic Maps Subm**

Find:  Any Gene

**WormBase** The Biology and Genome of C. elegans

**Select the dataset for this query**

Schema:

Database:

Dataset:

**Using MartView**

After choosing a DATASET above, select some FILTERS on the next page and then which data you want to EXPORT from the OUTPUT page. At any stage the COUNT button will calculate the number of entries you can expect in the final output.

MartView can generate a number of different types of output, including sequence and tabulated list data. Multiple output formats, including HTML, text and Microsoft Excel, are also supported.

wormmart  
refresh Help Desk

**Summary**

- ▶ start  
Not yet initialised
- ▶ filter  
Not yet initialised
- ▶ output  
Not yet initialised

# HapMap

- The HapMap Project is an international effort to identify and catalog genetic variation in human.

# HapMap MartView

BioMart (MartView) - Mozilla Firefox

File Edit View Go Bookmarks Tools Help

http://hapmart.hapmap.org/BioMart/martview

Getting Started Latest Headlines

International HapMap Project

Home | About the Project | Data | Publications

**International HapMap Project**

bioMart

**Summary**

- start  
(i) Not yet initialised
- filter  
(i) Not yet initialised
- output  
(i) Not yet initialised

**MartView**

Select the **dataset** for this query

Schema: rel20\_NCBI\_Build35

Database: HapMap\_rel20

Dataset: HapMap Population- Han Chinese in Beijing,China

**Using MartView**

After choosing a DATASET above, select some FILTERS on the next page and then which data you want to EXPORT from the OUTPUT page. At any stage the COUNT button will calculate the number of entries you can expect in the final output.

MartView can generate a number of different types of output, including sequence and tabulated list data. Multiple output formats, including HTML, text and Microsoft Excel, are also supported.

Help Desk / Suggestions

# Gramene

- A comparative mapping resource for grains
- Includes:
  - *Arabidopsis thaliana*
- <http://www.gramene.org>



# GrameneMart

Gramene BioMart Genome Browser (MartView) - Mozilla Firefox

File Edit View Go Bookmarks Tools Help

http://www.gramene.org/Multi/martview gramene

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A Comparative Mapping Resource

GRAMENE e!

Search for: Database: All Search Feedback

Genome Browser BLAST CMap Markers Protein Ontology Gene QTL Literature Species Resources About Gramene Site Map

new START FILTER OUTPUT export

Select the dataset for this query

Dataset:  Zea mays genes (FGENESH01)

Using MartView

After choosing a dataset, click the START button. This will generate a summary page showing the number of entries for each dataset. From there, you can choose which datasets to include in your final output. You can also choose which output format you want to use.

MartView can generate a number of different types of output, including sequence and tabulated list data. Multiple output formats, including HTML, text and Microsoft Excel, are also supported.

bioMart

refresh Help Desk

**Summary**

- ▶ start  
Not yet initialised
- ▶ filter  
Not yet initialised
- ▶ output  
Not yet initialised

# Other publicly available databases with BioMart

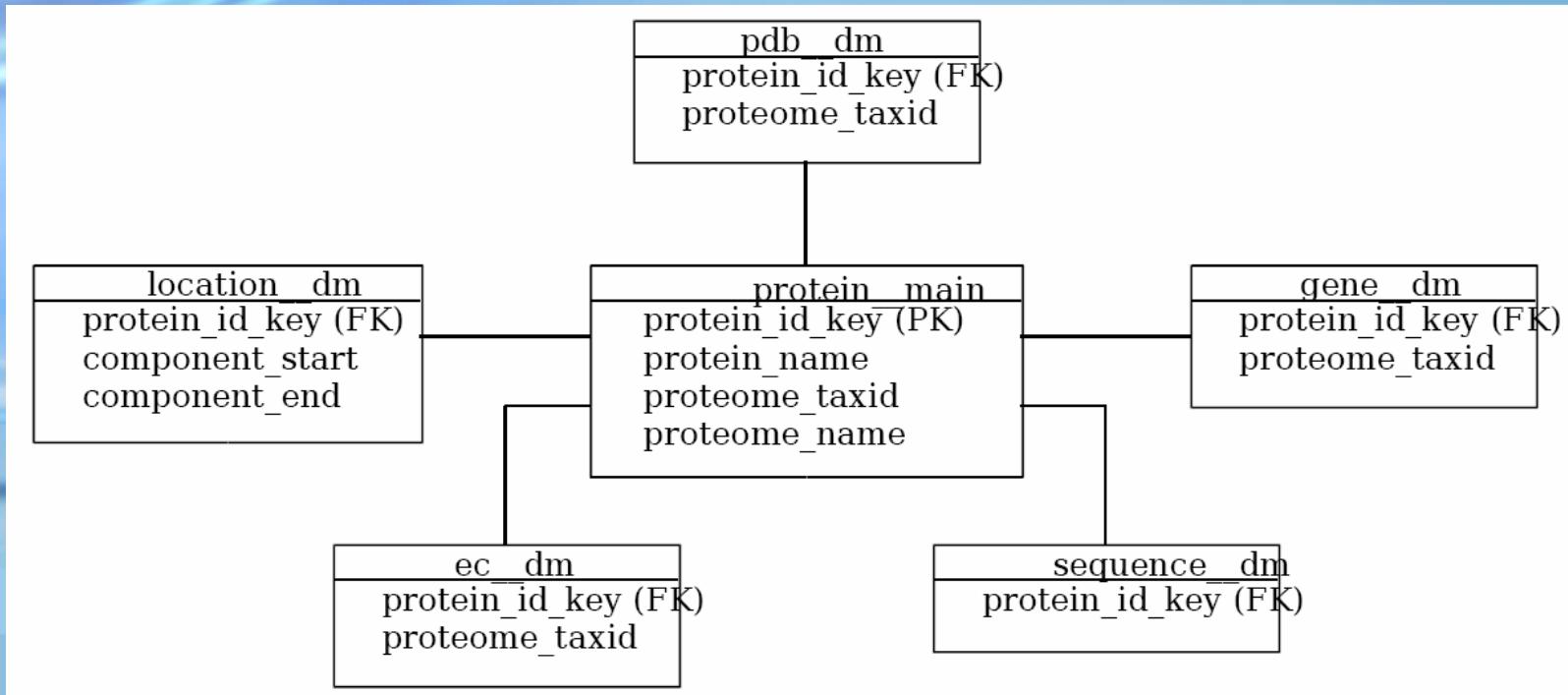
- euGenes
- VEGA
- Dictybase
- ZF-Models
- Uniprot
- MSD

# BioMart databases

- De-normalized
- Tables with 'redundant' information
- Query optimized
- Fast and flexible

# BioMart database schema's

Simple star-like schema's avoid complex joins and enable fast data retrieval



# BioMart user interfaces: MartShell

- Command-line BioMart user interface based on a structured query language: Mart Query Language (MQL)



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# BioMart user interfaces: MartShell

```
arek@localhost:~$ ./martshell.sh
Starting Interactive MartShell

MartShell: An Interactive User Interface to BioMart databases based on Mart Query Language (MQL)
type 'help' for a list of available commands, or type 'help command' to get help for a particular command.

MartShell> list marts;

ArrayExpress
Ensembl_28
MSD_3
SNP_28
UniProt_13
Vega_28

MartShell> use ArrayExpress.AE1;
MartShell> get experiment_accession, experiment_type ;
E-MEXP-2      compound_treatment_design,time_series_design
E-MEXP-1      time_series_design,compound_treatment_design
E-TOXM-1      compound treatment design,dose response design
E-MEXP-32     disease_state_design
E-MEXP-88     cellular_modification_design
E-MEXP-25     disease_state_design
MartShell>
```

# BioMart user interfaces: MartView

- Web-based user interface for BioMart
- Provides functionality for remote users to query all databases hosted by the BioMart server

# BioMart user interfaces: MartView

**START**

Select BioMart and Dataset

**FILTER**

Select a filter to restrict query e.g. Y chromosome

**OUTPUT**

Select the output (attributes) e.g. entrezgene

# BioMart user interfaces

- MartExplorer - stand alone client
- Perl and Java libraries
- MartEditor

# BioMart web service

- Web service:
  - A software system designed to support interoperable machine-to-machine interaction over a network.
  - Messages are typically conveyed using HTTP, and normally comprise XML in conjunction with other web-related standards



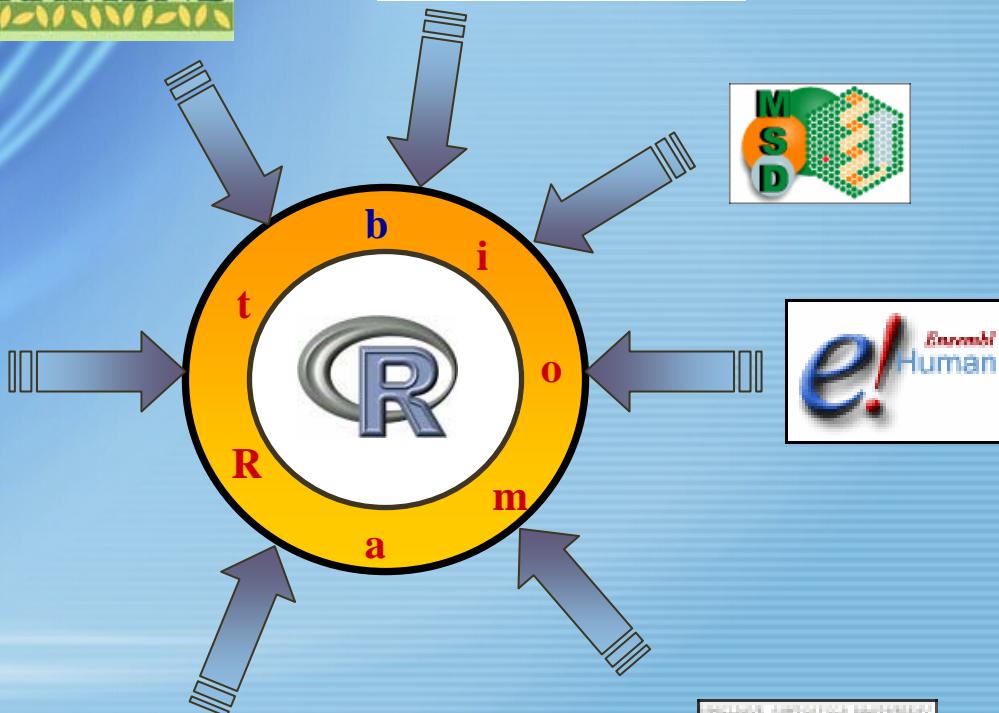
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# Integration of BioMart and R: biomaRt package



GRAMENE

Vega\*

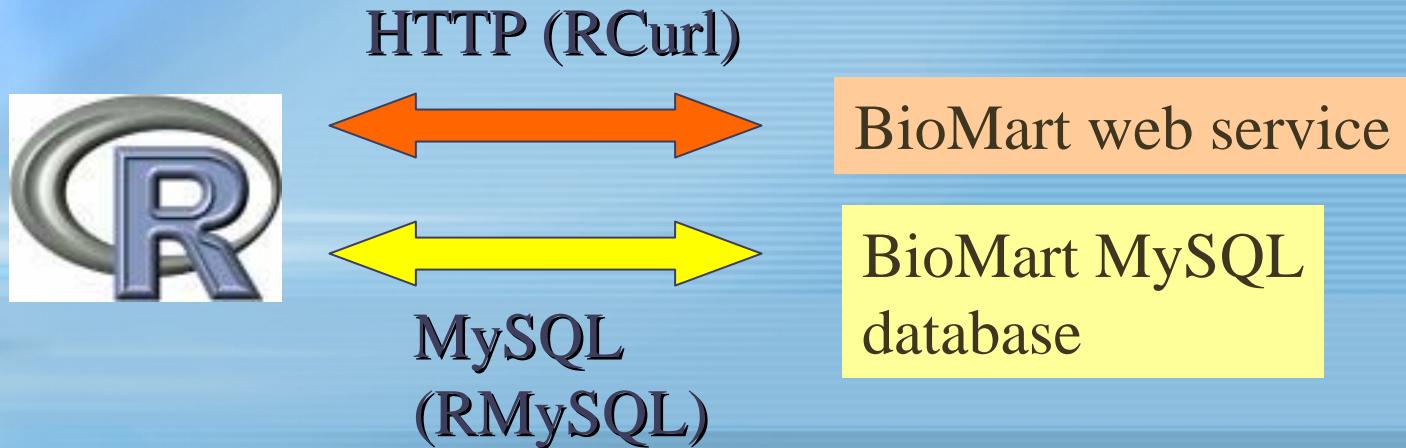


UniProt  
the universal  
protein resource

WormBase  
The Biology and  
Genome of  
*C. elegans*.

# biomaRt

- Direct HTTP queries to BioMart web services
- MySQL queries to BioMart databases



# biomaRt - Use

- Annotation of identifiers e.g. Affymetrix
- Retrieval of Gene Ontology, INTERPRO and other information
- Prioritizing groups of genes with particular properties
- Data mining

# Hands-on



- Installation
- Selecting a BioMart database and dataset
- Simple biomaRt functions tailored to Ensembl
- Generic biomaRt functions

# biomaRt installation

- biomaRt requires Rcurl  
<http://www.omegahat.org/Rcurl>
- biomaRt requires XML package
- RMySQL package is optional
- Platforms on which biomaRt has been installed:
  - Linux (curl <http://curl.haxx.se>)
  - OSX (curl)
  - Windows

# biomaRt installation

- Use biocLite

```
>source("http://www.bioconductor.org/bioc  
  Lite.R")  
> biocLite("biomaRt")
```

# Selecting a BioMart

```
> library( biomaRt )  
> listMarts()
```

# Selecting a BioMart

```
$biomart
```

```
[1] "dicty"    "ensembl"  "snp"      "vega"      "uniprot"  "msd"  
"wormbase"
```

```
$version
```

```
[1] "DICTYBASE (NORTHWESTERN)"  "ENSEMBL 39 (SANGER)"  
[3] "SNP 39 (SANGER)"          "VEGA 39 (SANGER)"  
[5] "UNIPROT PROTOTYPE 4-5 (EBI)" "MSD PROTOTYPE 4 (EBI)"  
[7] "WORMBASE CURRENT (CSHL)"
```

# Selecting a BioMart

```
> ensembl=useMart("ensembl")
```

# Selecting a dataset

> `listDatasets(ensembl)`

	dataset	version
1	<code>rnorvegicus_gene_ensembl</code>	RGSC3.4
2	<code>scerevisiae_gene_ensembl</code>	SGD1
3	<code>celegans_gene_ensembl</code>	CEL150
4	<code>trubripes_gene_ensembl</code>	FUGU4
5	<code>cintestinalis_gene_ensembl</code>	JGI2
6	<code>ptroglodytes_gene_ensembl</code>	CHIMP1A
7	<code>agambiae_gene_ensembl</code>	AgamP3
8	<code>hsapiens_gene_ensembl</code>	NCBI36

# Selecting a dataset

```
> ensembl =  
  useDataset("hsapiens_gene_ensembl",  
  mart=ensembl)
```

Or

```
> ensembl = useMart("ensembl",  
  dataset="hsapiens_gene_ensembl")
```

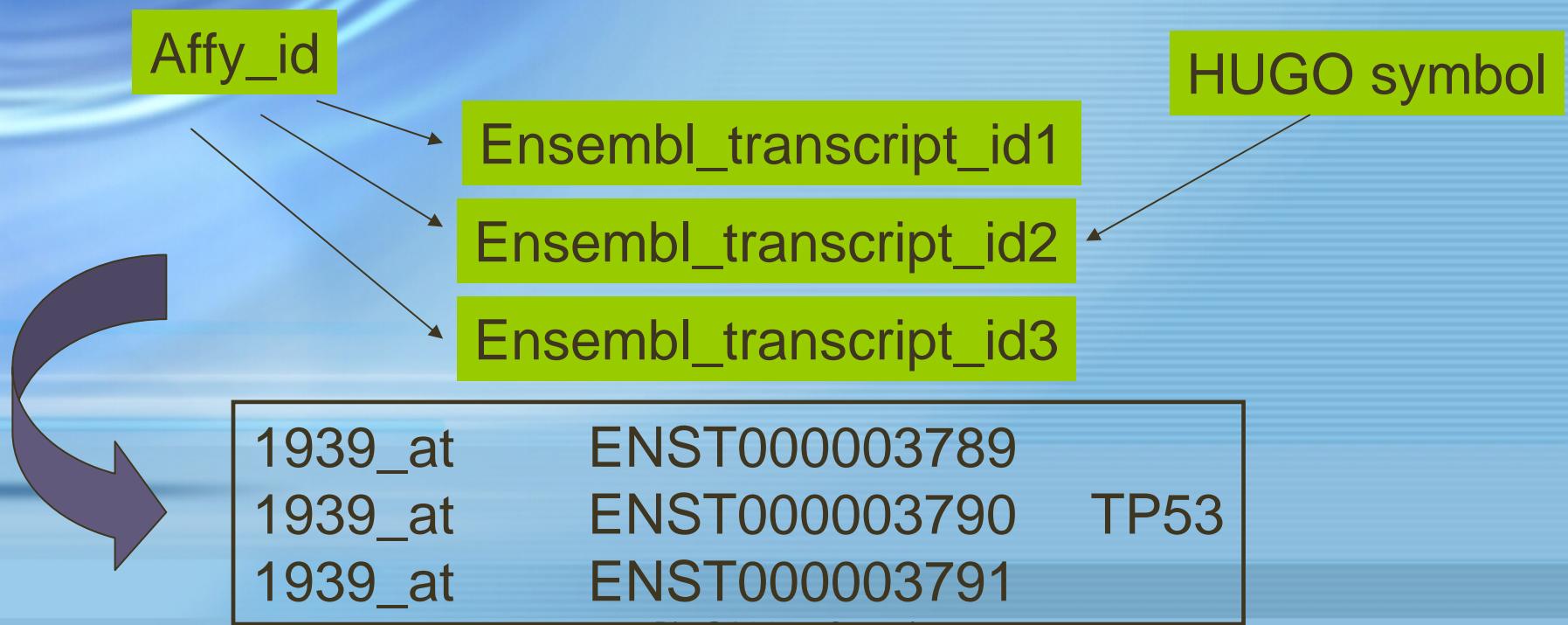


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# Simple biomarT functions tailored to Ensembl

# Ensembl annotation

- Ensembl annotation is at the transcript level



# getGene

Retrieves Gene annotations

- Gene symbol
- Description
- Chromosome name
- Band
- Start position
- End position
- Ensembl Gene ID
- Ensembl Transcript ID

# getGene

- Annotation of many types of identifiers such as:
  - EntrezGene
  - Affymetrix
  - Refseq
  - Embl
  - ...
- Output of all biomaRt “get” functions is a data.frame

# getGene

```
> affyids = c("202763_at","209310_s_at",
  "207500_at")  
  
> getGene(id=affyids,array="affy_hg_u133_plus_2",
  mart=ensembl)
```

	ID	symbol
1	202763_at	CASP3
2	207500_at	CASP5
3	209310_s_at	CASP4

# getGene

## description

- 1 Caspase-3 precursor (EC 3.4.22.-) (CASP-3) (Apopain) (Cysteine protease CPP32) (Yama protein) (CPP-32) (SREBP cleavage activity 1) (SCA-1) [Contains: Caspase-3 p17 subunit; Caspase-3 p12 subunit]. [Source:Uniprot/SWISSPROT;Acc:P42574]
- 2 Caspase-5 precursor (EC 3.4.22.-) (CASP-5) (ICH-3 protease) (TY protease) (ICE(rel)-III) [Contains: Caspase-5 subunit p20; Caspase-5 subunit p10]. [Source:Uniprot/SWISSPROT;Acc:P51878]
- 3 Caspase-4 precursor (EC 3.4.22.-) (CASP-4) (ICH-2 protease) (TX protease) (ICE(rel)-II) [Contains: Caspase-4 subunit 1; Caspase-4 subunit 2]. [Source:Uniprot/SWISSPROT;Acc:P49662]



# getGene

chromosome band strand chromosome\_start chromosome\_end

1	4	q35.1	-1	185785845	185807623
2	11	q22.3	-1	104370180	104384909
3	11	q22.3	-1	104318810	104345373

`ensembl_gene_id`      `ensembl_transcript_id`

ENSG00000164305	ENST00000308394
ENSG00000137757	ENST00000260315
ENSG00000196954	ENST00000355546

# getGene

Note:

- Ensembl does an independent mapping of affy probe sequences to genomes
- If there is no clear match then that probe is not assigned to a gene

# getGO

- Retrieve Gene Ontology annotation for a list of identifiers
- Many identifiers can be used
- Returns GO id, GO description and evidence code

# getGO

```
> getGO(id=affyids, array="affy_hg_u133_plus_2", mart=ensembl)
```

	ID	go_id	go_description	evidence_code
1	202763_at	GO:0005515	protein binding	IPI
2	202763_at	GO:0008234	cysteine-type peptidase activity	IEA
3	202763_at	GO:0030693	caspase activity	TAS
4	202763_at	GO:0006508	proteolysis	IDA
5	202763_at	GO:0006915	apoptosis	IEA
6	202763_at	GO:0006917	induction of apoptosis	TAS
7	202763_at	GO:0005737	cytoplasm	IDA
8	202763_at	GO:0005737	cytoplasm	IEA

# getINTERPRO

- INTERPRO is an integrated resource for protein families, domains and functional sites. It integrates secondary structure databases such as PROSITE, PRINTS, SMART, Pfam, ProDom, etc.
- Retrieve INTERPRO annotation for a list of identifiers
- Many identifiers can be used
- Returns INTERPRO id, description

# getINTERPRO

```
> getINTERPRO(id=affyids[1],  
array="affy_hg_u133_plus_2",  
mart=ensembl)
```

# getINTERPRO

	ID	interpro_id	description
1	202763_at	IPR001309	Caspase, p20 subunit
2	202763_at	IPR002398	Peptidase C14, caspase precursor p45
3	202763_at	IPR011600	Peptidase C14, caspase catalytic
4	202763_at	IPR002138	Peptidase C14, caspase non-catalytic subunit p10

	ensembl_gene_id	ensembl_transcript_id
1	ENSG00000164305	ENST00000308394
2	ENSG00000164305	ENST00000308394
3	ENSG00000164305	ENST00000308394
4	ENSG00000164305	ENST00000308394

# Pre-selection of features

- Select all Affymetrix identifiers on the hgu133plus2 chip for genes located on chromosome 16 between base pair 1100000 and 1250000

```
> features = getFeature(  
  array = "affy_hg_u133_plus_2",  
  chromosome = "16",  
  start = "1100000",  
  end="1250000", mart=ensembl)
```

# Pre-selection of features

	ensembl_transcript_id	chromosome_name	start_position	end_position	affy_hg_u133_plus_2
1	ENST00000358590	16	1143739	1211772	222960_at
2	ENST00000358590	16	1143739	1211772	205845_at
3	ENST00000356546	16	1143739	1211772	222960_at
4	ENST00000356546	16	1143739	1211772	205845_at
5	ENST00000234798	16	1211659	1215257	220339_s_at
6	ENST00000357113	16	1218338	1220215	207741_x_at

# Pre-selection of features

```
> unique(features[,5])
```

```
"222960_at" "205845_at" "220339_s_at" "207741_x_at" "215382_x_at"  
"210084_x_at" "205683_x_at" "207134_x_at" "217023_x_at" "216474_x_at"  
"214568_at"
```

# Pre-selection of features

- Select all entrezgene ids which have a "MAP kinase activity" GO term associated with it

```
> getFeature(type="entrezgene",
  GOID="GO:0004707", mart=ensembl)
```

# Pre-selection of features

	GO	entrezgene
1	GO:0004707	5598
2	GO:0004707	5598
3	GO:0004707	51701
4	GO:0004707	5596
5	GO:0004707	5595

# getSequence

- Retrieve sequences starting from a vector of identifiers or chromosomal coordinates
- 5'UTR
- 3'UTR
- cDNA
- protein

# getSequence

```
> getSequence(chromosome=3,  
  start=185514033,end=185535839,  
  seqType="5utr", mart=ensembl)
```

```
CCGGCTGCGCCTGCGGAGAAGCGGTGGCCGCCAGCGGGATCTGTGCAGGGAGCC  
GGAAATGGTTGTGGACTACGTCTGTGCGGCTGCGTGGGGCTCGGCCGCGCGACTG  
AAGGAGACTGAAGGGCGTTCCACATACGTTGTCCCACACAGCAGTACCCGTGC  
AGCCAGGAGCCCCAGGCTTCTATCCAGGTGCAAGCCCTACAGAATTGGGACCTAC  
GCTGGCGCCTACTATCCAGCCAAGGGGTGCAGCAGTTCCACTGGCGTGGCCCC  
CACCCCCAGTTTG
```



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# getSequence

```
>getSequence(chromosome=3,  
             start=185514033,end=185535839,  
             seqType="cdna", mart=ensembl)
```

```
CCGGCTGCGCCTGCGGAGAAGCGGTGGCCGCCAGCGGGATCTGTGCGGGAGCC  
GGAAATGGTTGTGGACTACGTCTGTGCGGCTGCGTGGGGCTCGGCCGCGCGACTG  
AAGGAGACTGAAGGGCGTTCCACATACGTTGTCCCACACAGCAGTACCTGTGC  
AGCCAGGAGCCCCAGGCTTCTATCCAGGTGCAAGCCCTACAGAATTGGGACCTAC  
GCTGGCGCCTACTATCCAGCCAAGGGGTGCAGCAGTTCCACTGGCGTGGCCCC  
CACCCCCAGTTTGATGAACCAGCCACCCAGATTGCTCCAAGAGGGAGCGTAAGA  
CGATCCGAATTGAGATCCAAACCAAGGAGGAAAGGATATCACAGAGGAGATCATG  
TCTGGGGCCCCGCACTGCCTCCACACCCACCCCTCCCC.....
```



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# getSequence

```
>getSequence(chromosome=3,  
             start=185514033,end=185535839,  
             seqType="peptide", mart=ensembl)
```

MNQPPQIAPKRERKTIRIDPNQGGKDITEEIMSGARTASTPT  
PPQTGGGLEPQANGETPQVAVIVRPDDRSQGAIIDRPGLPG  
PEHSPSESQPSSPSPTPSPSPVLEPGSEPNLAVLSIPGDTMT  
TI.....

# getSNP

- SNP: Single Nucleotide Polymorphisms, are common DNA sequence variations among individuals
- dbSNP is mirrored by Ensembl in its.snp BioMart.
- getSNP retrieves tsc-ids and refsnp identifiers together with allele, chromosome start and strand information.

# getSNP

```
> snpmart = useMart("snp", dataset =
  "hsapiens.snp")  
  
> snp=getSNP(chromosome = 8, start =
  148350, end = 148612, mart =
  snpmart)
```

tscid	refsnp_	id	allele	chrom_start	chrom_strand
1	TSC1723456	rs3969741	C/A	148394	1
2	TSC1421398	rs4046274	C/A	148394	1
3	TSC1421399	rs4046275	A/G	148411	1
4		rs13291	C/T	148462	1
5	TSC1421400	rs4046276	C/T	148462	1
6		rs4483971	C/T	148462	1
7		rs17355217	C/T	148462	1
8		rs12019378	T/G	148471	1
9	TSC1421401	rs4046277	G/A	148499	1
10		rs11136408	G/A	148525	1
11	TSC1421402	rs4046278	G/A	148533	1
12		rs17419210	C/T	148533	-1
13		rs28735600	G/A	148533	1
14	TSC1737607	rs3965587	C/T	148535	1
15		rs4378731	G/A	148601	1

# getHomolog

- 18 different species in Ensembl are interlinked
- biomaRt takes advantage of this to provide homology mappings between different species
- Combine two Ensembl datasets

# getHomolog

```
> human =  
  useMart("ensembl", "hsapiens_gene_ensembl")  
  
> mouse =  
  useMart("ensembl", "mmusculus_gene_ensembl")
```

# getHomolog

```
> homolog = getHomolog( id = "1939_at",
  to.array = "affy_mouse430_2",
  from.array = "affy_hg_u95av2",
  from.mart = human,
  to.mart = mouse )  
  
> homolog
```

V1

V2

V3

1	ENSMUSG00000059552	ENSMUST00000005371	1427739_a_at
2	ENSMUSG00000059552	ENSMUST00000005371	1426538_a_at

# getHomolog

```
> homolog = getHomolog( id = "NM_007294",
  to.array = "affy_mouse430_2",
  from.type = "refseq",
  from.mart = human,
  to.mart = mouse )
```

```
> homolog
```

	V1	V2	V3	
1	ENSMUSG00000017146	ENSMUST00000017290	1424629_at	
2	ENSMUSG00000017146	ENSMUST00000017290	1451417_at	
3	ENSMUSG00000017146	ENSMUST00000017290	1424630_a_at	



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# Generic biomaRt queries

# Generic biomarT queries

- Previous functions were all tailored to Ensembl BioMart
- Generic functions can be used to any available BioMart database and are modeled after MQL
- Generic functions enable one to query everything that is made available by the database

# Filters

- Filters define restrictions on the query
- Conceptually filters are inputs
- Example filters:
  - entrezgene
  - chromosome\_name

# listFilters

- Returns vector of all filters available in the selected BioMart

```
> listFilters(ensembl)
```

```
[1] "affy_hc_g110"  
[3] "affy_hg_u133_plus_2"  
[5] "affy_hg_u133a_2"
```

```
"affy_hg_focus"  
"affy_hg_u133a"  
"affy_hg_u133b"
```

```
.....
```

```
[15] "agilent_probe"  
[17] "ccds"  
[19] "embl"
```

```
"biotype"
```

```
"chromosome_name"  
"end"
```

```
.....
```



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# Attributes

- Attributes define the values which the user is interested in.
- Conceptually equal to output of the query
- Example attributes:
  - chromosome\_name
  - band

# listAttributes

```
> listAttributes(ensembl)
```

```
[1] "adf_embl"
```

.....

```
[14] "affy_hg_u95av2"
```

```
[15] "affy_hg_u95b"
```

.....

```
[21] "agilent_cgh"
```

```
[22] "agilent_probe"
```

```
[23] "allele"
```

```
[24] "allele_frequency"
```

```
[25] "band"
```

.....

# Generic biomarT queries

`hsapiens_gene_ensembl`



Attributes (e.g.,  
chromosome  
and band)

Filters (e.g.,  
“entrezgene”)

Values (e.g.,  
EntrezGene  
identifiers)

**biomarT query**

# getBM

- Generic biomaRt query function
- Contains no hard-coded information
- Is used by the simple biomaRt functions (which do contain hard-coded names of attributes and filters)

# getBM

```
> getBM(  
  attributes=c("affy_hg_u95av2","hgnc_symbol",  
  "chromosome_name","band"),  
  filters="affy_hg_u95av2",  
  values=c("1939_at","1454_at"), mart=mart)
```

		affy_hg_u95av2	hgnc_symbol	chromosome_name
		band		
1	1454_at	SMAD3		15 q22.33
2	1939_at	TP53		17 p13.1

# getBM: homology mapping

- Within one Ensembl dataset there are attributes providing mappings to the other Ensembl species
- Example:  
starting from the hsapiens dataset and a list of entrezgene ids we can query chromosomal positions of the corresponding genes in human, zebrafish, mouse and mosquito.

# getBM: homology mapping

```
> getBM(attributes=
  c("hgnc_symbol", "chromosome_name", "start_position",
    "mouse_chromosome", "mouse_chrom_start",
    "zebrafish_chromosome", "zebrafish_chrom_start",
    "mosquito_chromosome", "mosquito_chrom_start"),
  filter="entrezgene",
  values = c("673", "7157", "837"),
  mart=ensembl)
```

hgnc_symbol	chromosome_name	start_position
BRAF	7	140080754
TP53	17	7512464
CASP4	11	104318810

mouse_chromosome	mouse_chrom_start
6	39543731
11	69396600
9	5308874

zebrafish_chromosome	zebrafish_chrom_start
4	9473158
5	16155000
16	47717138

mosquito_chromosome	mosquito_chrom_start
2L	1974599
2R	20538788
	NA

# Using Wormbase

```
> listMarts()
```

```
$biomart
[1] "dicty"     "ensembl"   "snp"       "vega"      "uniprot"
     "msd"       "wormbase"
```

```
$version
[1] "DICTYBASE (NORTHWESTERN)"    "ENSEMBL 39
     (SANGER)"
[3] "SNP 39 (SANGER)"              "VEGA 39 (SANGER)"
[5] "UNIPROT PROTOTYPE 4-5 (EBI)" "MSD
     PROTOTYPE 4 (EBI)"
[7] "WORMBASE CURRENT (CSHL)"
```



BIOCONDUCTOR

# Using Wormbase

```
>wormbase=useMart("wormbase",dataset="gene")
> listFilters(wormbase)
> listAttributes(wormbase)
>getBM(attributes=c("name","rnai",
                      "rnai_phenotype","phenotype_desc"),
       filters="gene_name",
       values=c("unc-26","his-33"),
       mart=wormbase)
```

# Using Wormbase

name rnai	rnai_phenotype
phenotype_desc	

- 1 his-33 WBRNAi00000104 Emb | Nmo  
embryonic lethal | Nuclear morphology alteration in early embryo
- 2 his-33 WBRNAi00012233 WT  
wild type morphology
- 3 his-33 WBRNAi00024356 Ste  
sterile
- 4 his-33 WBRNAi00025036 Emb  
embryonic lethal

# Locally installed BioMarts

- Main use case currently is to use biomaRt to query public BioMart servers over the internet
- But you can also install BioMart server locally, populated with a copy of a public dataset (particular version), or populated with your own data
- Versioning is supported by naming convention



BIOCONDUCTOR

# Discussion

- Using biomaRt to query public web services gets you started quickly, is easy and gives you access to a large body of metadata in a uniform way
- Need to be online
- Online metadata can change behind your back; although there is possibility of connecting to a particular, immutable version of a dataset

# Reporting bugs

- Check with MartView if you get the same output
  - Yes: contact database e.g. [helpdesk@ensembl.org](mailto:helpdesk@ensembl.org)
  - No: contact me

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