

Package ‘MiPP’

November 17, 2025

Version 1.83.0

Date 2007-01-31

Title Misclassification Penalized Posterior Classification

Author HyungJun Cho <hj4cho@korea.ac.kr>,
Sukwoo Kim <s4kim@korea.ac.kr>,
Mat Soukup <soukup@fda.gov>, and
Jae K. Lee <jaeklee@virginia.edu>

Maintainer Sukwoo Kim <s4kim@korea.ac.kr>

Depends R (>= 2.4)

Imports Biobase, e1071, MASS, stats

Description This package finds optimal sets of genes that separate samples into two or more classes.

License GPL (>= 2)

URL <http://www.healthsystem.virginia.edu/internet/hes/biostat/bioinformatics/>

biocViews Microarray, Classification

git_url <https://git.bioconductor.org/packages/MiPP>

git_branch devel

git_last_commit b118e70

git_last_commit_date 2025-10-29

Repository Bioconductor 3.23

Date/Publication 2025-11-16

Contents

colon	2
cv.mipp.rule	2
get.mipp	3
get.mipp.lda	3
get.mipp.logistic	3
get.mipp.qda	3
get.mipp.svm.linear	3

get.mipp.svm.rbf	4
leuk1	4
leuk2	4
leukemia	5
linearkernel.decision.function	5
mipp	6
mipp.preproc	8
mipp.rule	9
mipp.seq	9
pre.select	11
quant.normal	12
quant.normal2	12
rbfkernel.decision.function	12
Index	13

colon	<i>Gene expression data for colon cancer</i>
-------	--

Description

This data set consists of gene expression of colon cancer study.

Usage

data(colon)

Format

A matrix containing 2000 probe sets and 2 classes (T, F)

Source

Alon, U., Barkai, N., Notterman, D.A., Gish, K., Ybarra, S., Mack, D., Levine, A.J. (1999). Broad Patterns of Gene Expression Revealed by Clustering Analysis of Tumor and Normal Colon Tissues probed by Oligonucleotide Arrays, PNAS, 96(12), 6745–6750.

cv.mipp.rule	<i>Fitting cross-validation MiPP</i>
--------------	--------------------------------------

Description

Fits cross-validation MiPP

get.mipp	<i>Choosing a rule</i>
----------	------------------------

Description

Choose a rule to compute MiPP

get.mipp.lda	<i>Fitting LDA to compute MiPP</i>
--------------	------------------------------------

Description

Fits LDA to compute MiPP

get.mipp.logistic	<i>Fitting logistic model to compute MiPP</i>
-------------------	---

Description

Fits logistic model to compute MiPP

get.mipp.qda	<i>Fitting QDA to compute MiPP</i>
--------------	------------------------------------

Description

Fits QDA to compute MiPP

get.mipp.svm.linear	<i>Fitting SVM (linear) to compute MiPP</i>
---------------------	---

Description

Fits SVM (linear) to compute MiPP

<code>get.mipp.svm.rbf</code>	<i>Fitting SVM (RBF) to compute MiPP</i>
-------------------------------	--

Description

Fits SVM (RBF) to compute MiPP

<code>leuk1</code>	<i>Gene expression data for leukemia</i>
--------------------	--

Description

This data set consists of gene expression of leukemia study.

Usage

```
data(leukemia)
```

Format

A matrix containing 6817 probe sets and 38 samples (2 classes: AML, ALL)

Source

Golub, T.R., Slonim, D.K., Tamayo, P., Huard, C., Gaasenbeek, M., Mesirov, P., Coller, H., Loh, M.L., Downing, J.R., Caliguri, M.A., Bloomfield, C.D., and Lander, E.S. (1999) Molecular classification of cancer: class discovery and class prediction by gene expression monitoring. *Science*, 286, 531-537.

<code>leuk2</code>	<i>Gene expression data for leukemia</i>
--------------------	--

Description

This data set consists of gene expression of leukemia study.

Usage

```
data(leukemia)
```

Format

A matrix containing 6817 probe sets and 34 samples (2 classes: AML, ALL)

Source

Golub, T.R., Slonim, D.K., Tamayo, P., Huard, C., Gaasenbeek, M., Mesirov, P., Coller, H., Loh, M.L., Downing, J.R., Caliguri, M.A., Bloomfield, C.D., and Lander, E.S. (1999) Molecular classification of cancer: class discovery and class prediction by gene expression monitoring. *Science*, 286, 531-537.

leukemia

Gene expression data for leukemia

Description

This data set consists of gene expression of leukemia study.

Usage

```
data(leukemia)
```

Format

A matrix containing 6817 probe sets and 2 classes (AML, ALL)

Source

Golub, T.R., Slonim, D.K., Tamayo, P., Huard, C., Gaasenbeek, M., Mesirov, P., Coller, H., Loh, M.L., Downing, J.R., Caliguri, M.A., Bloomfield, C.D., and Lander, E.S. (1999) Molecular classification of cancer: class discovery and class prediction by gene expression monitoring. *Science*, 286, 531-537.

```
linearkernel.decision.function
```

SVM (linear) kernel to compute MiPP

Description

SVM (linear) kernel to compute MiPP

mipp

*MiPP-based Classification***Description**

Finds optimal sets of genes for classification

Usage

```
mipp(x, y, x.test = NULL, y.test = NULL, probe.ID = NULL,
     rule = "lda", method.cut = "t.test", percent.cut = 0.01,
     model.sMiPP.margin = 0.01, min.sMiPP = 0.85, n.drops = 2,
     n.fold = 5, p.test = 1/3, n.split = 20,
     n.split.eval = 100)
```

Arguments

x	data matrix
y	class vector
x.test	test data matrix if available
y.test	test class vector if available
probe.ID	probe set IDs; if NULL, row numbers are assigned.
rule	classification rule: "lda", "qda", "logistic", "svmlin", "svmrbf"; the default is "lda".
method.cut	method for pre-selection; t-test is available.
percent.cut	proportion of pre-selected genes; the default is 0.01.
model.sMiPP.margin	smallest set of genes s.t. sMiPP <= (max sMiPP-model.sMiPP.margin); the default is 0.01.
min.sMiPP	Adding genes stops if max sMiPP is at least min.sMiPP; the default is 0.85.
n.drops	Adding genes stops if sMiPP decreases (n.drops) times, in addition to min.sMiPP criterion.; the default is 2.
n.fold	number of folds; default is 5.
p.test	partition percent of train and test samples when test samples are not available; the default is 1/3 for test set.
n.split	number of splits; the default is 20.
n.split.eval	numbr of splits for evalutation; the default is 100.

Value

model	candiadate genes (for each split if no indep set is available
model.eval	Optimal sets of genes for each split when no indep set is available

```
#####
#Example 1: When an independent test set is available

data(leukemia)

#Normalize combined data
leukemia <- cbind(leuk1, leuk2)
leukemia <- mipp.preproc(leukemia, data.type="MAS4")

#Train set
x.train <- leukemia[,1:38]
y.train <- factor(c(rep("ALL",27),rep("AML",11)))

#Test set
x.test <- leukemia[,39:72]
y.test <- factor(c(rep("ALL",20),rep("AML",14)))

#Compute MiPP
out <- mipp(x=x.train, y=y.train, x.test=x.test, y.test=y.test, probe.ID = 1:nrow(x.train), n.fold=5, percent.cut=0.5)

#Print candidate models
out$model

#####
#Example 2: When an independent test set is not available

data(colon)

#Normalize data
x <- mipp.preproc(colon)
y <- factor(c("T", "N", "T", "N", "T", "N", "T", "N", "T", "N",
  "T", "N", "T", "N", "T", "N", "T", "N", "T", "N",
  "T", "N", "T", "N", "T", "T", "T", "T", "T", "T",
  "T", "T", "T", "T", "T", "T", "T", "N", "T",
  "T", "N", "N", "T", "T", "T", "T", "N", "T", "N",
  "N", "T", "T", "N", "N", "T", "T", "T", "T", "N",
```

```

      "T", "N"))

#Deleting comtaminated chips
x <- x[,-c(51,55,45,49,56)]
y <- y[ -c(51,55,45,49,56)]

#Compute MiPP
out <- mipp(x=x, y=y, probe.ID = 1:nrow(x), n.fold=5, p.test=1/3, n.split=5, n.split.eval=100,
percent.cut= 0.1, rule="lda")

#Print candidate models for each split
out$model

#Print optimal models and independent evaluation for each split
out$model.eval

```

mipp.preproc

Preprocessing

Description

Performs IQR normalization, thesholding, and log2-transformation

Usage

```
mipp.preproc(x, data.type = "MAS5")
```

Arguments

x	data
data.type	data type is MAS5, MAS4, or dChip

See Also

[mipp](#)

Examples

```

library(MiPP)

data(colon)
colon.nor <- mipp.preproc(colon)

```


mipp.rule

*Computing MiPP***Description**

Computes MiPP

mipp.seq

*MiPP-based Classification***Description**

sequentially finds optimal sets of genes for classification

Usage

```
mipp.seq(x, y, x.test = NULL, y.test = NULL, probe.ID = NULL,
        rule = "lda", method.cut = "t.test", percent.cut = 0.01,
        model.sMiPP.margin = 0.01, min.sMiPP = 0.85, n.drops = 2,
        n.fold = 5, p.test = 1/3, n.split = 20, n.split.eval = 100,
        n.seq=3, cutoff.sMiPP=0.7, remove.gene.each.model="all")
```

Arguments

x	data matrix
y	class vector
x.test	test data matrix if available
y.test	test class vector if available
probe.ID	probe set IDs; if NULL, row numbers are assigned.
rule	classification rule: "lda", "qda", "logistic", "svmlin", "svmrbf"; the default is "lda".
method.cut	method for pre-selection; t-test is available.
percent.cut	proportion of pre-selected genes; the default is 0.01.
model.sMiPP.margin	smallest set of genes s.t. sMiPP <= (max sMiPP-model.sMiPP.margin); the default is 0.01.
min.sMiPP	Adding genes stops if max sMiPP is at least min.sMiPP; the default is 0.85.
n.drops	Adding genes stops if sMiPP decreases (n.drops) times, in addition to min.sMiPP criterion.; the default is 2.
n.fold	number of folds; default is 5.
p.test	partition percent of train and test samples when test samples are not available; the default is 1/3 for test set.

`n.split` number of splits; the default is 20.
`n.split.eval` numbr of splits for evaluation; the default is 100.
`n.seq` Number of sequential gene model selection; the default is 3.
`cutoff.sMiPP` Cutoff point of 5 percent sMiPP to select gene models
`remove.gene.each.model` Re-run after removing all genes in the selected models if "all" and the first gene for each of the selected models if "first"

Value

`model` candiadate genes (for each split if no indep set is available)
`model.eval` Optimal sets of genes for each split when no indep set is available
`genes.selected` a list of genes selected by sequential selection

Author(s)

Soukup M, Cho H, and Lee JK

References

Soukup M, Cho H, and Lee JK (2005). Robust classification modeling on microarray data using misclassification penalized posterior, *Bioinformatics*, 21 (Suppl): i423-i430.
 Soukup M and Lee JK (2004). Developing optimal prediction models for cancer classification using gene expression data, *Journal of Bioinformatics and Computational Biology*, 1(4) 681-694

Examples

```
#####
#Example 1: When an independent test set is available

data(leukemia)

#Normalize combined data
leukemia <- cbind(leuk1, leuk2)
leukemia <- mipp.preproc(leukemia, data.type="MAS4")

#Train set
x.train <- leukemia[,1:38]
y.train <- factor(c(rep("ALL",27),rep("AML",11)))

#Test set
x.test <- leukemia[,39:72]
y.test <- factor(c(rep("ALL",20),rep("AML",14)))

#Compute MiPP
out <- mipp.seq(x=x.train, y=y.train, x.test=x.test, y.test=y.test, n.fold=5, percent.cut=0.01, rule="lda", n.seq=3)

#Print candidate models
```

```

out$model

#Print the genes selected
out$genes.selected

#####
#Example 2: When an independent test set is not available

data(colon)

#Normalize data
x <- mipp.preproc(colon)
y <- factor(c("T", "N", "T", "N", "T", "N", "T", "N", "T", "N",
              "T", "N", "T", "N", "T", "N", "T", "N", "T", "N",
              "T", "N", "T", "N", "T", "T", "T", "T", "T", "T",
              "T", "T", "T", "T", "T", "T", "T", "T", "N", "T",
              "T", "N", "N", "T", "T", "T", "T", "N", "T", "N",
              "N", "T", "T", "N", "N", "T", "T", "T", "T", "N",
              "T", "N"))

#Deleting contaminated chips
x <- x[,-c(51,55,45,49,56)]
y <- y[ -c(51,55,45,49,56)]

#Compute MiPP
out <- mipp.seq(x=x, y=y, n.fold=5, p.test=1/3, n.split=5, n.split.eval=100,
percent.cut= 0.05, rule="lda", n.seq=2)

#Print candidate models for each split
out$model

#Print optimal models and independent evaluation for each split
out$model.eval

#Print the genes selected
out$genes.selected

```

pre.select

Pre-selection

Description

Pre-select genes

<code>quant.normal</code>	<i>Quantile normalization</i>
---------------------------	-------------------------------

Description

Performs quantile normalization

<code>quant.normal2</code>	<i>Quantile normalization</i>
----------------------------	-------------------------------

Description

Performs quantile normalization

<code>rbfkernel.decision.function</code>	<i>SVM (RBF) kernel to compute MiPP</i>
--	---

Description

SVM (RBF) kernel to compute MiPP

Index

* datasets

- colon, [2](#)
- leuk1, [4](#)
- leuk2, [4](#)
- leukemia, [5](#)

* models

- cv.mipp.rule, [2](#)
- get.mipp, [3](#)
- get.mipp.lda, [3](#)
- get.mipp.logistic, [3](#)
- get.mipp.qda, [3](#)
- get.mipp.svm.linear, [3](#)
- get.mipp.svm.rbf, [4](#)
- linearkernel.decision.function, [5](#)
- mipp, [6](#)
- mipp.preproc, [8](#)
- mipp.rule, [9](#)
- mipp.seq, [9](#)
- pre.select, [11](#)
- quant.normal, [12](#)
- quant.normal2, [12](#)
- rbfkernel.decision.function, [12](#)

colon, [2](#)

cv.mipp.rule, [2](#)

get.mipp, [3](#)

get.mipp.lda, [3](#)

get.mipp.logistic, [3](#)

get.mipp.qda, [3](#)

get.mipp.svm.linear, [3](#)

get.mipp.svm.rbf, [4](#)

leuk1, [4](#)

leuk2, [4](#)

leukemia, [5](#)

leukimia (leukemia), [5](#)

linearkernel.decision.function, [5](#)

mipp, [6](#), [8](#)

mipp.preproc, [8](#)

mipp.rule, [9](#)

mipp.seq, [9](#)

pre.select, [11](#)

quant.normal, [12](#)

quant.normal2, [12](#)

rbfkernel.decision.function, [12](#)