

Package ‘ChIPanalyser’

November 16, 2025

Type Package

Title ChIPanalyser: Predicting Transcription Factor Binding Sites

Version 1.33.0

Date 2017-09-01

Author Patrick C.N.Martin & Nicolae Radu Zabet

Maintainer Patrick C.N. Martin <pcnmartin@gmail.com>

Citation Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. Nucleic Acids Res., 43, 84–94.

Description ChIPanalyser is a package to predict and understand TF binding by utilizing a statistical thermodynamic model. The model incorporates 4 main factors thought to drive TF binding: Chromatin State, Binding energy, Number of bound molecules and a scaling factor modulating TF binding affinity. Taken together, ChIPanalyser produces ChIP-like profiles that closely mimic the patterns seen in real ChIP-seq data.

License GPL-3

Collate 2AllS4Class_ProfileParameters.R 3AllGenerics.R 4AllMethods.R
AllInitialize.R AllShowMethods.R computeChIPProfile.R
computeOccupancy.R computeOptimal.R computePWMScore.R
computeGenomeWide.R parallelInternalFunctionsDev.R
GenomicProfileGenericFunctions.R plotOccupancy.R
plotOptimalHeatMapDev.R DataPreprocessingDev.R
DataPreprocessingGenericFunctionsDev.R
profileAccuracyEstimateDev.R GAAnalysis.R GAGeneric.R

Depends R (>= 3.5.0), GenomicRanges, Biostrings, BSgenome, RcppRoll, parallel

Imports methods, IRanges,
S4Vectors, grDevices, graphics, stats, utils, rtracklayer, ROCR,
BiocManager, GenomeInfoDb, RColorBrewer

Suggests BSgenome.Dmelanogaster.UCSC.dm6, knitr, RUnit, BiocGenerics

Encoding UTF-8

LazyData true

biocViews Software, BiologicalQuestion, WorkflowStep, Transcription,
Sequencing, ChipOnChip, Coverage, Alignment, ChIPSeq,
SequenceMatching, DataImport ,PeakDetection

VignetteBuilder knitr

RoxygenNote 7.2.1

git_url <https://git.bioconductor.org/packages/ChIPAnalyser>

git_branch devel

git_last_commit 873269e

git_last_commit_date 2025-10-29

Repository Bioconductor 3.23

Date/Publication 2025-11-16

Contents

ChIPAnalyser-package	5
averageExpPWMScore	6
averageExpPWMScore-methods	7
backgroundSignal	8
backgroundSignal-methods	9
backgroundSignal<-	9
backgroundSignal<-methods	10
boundMolecules	11
boundMolecules-methods	12
boundMolecules<-	12
boundMolecules<-methods	13
BPFrequency	13
BPFrequency-methods	14
BPFrequency<-	15
BPFrequency<-methods	16
ChIPAnalyserData	16
chipMean	17
chipMean-methods	18
chipMean<-	19
chipMean<-methods	20
ChIPScore-class	20
chipSd	22
chipSd-methods	23
chipSd<-	23
chipSd<-methods	24
chipSmooth	24
chipSmooth-methods	25
chipSmooth<-	25
chipSmooth<-methods	26
computeChIPProfile	27
computeGenomeWideScores	29

computeOccupancy	30
computeOptimal	32
computePWMScore	34
DNASequenceLength	36
DNASequenceLength-methods	37
drop	38
drop-methods	39
evolve	39
generateStartingPopulation	41
genomicProfiles	42
genomicProfiles-class	43
genomicProfilesInternal-class	46
getHighestFitnessSolutions	48
getTestingData	49
getTrainingData	49
GRList-class	50
initialize-methods	51
lambdaPWM	51
lambdaPWM-methods	52
lambdaPWM<-	52
lambdaPWM<-methods	53
loci	53
loci-class	54
loci-methods	55
lociWidth	55
lociWidth-methods	56
lociWidth<-	57
lociWidth<-methods	58
maxPWMScore	58
maxPWMScore-methods	59
maxSignal	59
maxSignal-methods	60
maxSignal<-	61
maxSignal<-methods	62
minPWMScore	62
minPWMScore-methods	63
naturalLog	63
naturalLog-methods	64
naturalLog<-	65
naturalLog<-methods	66
noiseFilter	66
noiseFilter-methods	67
noiseFilter<-	67
noiseFilter<-methods	68
noOfSites	68
noOfSites-methods	69
noOfSites<-	70
noOfSites<-methods	71

nos-class	71
parameterOptions	72
parameterOptions-class	74
PFMFormat	77
PFMFormat-methods	78
PFMFormat<-	78
PFMFormat<-methods	79
ploidy	79
ploidy-methods	80
ploidy<-	81
ploidy<-methods	82
plotOccupancyProfile	82
plotOptimalHeatMaps	84
PositionFrequencyMatrix	86
PositionFrequencyMatrix-methods	87
PositionFrequencyMatrix<-	87
PositionFrequencyMatrix<-methods	88
PositionWeightMatrix	89
PositionWeightMatrix-methods	90
PositionWeightMatrix<-	90
PositionWeightMatrix<-methods	91
processingChIP	91
profileAccuracyEstimate	93
profiles-methods	95
PWMPseudocount	95
PWMPseudocount-methods	96
PWMPseudocount<-	96
PWMPseudocount<-methods	97
PWMThreshold	98
PWMThreshold-methods	99
PWMThreshold<-	99
PWMThreshold<-methods	100
removeBackground	100
removeBackground-methods	101
removeBackground<-	101
removeBackground<-methods	102
scores	103
scores-methods	104
searchSites	104
setChromatinStates	106
show-methods	107
singleRun	107
splitData	108
stepSize	109
stepSize-methods	110
stepSize<-	110
stepSize<-methods	111
strandRule	111

strandRule-methods	112
strandRule<-	113
strandRule<-methods	114
whichstrand	114
whichstrand-methods	115
whichstrand<-	115
whichstrand<-methods	116

Index	117
--------------	------------

ChIPAnalyser-package *ChIPAnalyser: Predicting Transcription Factor Binding Sites*

Description

ChIPAnalyser is a package to predict and understand TF binding by utilizing a statistical thermodynamic model. The model incorporates 4 main factors thought to drive TF binding: Chromatin State, Binding energy, Number of bound molecules and a scaling factor modulating TF binding affinity. Taken together, ChIPAnalyser produces ChIP-like profiles that closely mimic the patterns seen in real ChIP-seq data.

Details

The DESCRIPTION file: This package was not yet installed at build time.

Index: This package was not yet installed at build time.

Author(s)

Patrick C.N. Martin <pm16057@essex.ac.uk>

And

Nicolae Radu Zabet <nzabet@essex.ac.uk>

Maintainer: Patrick C.N. Martin <pcnmartin@gmail.com>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Examples

```
#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome
```

```

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
#Building data objects
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR",BPFrequency=DNASequenceSet)

chip<-processingChIP(chip,top)
# Computing Genome Wide
GenomeWide <- computeGenomeWideScores(DNASequenceSet = DNASequenceSet,
  genomicsProfiles = GPP)

#Compute PWM Scores
PWMScores <- computePWMScore(genomicsProfiles = GenomeWide,
  DNASequenceSet = DNASequenceSet,
  loci = top, chromatinState = Access)
#Compute Occupancy
Occupancy <- computeOccupancy(genomicsProfiles = PWMScores,
  parameterOptions = OPP)

#Compute ChIP profiles
chipProfile <- computeChIPProfile(genomicProfiles = Occupancy,
  loci = top,
  parameterOptions = OPP)
#Estimating accuracy estimate
AccuracyEstimate <- profileAccuracyEstimate(genomicProfiles = chipProfile,
  ChIPScore = chip,
  parameterOptions = OPP)

```

averageExpPWMScore	<i>Accessor for averageExpPWMScore slot in a genomicProfiles object.</i>
--------------------	--

Description

Extract or Access averageExpPWMScore slot in a [genomicProfiles](#)

Usage

```
averageExpPWMScore(object)
```

Arguments

object	object is a genomicProfiles
--------	-----------------------------

Details

As a general rule, averageExpPWMScore is computed and updated internally by [computeGenomeWideScores](#). Ideally, this slot should not be updated by user. The averageExpPWMScore is the sum of the exponential of every PWM score for a given DNA sequence and divided by the length of the said DNA sequence ([DNASequenceLength](#)). This can either be the full length sequence or only the accessible sequence (see [computeGenomeWideScores](#)).

Value

Returns the averageExpPWMScore of a [genomicProfiles](#) when computed.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Examples

```
# Accessing Data
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata", package="ChIPAnalyser"), "BEAF-32.pfm")
# Building genomicProfiles object
GPP <- genomicProfiles(PFM=PFM, PFMFormat="JASPAR")
# Extracting AllSitesAboveThreshold slot
averageExpPWMScore(GPP)

## Note this slot is now empty as nothing has yet been computed
```

averageExpPWMScore-methods

~~ *Methods for Function averageExpPWMScore* ~~

Description

~~ Methods for function averageExpPWMScore ~~

Methods:

signature(object = "genomicProfilesInternal")

backgroundSignal	<i>Accessor method for the backgroundSignal slot in a parameterOptions object.</i>
------------------	--

Description

Extract or access the backgroundSignal slot in a [parameterOptions](#) object.

Usage

```
backgroundSignal(object)
```

Arguments

object object is an [parameterOptions](#)

Details

Default Value: 0

When computing [computeOccupancy](#), a ChIP-seq background signal is used to scale Occupancy by considering both a backgroundSignal and a [maxSignal](#). The backgroundSignal is also used to normalise occupancies against maxOccupancy. The backgroundSignal usually comes from experimental data and is provided by user. As a general rule, if ChIP-seq data is available and will be used in [computeChIPProfile](#), [profileAccuracyEstimate](#) or [plotOccupancyProfile](#), it is advised to use the backgroundSignal from this data. We strongly encourage to set values when building a [parameterOptions](#) object.

Value

Returns a backgroundSignal of a [parameterOptions](#) object.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Building occupancyProfileParameters object
OPP <- parameterOptions()
#Viewing single value in object
backgroundSignal(OPP)
```

backgroundSignal-methods

~~ Methods for Function backgroundSignal ~~

Description

~~ Methods for function backgroundSignal ~~

Methods:

signature(object = "parameterOptions")

backgroundSignal<- *Setter method for backgroundSignal slot in a [parameterOptions](#)*

Description

Setter method for backgroundSignal slot in a [parameterOptions](#)

Usage

```
backgroundSignal(object)<-value
```

Arguments

object	object is an parameterOptions object.
value	value is the value to be assigned to the backgroundSignal slot in parameterOptions . backgroundSignal should be a positive value. Default value is 0.

Details

Default value: 0. When computing [computeOccupancy](#), a ChIP-seq background signal is used to scale Occupancy by considering both a backgroundSignal and a [maxSignal](#). The backgroundSignal is also used to normalise occupancies to maxOccupancy. The backgroundSignal usually comes from experimental data and is provided by user. As a general rule, if ChIP-seq data is available and will be used in [computeChIPProfile](#), [profileAccuracyEstimate](#) or [plotOccupancyProfile](#), it is advised to use the backgroundSignal from this data. We strongly encourage to set values when building a [parameterOptions](#) object.

Value

Returns a [parameterOptions](#) object with a new value assigned to the backgroundSignal slot.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Building occupancyProfileParameters object
OPP <- parameterOptions()
# Setting new value for backgroundSignal
backgroundSignal(OPP) <- 0.2
# Viewing whole object with new updated value
OPP
#Viewing single value in object
backgroundSignal(OPP)
```

backgroundSignal<-methods

~~ *Methods for Function* backgroundSignal<- ~~

Description

~~ Methods for function backgroundSignal<- ~~

Methods:

backgroundSignal(object)<-value

boundMolecules	<i>Accessor methods for boundMolecules slot in parameterOptions object.</i>
----------------	---

Description

Extract or Access boundMolecules slot in [parameterOptions](#) object.

Usage

```
boundMolecules(object)
```

Arguments

object object is a [parameterOptions](#) object.

Details

Default value: 1000

When computing occupancy ([computeOccupancy](#)), a value for the number of bound Molecules to DNA is needed. This value can be updated and set in a [parameterOptions](#) object. If the number of molecules is unknown, it is possible to infer this value with [computeOptimal](#). We strongly encourage to set values when building a [parameterOptions](#) object.

Value

Returns boundMolecules slot in [parameterOptions](#) object.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. Nucleic Acids Res., 43, 84–94.

Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
#Checking single value by slot accessor
boundMolecules(OPP)
```

boundMolecules-methods

~~ Methods for Function boundMolecules ~~

Description

~~ Methods for function boundMolecules ~~

Methods:

signature(object = "parameterOptions")

boundMolecules<- *Setter method for the boundMolecules slot in a [parameterOptions](#) object.*

Description

Setter method for the boundMolecules slot in a [parameterOptions](#) object.

Usage

boundMolecules(object)<-value

Arguments

object	object is a parameterOptions object.
value	value is a positive integer or vector of positive integers describing the number of molecules bound to DNA. Default value is 1000.

Details

Default value: 1000 When computing occupancy ([computeOccupancy](#)), a value for the number of bound Molecules to DNA is needed. This value can be updated and set in a [parameterOptions](#) object. If the number of molecules is unknown, it is possible to infer this value with [computeOptimal](#). We strongly encourage to set values when building a [parameterOptions](#) object.

Value

Returns a [parameterOptions](#) object with an updated value for boundMolecules.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
# Setting new boundMolecules value in OPP
boundMolecules(OPP) <- 5000
#Checking value in whole object
OPP
#Checking single value by slot accessor
boundMolecules(OPP)
```

```
boundMolecules<-methods
```

```
~~ Methods for Function boundMolecules<- ~~
```

Description

~~ Methods for function boundMolecules<- ~~

Methods:

```
signature(object = "parameterOptions", value = "vector")
```

BPFrequency	<i>Accessor method for BPFrequency slot in a genomicProfiles object.</i>
-------------	--

Description

Extract or Access BPFrequency slot in a [genomicProfiles](#) object.

Usage

```
BPFrequency(object)
```

Arguments

object object is a [genomicProfiles](#)

Details

Default value is `c(0.25,0.25,0.25,0.25)` When generating a Position Weight Matrix from a Position Frequency Matrix, the probability of occurrence of each base pair (Base Pair Frequency) is necessary (as originally described by Gary Stormo). It is possible to set custom values for BPFfrequency with a vector of length 4 containing the probability of occurrence of each base pair (A,C,G,T) in order. If Base pair frequency is unknown, BPFfrequency will compute base pair frequency from a DNA sequence. The nature of this sequence can be a BSgenome or a [DNAStringSet](#). In order to decrease run time, it is advised to use [DNAStringSet](#)

Value

Returns BPFfrequency slot in [genomicProfiles](#) object.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Examples

```
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata", package="ChIPAnalyser"), "BEAF-32.pfm")
# Building genomicProfiles object
GPP <- genomicProfiles(PFM=PFM, PFMFormat="JASPAR")
#Extracting BPFfrequency slot
BPFfrequency(GPP)
```

BPFfrequency-methods *~~ Methods for Function BPFfrequency ~~*

Description

~~ Methods for function BPFfrequency ~~

Methods:

`signature(object = "genomicProfilesInternal")`

BPFrequency<- *Setter method for BPFrequency slot in a [genomicProfiles](#) object.*

Description

Setter method for BPFrequency slot in a [genomicProfiles](#) object. If base pair frequency is unknown, BPFrequency will compute base pair frequency from a DNA sequence.

Usage

```
BPFrequency(object)<-value
```

Arguments

object	object is a genomicProfiles object.
value	value can three different objects: A vector of length 4 containing the probability of occurrence of each base pair (A,C,G,T) in order. Default value is c(0.25,0.25,0.25,0.25). A BSgenome of the organism of interest. The base pair frequency will automatically be computed and updated in genomicProfiles. A DNAStringSet of the organisme of interest. The base pair frequency will automatically be computed and updated in genomicProfiles (Prefered method).

Details

Default value is c(0.25,0.25,0.25,0.25) When generating a Postion Weight Matrix from a Position Frequency Matrix, the probability of occurrence of each base pair (Base Pair Frequency) is necessary (as originally described by Gary Stormo). It is possible to set custom values for BPFrequency with a vector of length 4 containing the probability of occurrence of each base pair (A,C,G,T) in order. If Base pair frequency is unknown, BPFrequency will compute base pair frequency from a DNA sequence when building a [genomicProfiles](#) object. The nature of this sequence can be aBSgenome object or a [DNAStringSet](#). In order to decrease run time, it is advised to use [DNAStringSet](#).

Value

Returns a [genomicProfiles](#) object with an updated value for BPFrequency.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
# Building genomicProfiles object
GPP <- genomicProfiles(PFM=PFM,PFMFormat = "JASPAR", BPFrequency=DNASequenceSet)
# Updating BPFrequency
## !! Note!! BPFrequency is used to compute PWM from PFM
## IF updated after building GPP, then it will not influence PWM
## Advised to build with BPFrequency directly
BPFrequency(GPP) <- DNASequenceSet
BPFrequency(GPP) <- c(0.25,0.25,0.25,0.25)
```

BPFrequency<-methods *~~ Methods for Function BPFrequency<- ~~*

Description

~~ Methods for function BPFrequency<- ~~

Methods:

```
signature(object = "genomicProfilesInternal", value = "DNAStringSet")
signature(object = "genomicProfilesInternal", value = "vector")
```

ChIPAnalyserData	<i>ChIPAnalyserData</i>
------------------	-------------------------

Description

ChIPAnalyserData is derived from real biological data. The source organism is *Drosophila melanogaster*. The data can be described as genomic data as it contains DNA sequences, loci, genetic information, DNA accessibility data and ChIP-seq data.

Usage

```
data(ChIPAnalyserData)
```

Format

1. Accessis [GRanges](#) containing DNA Accesibility data for the sequences described above.
2. csis [GRanges](#) containing Chromatin State data for the sequences described above.
3. topis [GRanges](#) containing a locus of interest. In this case *eve strip Locus* on chromosome 2R in *Drosophila melanogaster*
4. chipis a [GRanges](#) containing ChIP score of the eve strip locus in *Drosophila melanogaster*.
5. geneRefis a [GRanges](#) containing UCSC gene reference information

Value

Returns a set of Rdata objects as described above.

Source

Transcription Factor PFM: Berkeley Drosophila Transcription Network Project (bdtncp.lbl.gov)

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
data(ChIPanalyserData)
```

chipMean

Accessor method for [chipMean](#) slot in a [parameterOptions](#) object.

Description

Accessor method for [chipMean](#) slot in a [parameterOptions](#) object.

Usage

```
chipMean(object)
```

Arguments

object object is a [parameterOptions](#)

Details

Default vlaue : 150 When computing ChIP-seq like profiles ([computeChIPProfile](#), the occupancy values given by `computeOccupancy` are transformed into ChIP-seq like profiles. The average size of a ChIP-seq peak was described by Kaplan (Kaplan et al. , 2011). It is advised to use the average width of ChIP peaks from actual ChIP-seq data. We strongly encourage to set values when building a [parameterOptions](#) object.

Value

Returns chipMean slot from a [parameterOptions](#) object.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Kaplan T.,Li X.-Y.,Sabo P.J.,Thomas S.,Stamatoyannopoulos J.A., Biggin M.D., EisenM.B. Quantitative models of the mechanisms that control genome-wide patterns of transcription factor binding during early Drosophila development, *PLoS Genet.*,2011, vol. 7 pg. e1001290

Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
#Accessing chipMean slot in OPP
chipMean(OPP)
```

chipMean-methods *~~ Methods for Function chipMean ~~*

Description

~~ Methods for function chipMean ~~

Methods:

`chipMean(object)`

chipMean<- *Access methods for chipMean slot in [parameterOptions](#) object.*

Description

Access methods for chipMean slot in [parameterOptions](#) object.

Usage

```
chipMean(object)<-value
```

Arguments

object	object is a parameterOptions object.
value	value is a positive numeric value that will be assigned to the chipMean slot. chipMean describes the average size of a ChIP-seq peak in base pairs.

Details

Default vlaue : 150 When computing ChIP-seq like profiles ([computeChIPProfile](#), the occupancy values given by `computeOccupancy` are transformed into ChIP-seq like profiles. The average size of a ChIP-seq peak was described by Kaplan (Kaplan et al. , 2011). It is advised to use the average width of ChIP peaks from actual ChIP-seq data. We strongly encourage to set values when building a [parameterOptions](#) object.

Value

Returns a [parameterOptions](#) object with an updated value for chipMean slot.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Kaplan T.,Li X.-Y.,Sabo P.J.,Thomas S.,Stamatoyannopoulos J.A., Biggin M.D.,EisenM.B. Quantitative models of the mechanisms that control genome-wide patterns of transcription factor binding during early Drosophila development, *PLoS Genet.*,2011, vol. 7 pg. e1001290

Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
# Setting new value for slot
chipMean(OPP) <- 250
```

```
chipMean<-methods      ~~ Methods for Function chipMean<- ~~
```

Description

```
~~ Methods for function chipMean<- ~~
```

Methods:

```
chipMean(object)<-value
```

```
ChIPScore-class      Class "ChIPScore"
```

Description

ChIPScore is the result of the [processingChIP](#) function. This object contains the extracted ChIP Score from ChIP data, the loci of interest and optional parameters associated to ChIPAnalyser. The loci of interest will either be user provided or the top n regions as defined by the reduce argument in processingChIP. This object has the sole purpose of aiding the storage and parsing of data and parameters.

Objects from the Class

Object of this class are created internally and will be parsed to other objects as is.

Slots

scores: Object of class "list" List of extracted ChIP scores
loci: Object of class "loci" GRanges containing loci of interest
ploidy: Object of class "numeric" Ploidy level of the organism
boundMolecules: Object of class "vector" Number of Bound molecules to the DNA
backgroundSignal: Object of class "numeric" ChIP background signal (average ChIP score)
maxSignal: Object of class "numeric" max ChIP signal
lociWidth: Object of class "numeric" Width of loci if reduce is used and no loci are provided
chipMean: Object of class "numeric" Average ChIP peak width
chipSd: Object of class "numeric" Standard Deviation of ChIP peak width
chipSmooth: Object of class "vector" Smoothing window width for ChIP score
stepSize: Object of class "numeric" Defining resolution size of ChIP like profiles (10bp = signal will be only considered every 10bp)
removeBackground: Object of class "numeric" Signal Threshold to be removed. Default removes all negative scores

noiseFilter: Object of class "character" Type of noise filter to be used on ChIP data.
PWMThreshold: Object of class "numeric" Threshold of PWM scores that will be selected
strandRule: Object of class "character" Rule to compute strand score (max, mean or sum)
whichstrand: Object of class "character" Which strand should be used to compute PWM scores.
lambdaPWM: Object of class "vector" Lambda value - Scaling factor to the PWM
naturalLog: Object of class "logical" PFM to PWM conversion log transform (natural log or log2)
noOfSites: Object of class "nos" Number of Sites in the PWM that should be used to compute PWM scores.
PWMPseudocount: Object of class "numeric" PWM pseudocount value for PFM to PWM conversion.
paramTag: Object of class "character" Internal Tag - Code progression

Extends

Class "[parameterOptions](#)", directly.

Methods

```

.loci<- signature(object = "ChIPScore", value = "loci"): ...
.scores<- signature(object = "ChIPScore", value = "list"): ...
initialize signature(.Object = "ChIPScore"): ...
loci signature(object = "ChIPScore"): ...
scores signature(object = "ChIPScore"): ...
show signature(object = "ChIPScore"): ...
  
```

Author(s)

Patrick C.N. Martin

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

See Also

[processingChIP](#)

Examples

```
showClass("ChIPScore")
```

chipSd	Accessor method for chipSd slot in a parameterOptions object.
--------	---

Description

Access or Extract chipSd slot in a [parameterOptions](#) object.

Usage

```
chipSd(object)
```

Arguments

object object is a [parameterOptions](#)

Details

When computing ChIP-seq like profiles ([computeChIPProfile](#), the occupancy values given by [computeOccupancy](#) are transformed into ChIP-seq like profiles. The average size of a ChIP-seq peak was described by Kaplan (Kaplan et al. , 2011). The average peak size is subject to variation. This variation is accounted for with chipSd. It is advised to use the standard deviation of ChIP peak width from actual ChIP-seq data. We strongly encourage to set values when building a [parameterOptions](#) object.

Value

Returns a [parameterOptions](#) object with an updated value for chipSd.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Kaplan T.,Li X.-Y.,Sabo P.J.,Thomas S.,Stamatoyannopoulos J.A., Biggin M.D., Eisen M.B. Quantitative models of the mechanisms that control genome-wide patterns of transcription factor binding during early Drosophila development, *PLoS Genet.*,2011, vol. 7 pg. e1001290

Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
# Accessing chipSd slot
chipSd(OPP)
```

chipSd-methods	~~ <i>Methods for Function chipSd</i> ~~
----------------	--

Description

~~ Methods for function chipSd ~~

Methods:

chipSd(object)

chipSd<-	Setter methods for chipSd slot in a parameterOptions object.
----------	--

Description

Setter methods for chipSd slot in a [parameterOptions](#) object.

Usage

chipSd(object)<-value

Arguments

object	object is parameterOptions object.
value	value is a positive numeric value that will be assigned to chipSd slot. Default value is 150.

Details

When computing ChIP-seq like profiles ([computeChIPProfile](#), the occupancy values given by `computeOccupancy` are transformed into ChIP-seq like profiles. The average size of a ChIP-seq peak was described by Kaplan (Kaplan et al. , 2011). The average peak size is subject to variation. This variation is accounted for with `chipSd`. It is advised to use the standard deviation of ChIP peak width from actual ChIP-seq data. We strongly encourage to set values when building a [parameterOptions](#) object.

Value

Returns a [parameterOptions](#) object with an updated value for `chipSd`.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Kaplan T., Li X.-Y., Sabo P.J., Thomas S., Stamatoyannopoulos J.A., Biggin M.D., Eisen M.B. Quantitative models of the mechanisms that control genome-wide patterns of transcription factor binding during early *Drosophila* development, *PLoS Genet.*, 2011, vol. 7 pg. e1001290

Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
# Setting new value for chipSd slot
chipSd(OPP) <- 250
```

chipSd<-methods	~~ <i>Methods for Function chipSd<-</i> ~~
-----------------	---

Description

~~ Methods for function chipSd<- ~~

Methods:

chipSd(object)<-value

chipSmooth	<i>Accessor methods for chipSmooth slot in a parameterOptions object.</i>
------------	---

Description

Access or Extract chipSmooth slot in a [parameterOptions](#) object.

Usage

```
chipSmooth(object)
```

Arguments

object object is a [parameterOptions](#) object.

Details

When computing ChIP-seq like ([computeChIPProfile](#)) profile from occupancy data (see [computeOccupancy](#)), the profiles are smoothed using a window of a given size. The default value is set at 250 base pairs. If chipSmooth is set to 0 then the profile will not be smoothed. We strongly encourage to set values when building a [parameterOptions](#) object.

Value

Returns the chipSmooth slot in an [parameterOptions](#) object.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
# Accessing chipSd slot
chipSmooth(OPP)
```

chipSmooth-methods *~~ Methods for Function chipSmooth ~~*

Description

~~ Methods for function chipSmooth ~~

Methods:

signature(object = "parameterOptions")

chipSmooth<- *Setter method for chipSmooth slot in [parameterOptions](#) object.*

Description

Setter method for chipSmooth slot in [parameterOptions](#) object.

Usage

```
chipSmooth(object) <- value
```

Arguments

object object is a [parameterOptions](#) object.
 value value is the positive numeric value to be assigned to the chipSmooth slot in [parameterOptions](#) Default value is 250 base pairs.

Details

When computing ChIP-seq like ([computeChIPProfile](#)) profile from occupancy data (see [computeOccupancy](#)), the profiles are smoothed using a window of a given size. The default value is set at 250 base pairs. If chipSmooth is set to 0 then the profile will not be smoothed. We strongly encourage to set values when building a [parameterOptions](#) object.

Value

Returns a [parameterOptions](#) object with an updated value for chipSmooth slot.

Author(s)

Patrick C.N Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
# Setting new value for chipSd slot
chipSmooth(OPP) <- 250
```

chipSmooth<-methods *~~ Methods for Function chipSmooth<- ~~*

Description

~~ Methods for function chipSmooth<- ~~

Methods:

signature(object = "parameterOptions", value = "vector")

computeChIPProfile	<i>Computing ChIP-seq like profiles from Occupancy data.</i>
--------------------	--

Description

computeChIPProfile compute ChIP-seq like profile from occupancy data. Occupancy data is computed using [computeOccupancy](#).

Usage

```
computeChIPProfile(genomicProfiles, loci, parameterOptions = NULL,
  norm = TRUE, method = c("moving_kernel", "truncated_kernel", "exact"),
  peakSignificantThreshold= NULL, cores=1, verbose = TRUE)
```

Arguments

genomicProfiles

genomicProfiles is the result of [computeOccupancy](#). This object should be a [genomicProfiles](#) object.

loci

loci is either a [GRanges](#) or [ChIPScore](#) object. ChIPScore-class will be the result of [processingChIP](#). This object represents the set of Loci you are interested in analysing. If you have followed the full ChIPanalyzer pipe line, you would have used the processingChIP function that would return a ChIPScore-class object containing your loci of interest. GRanges are also supported if you are only using part of the pipeline.

parameterOptions

parameterOptions is a [parameterOptions](#) object. This object is used to store the numerous parameters offered by ChIPanalyzer. This argument is optional as all arguments are also parse in both ChIPScore-class and genomicProfiles objects. If you wanted to make some last minute changes, [parameterOptions](#) is the way to go. We recommend that you set your desired options before hand.

norm

norm is a logical value. If TRUE, the ChIP-seq like profile will be normalised towards maximum Occupancy. If FALSE, the profile will be left as is.

method

method is a character string of one of the following: c("moving_kernel", "truncated_kernel", "exact"). If set to moving_kernel, the peaks will be approximated using Rcpp (Default). If set to truncated_kernel, the peaks will be approximated however this method does not require Rcpp. If set to exact, the peaks will not be approximated.

peakSignificantThreshold

peakSignificantThreshold is a threshold at which peaks will be selected. **IMPORTANT:** if you select "moving_kernel" as described in method then this threshold is a numeric value describing the peak tail height cutoff value (Default = 0.001). In the case of "truncated_kernel" and "exact", the threshold represents a distance in base pair from the peak summit at which the peak should be cut (Default = 1250). The default is set to NULL in this function. This just means that either the value is provided by user with the appropriate method. If not, the default will be selected depending on the method selected.

cores	cores is the number of cores that will be used to compute ChIP profiles.
verbose	verbose is a logical value. If TRUE, progress messages will be displayed in console. If FALSE, no progress messages will be displayed in console.

Details

computeChIPProfile converts Transcription Factor occupancy to a profile resembling the one of a ChIP-seq profile. Internally a few parameters are required to build a ChIP like profile. These parameters are either defined and stored in a [ChIPScore](#) object (Parameters are updated based on your ChIP data), a [genomicProfiles](#) (user defined at the start of the analysis) or a [parameterOptions](#) (if you want to update values as you go along)

Value

Returns a [genomicProfiles](#) object containing all ChIP-seq like profile for every combination of [lambdaPWM](#) and [boundMolecules](#) provided by the user.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
#Extracting Data
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
# Building genomicProfiles object
GPP <- genomicProfiles(PFM=PFM, PFMFormat="JASPAR",BPFrequency=DNASequenceSet)

# Computing Genome Wide
GenomeWide <- computeGenomeWideScores(genomicProfiles = GPP,
  DNASequenceSet = DNASequenceSet)
```

```
#Compute PWM Scores
PWMScores <- computePWMScore(genomicProfiles = GenomeWide,
                             DNASequenceSet = DNASequenceSet, loci = top, chromatinState = Access)
#Compute Occupancy
Occupancy <- computeOccupancy(genomicProfiles = PWMScores)

#Compute ChIP profiles
chipProfile <- computeChIPProfile(genomicProfiles=Occupancy,loci=top)
chipProfile
```

computeGenomeWideScores

Computing Genome Wide scores

Description

computeGenomeWideScores compute the max and min PWM score over the entire genome.

Usage

```
computeGenomeWideScores(genomicProfiles, DNASequenceSet, chromatinState = NULL, parameterOptions = NULL)
```

Arguments

genomicProfiles	genomicProfiles is a genomicProfiles object containing the PFM, PWM of interest.
DNASequenceSet	DNASequenceSet is a BSgenome or DNASTringSet containing the sequence of the organism of interest.
chromatinState	chromatinState is a GRanges object containing the chromatin States. This can either represent regions of accessible DNA or Chromatin state affinities.
parameterOptions	parameterOptions is a parameterOptions object containing parameters that you wish to change. The genomicProfiles object will be updated using the values assigned to parameterOptions
cores	cores is the number of cores that will be used (Numeric value - Default = 1)
verbose	verbose is a logical value that will determine if internal progress message will be printed.

Details

computeGenomeWideScores function computes PWM scores over the entire genome (or accessible Genome if chromatin State are provided). Genome wide scores are used to determine the maximum and minimum PWM score as well as the average exponential score. These scores will in turn be used to determine which score are above the PWM threshold. The average exponential score is an integrale part of the equation used to compute Occupancy. Using default settings, ChIPAnalyser will only compute occupancy on the top 70% of PWM scores. This threshold can be changed. See [PWMThreshold](#)

Value

Returns a `genomicProfiles` object with updated values for max score, min score and average-ExpPWMScore.

Author(s)

Patrick C.N Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
# Building genomicProfiles object
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR", BPFrequency=DNASequenceSet)

# Computing Genome Wide
GenomeWide <- computeGenomeWideScores(genomicProfiles = GPP,
  DNASequenceSet = DNASequenceSet)
```

computeOccupancy

Compute Occupancy values from PWM Scores based on model.

Description

computeOccupancy will compute the Occupancy from PWM Scores. As described in detail in the vignette, ChIPanalyser uses PWM Scores, DNA Accessibility data, the number of bound molecules and a scaling factor of Transcription Factor specificity. This function will compute occupancy using the values assigned to each variable.

Usage

```
computeOccupancy(genomicProfiles,parameterOptions = NULL,
  norm = TRUE, verbose = TRUE)
```

Arguments

genomicProfiles	genomicProfiles is a genomicProfiles object resulting from computePWMscore . It is important to use this resulting object as the occupancy will only be computed for sites above a threshold.
parameterOptions	parameterOptions is a parameterOptions object containing the adequate values assigned to each Parameter. If not Supplied (parameterOptions = NULL), a new object will be created internally using default values.
norm	norm a logical value which determines if the occupancy should be normalised or not.
verbose	verbose a logical value which determines if progress messages are printed or not.

Details

computeOccupancy will compute the Occupancy from PWM Scores. As described in detail in the vignette, ChIPAnalyser uses PWM Scores, DNA Accessibility data, the number of bound molecules and a scaling factor of Transcription Factor specificity. This function will compute occupancy using the values assigned to each variable. It should also be noted that the [parameterOptions](#) object contains a set of parameters used to compute Occupancy (not only restricted to this). These parameters are often dependant on real ChIP-Seq data and will influence the goodness of fit between the predicted model and real ChIP-seq data. We strongly advise that the values assigned to each parameter should be customized in order to increase the model agreement with real world biological data.

Value

computeOccupancy will return a [genomicProfiles](#). The main difference will reside in the [profiles](#) slot. This slot is generally a list or [GRangesList](#). Within these list type structures are enclosed [GRanges](#) containing the positions of site above threshold, PWM Scores and Occupancy for each site. The series of GRanges will depend on the number of loci that are tested and the number of element in the list will depend on the various combinations of lambdaPWM and boundMolecules.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Examples

```
#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata", package="ChIPAnalyser"), "BEAF-32.pfm")
```

```

#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)

#Building data objects
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR",BPFrequency=DNASequenceSet)
OPP <- parameterOptions()
# Computing Genome Wide
GenomeWide <- computeGenomeWideScores(genomicProfiles = GPP,
  DNASequenceSet = DNASequenceSet)

#Compute PWM Scores
PWMScores <- computePWMScore(genomicProfiles = GenomeWide,
  DNASequenceSet = DNASequenceSet,
  loci = top,
  chromatinState = Access)
#Compute Occupancy
Occupancy <- computeOccupancy(genomicProfiles = PWMScores,
  parameterOptions = OPP)
Occupancy

```

computeOptimal

compute Optimal Parameters

Description

ChIPAnalyser contains a set of functions some of which require two parameters known as [lambdaPWM](#) and as [boundMolecules](#). These two parameters are not always known. computeOptimal will compute these values by maximising the correlation and minimising the Mean Squared Error between a predicted ChIP-seq-like profile and a real ChIP-seq profile for a given loci.

Usage

```

computeOptimal(genomicProfiles,DNASequenceSet, ChIPScore,chromatinState = NULL,
  parameterOptions = NULL, optimalMethod = "all",rank=FALSE,returnAll=TRUE,
  peakMethod="moving_kernel",cores=1)

```

Arguments

genomicProfiles

genomicProfiles is a [genomicProfiles](#) object containing at least a Position Frequency Matrix or a Position Weight Matrix. It is strongly advised to cus-

	tomize this object to increase goodness of fit of the model when compared to real ChIP-seq data.
DNASequenceSet	DNASequenceSet is a DNAStringSet or a BSgenome of the full sequence of the organism of interest.
ChIPScore	ChIPScore is a named list containing ChIP-seq enrichments for each Loci of interest. This Profile should be normalised to a base pair level. In other words, there should be an enrichment score for each base pair of a given Locus.
chromatinState	chromatinState is a GRanges object containing either accesible sites or DNA affinity scores.
parameterOptions	parameterOptions is a parameterOptions object. If this object is not provided (parameterOptions = NULL), a new object will be created internally. However, it is strongly advised to tailor this object to maximise the goodness of fit of the model when compared to ChIP-seq data.
optimalMethod	optimalMethod is a character string which determines which method for optimal parameter selection should be selected. optimalMethod can be one of the following: pearson, spearman, kendall, ks, fscore, geometric,MSE, or all. Default is set to all.
rank	rank is a logical value indicating if optimal parameters should be based on rank (parameter combination occuring the most over all regions) or average score (best performing combination of paramters on average over all regions selected). DEFAULT = FALSE
returnAll	returnAll is a logical value indicating if all internal objects should be returned. DEFAULT = TRUE. Internal objects are the following: Occupancy Scores, ChIP like profiles, goodness of fit metrics and optimal paramters. If set to FALSE, computeOptimal will only return the optimal parameters.
peakMethod	peakMethod is a character string of one of the following: c("moving_kernel","truncated_kernel","exact"). If set to moving_kernel, the peaks will be approximated using Rcpp (Default). If set to truncated_kernel, the peaks will be approximated however this method does not require Rcpp. If set to exact, the peaks will not be approximated.
cores	cores is the number cores that will be used to compute optimal set of parameters.

Details

In order to backward infer the values of [lambdaPWM](#) and [boundMolecules](#), it is possible to use the computeOptimal to find these parameters. It should be noted that this functions requires a ChIP-seq data input. ChIPScore (ChIP-seq data). This should be the output of the [processingChIP](#) function.

Value

computeOptimal returns a list respectively described as the optimal set of Parameters (lambda - [lambdaPWM](#) and [boundMolecules](#)), the optimal matrix (a matrix containing accuracy estimates dependant on the parameter chosen), and finally the chosen parameter. If the parameter that was chosen was "all", then each element of this list will contain the optimal set of parameters, optimal matrices for all of the aforementioned paramters (see optimalMethod).

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Examples

```
#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
chip<-processingChIP(chip,top)
#Building data objects
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR",BPFrequency=DNASequenceSet)
OPP <- parameterOptions()
#Computing Optimal set of Parameters
optimalParam <- computeOptimal(genomicProfiles = GPP,
  DNASequenceSet = DNASequenceSet,
  ChIPScore = chip,
  chromatinState = Access,
  parameterOptions = OPP,
  parameter = "all",
  peakMethod="moving_kernel")
```

computePWMScore

Compute PWM Scores of sites above threshold.

Description

computePWMScore will compute and extract all sites that exhibit a PWM Score higher than a threshold. This threshold (see [PWMThreshold](#)) will determine the percentage of total sites that should NOT be considered.

Usage

```
computePWMScore(genomicProfiles,DNASequenceSet,
  loci = NULL, chromatinState = NULL,parameterOptions=NULL,cores=1, verbose = TRUE)
```

Arguments

DNASequenceSet	DNASequenceSet is a DNAStrngSet or a BSgenome containing the full sequence of the organism of interest.
genomicProfiles	genomicProfiles is a genomicProfiles object resulting from the computeGenomeWideScores function.
loci	loci is a GRanges object containing the Loci of interest or a ChIPScore object result of processingChIP function.
parameterOptions	parameterOptions is a parameterOptions object containing parameters that you wish to parse/change when computing PWM Scores.
chromatinState	chromatinState is a GRanges object sites of accessible DNA or DNA affinity scores.
cores	cores is the number of cores used to compute PWM Scores.
verbose	verbose is a logical value indicating if progress messages should be printed or not.

Details

After determining genome wide scores, it is possible to only compute and extract high affinity sites (in the sense that they have a high PWM Score). If a [PWMThreshold](#) is not set by user, the default value is set at 0.7. This means that 70 % of sites will NOT be selected. Only the top 30 % will be computed and extracted. If one is interested in all PWM Scores at a genome wide scale (or accessible DNA), this is possible by setting [PWMThreshold](#) to zero.

Value

[computePWMScore](#) will return a [genomicProfiles](#) object. The [profiles](#) slot will have been updated. This slot will now contain a [GRangesList](#) with each element being a [GRanges](#). This GRanges will contain position of each sites (start, end and strand) and the PWM Score associated to that site.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. Nucleic Acids Res., 43, 84–94.

Examples

```
#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome
```

```

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
chip<-processingChIP(chip,top)
#Building data objects
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR",BPFrequency=DNASequenceSet)

# Computing Genome Wide
GenomeWide <- computeGenomeWideScores(DNASequenceSet = DNASequenceSet,
  genomicProfiles = GPP)

#Compute PWM Scores
PWMScores <- computePWMScore(DNASequenceSet = DNASequenceSet,
  genomicProfiles = GenomeWide,
  loci = chip, chromatinState = Access)
PWMScores

```

DNASequenceLength	<i>Accessor method for DNASequenceLength slot in a genomicProfiles</i>
-------------------	--

Description

Accessor method for DNASequenceLength slot in a [genomicProfiles](#)

Usage

```
DNASequenceLength(object)
```

Arguments

object object is a [genomicProfiles](#)

Details

The model on which is based ChIPanalyser requires the length of the DNA sequence used to compute scores. In this circumstance, this DNA Length is the total length of the DNA of the organism of interest or the the Accessible DNA at a genome wide scale.

Value

Returns DNASequenceLength slot in a [genomicProfiles](#) object.

Author(s)

Patrick C. N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Examples

```
#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)

#Building genomicProfiles object
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR",BPFrequency=DNASequenceSet)
# Computing Genome Wide
GenomceWide <- computeGenomeWideScores(DNASequenceSet = DNASequenceSet,
  genomicProfiles = GPP)

DNASequenceLength(GenomceWide)
```

DNASequenceLength-methods

~~ *Methods for Function* DNASequenceLength ~~

Description

~~ Methods for function DNASequenceLength ~~

Methods:

signature(object = "genomicProfilesInternal")

drop	<i>Accessor Method for the drop slot in a genomicProfiles object.</i>
------	---

Description

Accessor Method for the drop slot in a [genomicProfiles](#) object.

Usage

```
drop(object)
```

Arguments

object object is a [genomicProfiles](#) object.

Details

During certain computations, it is possible that the Loci of interest do not show any overlap with accessible DNA. If this were to be the case, a warning message will appear in the console but these inaccessible Loci will be stored in this slot. It is also for these reasons that it is imperative for Loci of interest to be named (in this case, a named [GRanges](#)).

Value

Returns a character string with loci containing no accessible DNA.

Author(s)

Patrick C.N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Examples

```
# Loading data
data(ChIPAnalyserData)
#Loading PFM files
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"), "BEAF-32.pfm")
#Building data objects
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR")

# Loci with no acces - a warning message will be issued
#if loci do not contain accessible DNA
# Otherwise this slot will remain empty

drop(GPP)
```

drop-methods	<i>~~ Methods for Function drop ~~</i>
--------------	--

Description

~~ Methods for function drop ~~

Methods:

signature(object = "genomicProfilesInternal")

evolve	<i>Running the ChIPAnalyser implementation of a Genetic algorithm.</i>
--------	--

Description

evolve pushes a starting population to evolve in a genetic algorithm.

Usage

```
evolve(population,DNASequenceSet,ChIPScore,
       genomicProfiles,parameters=NULL,generations=100,mutationProbability=0.3,
       offsprings=5,chromatinState=NULL,
       method="geometric", lambda=TRUE,
       checkpoint=TRUE,
       filename=NULL, cores=1)
```

Arguments

population	numeric value describing the number of individuals in the starting population. Alternatively - a starting population list as returned by generateStartingPopulation. NOTE: if numeric - the parameter argument is also required.
DNASequenceSet	DNAStringSet object containing DNA sequences of interest (Extracted from BSgenome)
ChIPScore	ChIPScore object as returned by the processingChIPfunction
genomicProfiles	genomicProfiles object containing minimal information (such as the PWM)
parameters	vector or list containing each parameter that should be added to the chromosome. See generateStartingPopulation
generations	numeric describing the number of generation before the Genetic algorithm should halt.
mutationProbability	numeric descrbining the rate of mutations for each surviving individual
offsprings	numeric descrbining the number of individuals surviving to the next generation

<code>chromatinState</code>	GRanges object containing chromatin state information. Each state should be labeled in a meta data column named "name". It is advised to use numeric values for each state name.
<code>method</code>	character string describing the scoring metric that should be used. ChIPAnalyser offers twelve different metrics: correlation coefficients (Pearson, Spearman and Kendall), Mean Squared Error (MSE), Kolmogorov–Smirnov Distance, precision, recall, accuracy, F-score, Matthew's correlation coefficient (MCC) and Area Under Curve Receiver Operator Characteristic (AUC ROC or just AUC)
<code>lambda</code>	logical describing if lambda value should be pre-computed. Setting to TRUE increases the speed of the algorithm.
<code>checkpoint</code>	logical describing if population parameters at each generations should be saved.
<code>filename</code>	character string that will serve as a prefix to the saved intermediate files.
<code>cores</code>	numeric describing the number of cores used to run the GA.

Details

ChIPAnalyser offers a way of finding optimal solution by using a genetic algorithm. Instead of running the standard analysis, TF binding affinities to chromatin states can be extracted via this more complex method. It should be noted that this method is better suited for the analysis of chromatin states. While the algorithm still works with simple DNA Accessibility, it would potentially take more time for accuracy minor gains.

Value

Returns a named list with three elements.

- `database` saves the data frame containing all scores for each individual since generation 1
- `population` saves the last population with chromosome values
- `fittest` saves the fittest individual for a given generation

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

Examples

```
library(ChIPAnalyser)
data(ChIPAnalyserData)
# See GA vignette for usage
```

`generateStartingPopulation`*Generate Starting population for ChIPAnalyser Genetic algorithm*

Description

`generateStartingPopulation` generates a starting population with random traits for each individual

Usage

```
generateStartingPopulation(population,parameters,names=NULL)
```

Arguments

<code>population</code>	numeric value describing the number of individuals in the starting population.
<code>parameters</code>	vector or list containing each parameter that should be added to the chromosome.
<code>names</code>	character describing names that should be added to each individual.

Details

`generateStartingPopulation` generates a starting population to be used in the genetic algorithm implemented in ChIPAnalyser. There are two main ways a starting population can be generated:

1. by name Using names of each parameter that should be parse to each "chromosome". The possible paramters are N, lambda, PWMThreshold, CS (DNAAffinity or DNAAccessibility also works). CS values should also contain a numeric value associated to each chromatin state you wish to parse. e.g CS1 ... CS14 This will generate a value by sampling from a set of predefined value for each paramters.
2. by value range Using a named list (names for each parameters). Each element of the list should contain three numeric values : length of range, min value, max value. (Internally - values are parse to `runif`)

Value

Returns a list of individuals with a random traits

Author(s)

Patrick C.N. Martin

Examples

```
## by name
param <- c("N","lambda","PWMThreshold","CS1","CS2","CS3")

pop <- generateStartingPopulation(20,param, names = NULL)

# by range
paramValue <- list(c(10,1,1000),c(10,0,5),c(10,0,0.9),c(10,0,1),c(10,0,1),c(10,0,1))

pop <- generateStartingPopulation(20,paramValue,names= param)
```

genomicProfiles	<i>Genomic Profile object</i>
-----------------	-------------------------------

Description

genomicProfiles is an S4 object serving two purposes: (i) storing internal computed data and (ii) storing parameter options. This object is parsed through the different steps of the pipeline to facilitate that parsing and changing of parameters.

Usage

```
genomicProfiles(..., parameterOptions = NULL, genomicProfiles = NULL, ChIPScore = NULL)
```

Arguments

...	Any of the user available slots in genomicProfiles.
parameterOptions	If some parameters were already previously computed or stored in a parameterOptions, parsing this object will use those values instead of the default ones.
genomicProfiles	If some parameters were already previously computed or stored in a genomicProfiles, parsing this object will use those values instead of the default ones.
ChIPScore	If some parameters were already previously computed or stored in a ChIPScore, parsing this object will use those values instead of the default ones.

Details

The genomicProfiles object serves the purpose of storing, and parsing parameters and computed data between the different steps of the pipeline. When creating a genomicProfiles object it is possible to use previously computed values by simply parsing the object to the constructor function.

Value

Returns a genomicProfiles object with updated slots for all parameters parsed.

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

See Also

[genomicProfiles](#)
[parameterOptions](#)

Examples

```
PFM <- file.path(system.file("extdata", package="ChIPAnalyser"), "BEAF-32.pfm")
genomicProfiles()
genomicProfiles(PFM=PFM, PFMFormat="JASPAR")
```

genomicProfiles-class *Class "genomicProfiles"*

Description

genomicProfiles is an S4 object serving two purposes: (i) storing internal computed data and (ii) storing parameter options. This object is parsed through the different steps of the pipeline to facilitate that parsing and changing of parameters.

Objects from the Class

Objects can be created by calls of the form `genomicProfiles(ploidy, boundMolecules, backgroundSignal, maxSignal, lociWidth, chipMean, chipSd, chipSmooth, stepSize, noiseFilter, removeBackground, lambdaPWM, PWMpseudocount, naturalLog, noOfSites, PWMThreshold, strandRule, whichstrand, PFM, PWM, PFMFormat, BPFrequency, minPWMScore, maxPWMScore, profiles, DNASequenceLength, averageExpPWMScore)`.

Slots

PWM: Object of class "matrix": A Position Weight Matrix (either supplied or internally computed if PFM is provided)

PFM: Object of class "matrix": A Position Frequency Matrix (may also be a path to file containing PFM)

PFMFormat: Object of class "character": A character string of one of the following: raw, trans-fac, JASPAR or sequences

BPFrequency: Object of class "vector": Base Pair Frequency in the genome (if a DNA sequence is provided (as a [DNAStrngSet](#) or BSgenome), will be automatically computed internally). Default:c(0.25,0.25,0.25,0.25)

minPWMScore: Object of class "vector": Lowest PWM score accross the genome (computed and updated internally)

maxPWMScore: Object of class "vector": Highest PWM score across the genome (computed and updated internally)

profiles: Object of class "GRList": Contains GRanges with sites above threshold and associated metrics (PWMscore and Occupancy) - Computed Internally

DNASequenceLength: Object of class "vector": Length of the Genome (or accesible genome) - computed internally

averageExpPWMScore: Object of class "vector": Average exponential PWM score across the genome (or accesible genome) - computed internally

ZeroBackground: Object of class "vector": Internal background value (computed internally)

drop: Object of class "vector": Stores Loci that do contain accesible DNA if it were to be the case (computed and updated internally)

tags: Object of class "character" ~Internal Tags~

ploidy: Object of class "numeric": A numeric Value describing the ploidy of the organism. Default: 2

boundMolecules: Object of class "vector": A vector (or single value) containing the number of bound Molecules (bound Transcription Factors): Default: 1000

backgroundSignal: Object of class "numeric": A numeric value describing the ChIP-seq background Signal (average signal from real ChIP seq data). Default: 0

maxSignal: Object of class "numeric": A numeric value describing the highest ChIP-seq signal (from real ChIP-seq data). Default: 1

lociWidth: Object of class "numeric" ~~

chipMean: Object of class "numeric": A numeric value describing the mean width of a ChIP- seq peak. Default:150

chipSd: Object of class "numeric": A numeric value describing the standard deviation of ChIP-seq peaks. Default: 150

chipSmooth: Object of class "vector": A numeric value describing the width of the window used to smooth Occupancy profiles into ChIP profiles. Default:250

stepSize: Object of class "numeric": A numeric value describing the step Size (in base pairs) between each ChIP-seq score. Default:10 (Scored every 10 base pairs)

removeBackground: Object of class "numeric": A numeric value describing the value at which score should be removed. Default:0 (If negative scores then remove)

noiseFilter: Object of class "character" ~Describes the noiseFilter method that will be applied to ChIP data (Zero, mean, median, sigmoid)~

PWMThreshold: Object of class "numeric": Threshold at which PWM Score should be selected (only sites above threshold will be selected - between 0 and 1)

strandRule: Object of class "character": "mean", "max" or "sum" will dertermine how strand should be handle for computing PWM Scores. Default : "max"

whichstrand: Object of class "character": "+", "-" or "+-" on which strand should PWM Score be computed. Default: "+-"

lambdaPWM: Object of class "vector" A vector (or single value) containing values for lambdaPWM Default: 1

naturalLog: Object of class "logical": A logical value describing if natural Log will be used to compute the PWM (if FALSE then log2 will be used). Default: TRUE

noOfSites: Object of class "numeric" A Positive integer describing number of sites (in base pair) should be used from the PFM to compute PWM. Default = 0 (Full width of binding site will be used when set to 0)

PWMPseudocount: Object of class "numeric": A numeric value describing a PWMPseudocount for PWM computation. Default: 1

paramTag: Object of class "character" ~Internal~

Extends

Class "[genomicProfilesInternal](#)", directly. Class "[parameterOptions](#)", directly.

Methods

initialize signature(.Object = "genomicProfiles"): ...

show signature(object = "genomicProfiles"): ...

Author(s)

Partick C. N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

See Also

[genomicProfiles](#) [parameterOptions](#)

Examples

```
showClass("genomicProfiles")
```

```
genomicProfilesInternal-class
      Class "genomicProfilesInternal"
```

Description

Non exported class. Represents the stripped down version of genomicProfiles.

Objects from the Class

Created Internally.

Slots

```
PWM: Object of class "matrix" ~~
PFM: Object of class "matrix" ~~
PFMFormat: Object of class "character" ~~
BPFfrequency: Object of class "vector" ~~
minPWMScore: Object of class "vector" ~~
maxPWMScore: Object of class "vector" ~~
profiles: Object of class "GRLIST" ~~
DNASequenceLength: Object of class "vector" ~~
averageExpPWMScore: Object of class "vector" ~~
ZeroBackground: Object of class "vector" ~~
drop: Object of class "vector" ~~
tags: Object of class "character" ~~
```

Methods

```
.averageExpPWMScore<- signature(object = "genomicProfilesInternal", value = "numeric"):
...
.DNASequenceLength<- signature(object = "genomicProfilesInternal", value = "vector"):
...
.drop<- signature(object = "genomicProfilesInternal", value = "vector"): ...
.generatePWM signature(object = "genomicProfilesInternal"): ...
.maxPWMScore<- signature(object = "genomicProfilesInternal", value = "vector"): ...
.minPWMScore<- signature(object = "genomicProfilesInternal", value = "vector"): ...
.profiles<- signature(object = "genomicProfilesInternal", value = "GRLIST"): ...
.tags signature(object = "genomicProfilesInternal"): ...
.tags<- signature(object = "genomicProfilesInternal", value = "character"): ...
averageExpPWMScore signature(object = "genomicProfilesInternal"): ...
```

```

BPFfrequency signature(object = "genomicProfilesInternal"): ...
BPFfrequency<- signature(object = "genomicProfilesInternal", value = "DNAStrngSet"):
  ...
BPFfrequency<- signature(object = "genomicProfilesInternal", value = "vector"): ...
DNASequenceLength signature(object = "genomicProfilesInternal"): ...
drop signature(object = "genomicProfilesInternal"): ...
maxPWMScore signature(object = "genomicProfilesInternal"): ...
minPWMScore signature(object = "genomicProfilesInternal"): ...
PFMFormat signature(object = "genomicProfilesInternal"): ...
PFMFormat<- signature(object = "genomicProfilesInternal", value = "character"): ...
PositionFrequencyMatrix signature(object = "genomicProfilesInternal"): ...
PositionFrequencyMatrix<- signature(object = "genomicProfilesInternal", value = "character"):
  ...
PositionFrequencyMatrix<- signature(object = "genomicProfilesInternal", value = "matrix"):
  ...
PositionWeightMatrix signature(object = "genomicProfilesInternal"): ...
PositionWeightMatrix<- signature(object = "genomicProfilesInternal", value = "matrix"):
  ...
profiles signature(object = "genomicProfilesInternal"): ...

```

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

See Also

[genomicProfiles](#)
[parameterOptions](#)

Examples

```
showClass("genomicProfilesInternal")
```

`getHighestFitnessSolutions`*Get Highest Fitness Solutions*

Description

`getHighestFitnessSolutions` extract best solution from a ChIPAnalyser GA/evolve Run.

Usage

```
getHighestFitnessSolutions(population,child=2,method="geometric")
```

Arguments

<code>population</code>	Population list as output by the evolve function.
<code>child</code>	numeric describing the number of solution to be extracted from Population list.
<code>method</code>	character string describing which scoring method should be used and selected from "geometric", "ks", "MSE", "pearson", "spearman", "kendall", "recall", "precesion", "fscore", "MCC", "AUC" or "AUC".

Details

This function only serves as a way of extracting data from the poppulation list. Ultimately - it is just a wrapper for some indexing.

Value

Return the index of the top "child" solutions.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

Examples

```
library(ChIPAnalyser)
data(ChIPAnalyserData)
# See GA vignette for usage
```

getTestingData	<i>Extract testing data from ChIPscore object</i>
----------------	---

Description

getTestingData extracts selected regions from ChIPscore object to be used as testing set.

Usage

```
getTestingData(ChIPscore, loci = 1)
```

Arguments

ChIPscore	ChIPscore object as returned by processingChIP
loci	numeric describing index of loci to be used as testing data.

Value

Returns ChIPscore object with the selected testing loci.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

Examples

```
library(ChIPAnalyser)
data(ChIPAnalyserData)
# See GA vignette for usage
test <- processingChIP(chip, top)
test <- getTestingData(test, 1:2)
```

getTrainingData	<i>Extract training data from ChIPscore object</i>
-----------------	--

Description

getTrainingData extracts selected regions from ChIPScore object to be used as training set.

Usage

```
getTrainingData(ChIPscore, loci = 1)
```

Arguments

ChIPscore ChIPscore object as returned by processingChIP
 loci numeric describing index of loci to be used as training data.

Value

Returns ChIPscore object with the selected training loci.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

Examples

```
library(ChIPAnalyser)
data(ChIPAnalyserData)
# See GA vignette for usage
test <- processingChIP(chip, top)
test <- getTrainingData(test, 1:2)
```

GRList-class

Class "GRList"

Description

Virtual Class to handle multiple data types for one slot ([profiles](#))

Objects from the Class

A virtual Class: No objects may be created from it.

Methods

[GRList-class](#) The purpose of this virtual classe is to store data of two different formats in one slot: GRangesList and Lists

Author(s)

Patrick C. N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. Nucleic Acids Res., 43, 84–94.

Examples

```
showClass("GRList")
```

initialize-methods	~~ <i>Methods for Function initialize</i> ~~
--------------------	--

Description

~~ Methods for function initialize ~~

Methods:

signature(.Object = "ChIPScore") Initialize ChIPScore

signature(.Object = "genomicProfiles") Initialize genomicProfiles

signature(.Object = "parameterOptions") Initialize parameterOptions

lambdaPWM	<i>Accessor Method for the lambdaPWM slot in a parameterOptions object</i>
-----------	--

Description

Accessor Method for the lambdaPWM slot in a [parameterOptions](#) object

Usage

```
lambdaPWM(object)
```

Arguments

object object is [parameterOptions](#) object

Details

The model underlying ChIPanalyser internally infers two paramters: number of bound molecules and lambda. Lambda represents a scaling factor for the Position weight matrix (PWM). This can be described as how well does a TF discriminate between high affinity and very high affinity sites.

Value

Returns the value assigned to the lambdaPWM slot in a [parameterOptions](#) object.

Author(s)

Patrick C. N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. Nucleic Acids Res., 43, 84–94.

Examples

```
# Loading data
data(ChIPanalyserData)

#Building data objects
GPP <- parameterOptions(lambdaPWM=1)
#Setting new Value for lambdaPWM
lambdaPWM(GPP)
```

lambdaPWM-methods *~~ Methods for Function lambdaPWM ~~*

Description

~~ Methods for function lambdaPWM ~~

Methods:

lambdaPWM(object)

lambdaPWM<- *Setter Method for the lambdaPWM slot in a [parameterOptions](#) object*

Description

Setter Method for the lambdaPWM slot in a [parameterOptions](#) object

Usage

```
lambdaPWM(object)<-value
```

Arguments

object	object is parameterOptions object
value	value is the numeric value to be assigned to the lambdaPWM slot. Default set at 1.

Details

The model underlying ChIPanalyser internally infers two parameters: number of bound molecules and lambda. Lambda represents a scaling factor for the Position weight matrix (PWM). This can be described as how well does a TF discriminate between high affinity and very high affinity sites.

Value

Returns the value assigned to the lambdaPWM slot in a [parameterOptions](#) object.

Author(s)

Patrick C. N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Examples

```
# Loading data
data(ChIPAnalyserData)

#Building data objects
GPP <- parameterOptions(lambdaPWM=1)
#Setting new Value for lambdaPWM
lambdaPWM(GPP) <- 2
```

lambdaPWM<-methods	~~ <i>Methods for Function lambdaPWM<-</i> ~~
--------------------	--

Description

Setter method for the lambdaPWM slot in the parameterOptions

Methods:

```
lambdaPWM(object)<-value
```

loci	<i>Accessor Method for the loci slot in a ChIPScore object</i>
------	--

Description

Setter Method for the loci slot in a [ChIPScore](#) object

Usage

```
loci(object)
```

Arguments

object object is [ChIPScore](#) object

Details

When using the [processingChIP](#), this functions will return a name GRanges with the loci of interest. These loci will either result from user input or extracted from the ChIP profiles (see [processingChIP](#) and [lociWidth](#)). This functions enables you to extract those loci from the [ChIPScore](#) object.

Value

Returns the value assigned to the loci slot in a [ChIPScore](#) object.

Author(s)

Patrick C. N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. Nucleic Acids Res., 43, 84–94.

Examples

```
# Loading data
data(ChIPAnalyserData)

chip<-processingChIP(chip,top)
loci(chip)
```

loci-class	Class "loci"
------------	--------------

Description

Setter for Loci of interest parsed to or extracted from the ChIPScore object

Objects from the Class

A virtual Class: No objects may be created from it.

Methods

.loci<- signature(object = "ChIPScore", value = "loci"): ...

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. Nucleic Acids Res., 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. CSBJ, 18, 3590-3605.

See Also

[ChIPScore](#)

Examples

```
showClass("loci")
```

loci-methods	<i>~~ Methods for Function loci ~~</i>
--------------	--

Description

Accessor method for the loci slot in [ChIPScore](#)

Methods:

loci{Object} Loci of interest parsed to or extracted from the ChIPScore object

lociWidth	<i>Accessor Method for the lociWidth slot in a parameterOptions object</i>
-----------	--

Description

Setter Method for the lociWidth slot in a [parameterOptions](#) object

Usage

```
lociWidth(object)
```

Arguments

object object is [parameterOptions](#) object

Details

When using the [processingChIP](#) function, the provided ChIP scores will be split into bins of a given size. lociWidth determines the Size of that bin. Default is set at 20 000 bp. This means that the ChIP profiles provided will be split into bins of 20 000 bp over the entire profile provided if no loci of interest is provided.

Value

Returns the value assigned to the lociWidth slot in a [parameterOptions](#) object.

Author(s)

Patrick C. N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. Nucleic Acids Res., 43, 84–94.

Examples

```
# Loading data
data(ChIPAnalyserData)

#Building data objects
GPP <- parameterOptions(lociWidth=20000)
#Accessing new Value for lociWidth
lociWidth(GPP)
```

lociWidth-methods

~~ *Methods for Function lociWidth* ~~

Description

Accessor method for the loci slot in [ChIPScore](#)

Methods:

`lociWidth(object)` Setting width of regions when using the reduce argument and NOT providing your own loci when using the [processingChIP](#) function.

lociWidth<-	<i>Setter Method for the lociWidth slot in a parameterOptions object</i>
-------------	--

Description

Setter Method for the lociWidth slot in a [parameterOptions](#) object

Usage

```
lociWidth(object)<-value
```

Arguments

object	object is parameterOptions object
value	value is the numeric value to be assigned to the lociWidth slot. Default set at 1.

Details

When using the [processingChIP](#) function, the provided ChIP scores will be split into bins of a given size. lociWidth determines the Size of that bin. Default is set at 20 000 bp. This mean that the ChIP profiles provided will be split into bins of 20 000 bp over the entire profile provided if no loci of interest is provided.

Value

Returns the value assigned to the lociWidth slot in a [parameterOptions](#) object.

Author(s)

Patrick C. N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. Nucleic Acids Res., 43, 84–94.

Examples

```
# Loading data
data(ChIPanalyserData)

#Building data objects
GPP <- parameterOptions(lociWidth=20000)
#Setting new Value for lociWidth
lociWidth(GPP) <- 30000
```

```
lociWidth<-methods      ~~ Methods for Function lociWidth<- ~~
```

Description

Setter method for the loci slot in [ChIPScore](#)

Methods:

```
lociWidth(Object)<-value
```

```
maxPWMScore      Accessor function for maxPWMScore slot in a genomicProfiles ob-
                  ject.
```

Description

Accessor function for maxPWMScore slot in a [genomicProfiles](#) object.

Usage

```
maxPWMScore(object)
```

Arguments

object object is a [genomicProfiles](#) object.

Details

maxPWMScore is a numerical value that can be described as the highest PWM score computed at a genome wide scale. This value is computed and updated in the [genomicProfiles](#) object after using the [computeGenomeWideScores](#).

Value

Returns the value of assigned to the maxPWMScore slot in a [genomicProfiles](#) object.

Author(s)

Patrick C. N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Examples

```
# Loading data
#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
#Building data objects
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR")

# Computing Genome Wide
GenomeWide <- computeGenomeWideScores(DNASequenceSet = DNASequenceSet,
  genomicProfiles = GPP)
maxPWMScore(GenomeWide)
## If used before computeGenomeWidePWMScore, will return NULL
```

maxPWMScore-methods *~~ Methods for Function maxPWMScore ~~*

Description

Accessor method for maxPWMScore

Methods:

maxPWMScore(object)

maxSignal	Accessor method for the maxSignal slot in a parameterOptions object.
-----------	--

Description

Accessor method for the maxSignal slot in a [parameterOptions](#) object.

Usage

maxSignal(object)

Arguments

object object is a `parameterOptions` object.

Details

In the context of ChIPanalyzer, maxSignal represents the maximum normalised ChIP-Seq signal of a given Transcription factor (or DNA binding protein). Although, A default value of 1 has been assigned to this slot, we strongly recommend to tailor this value accordingly. We strongly encourage to set values when building a `parameterOptions` object.

Value

Returns the value assigned to the maxSignal slot in a `parameterOptions` object.

Author(s)

Patrick C.N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
#Setting new Value for maxSignal
maxSignal(OPP)
```

maxSignal-methods *~~ Methods for Function maxSignal ~~*

Description

Accessor method for maxSignal

Methods:

maxSignal(object) Maximum ChIP signal extracted from ChIP data (see processingChIP)

maxSignal<-	Setter method for maxSignal slot in a parameterOptions object.
-------------	--

Description

Setter method for maxSignal slot in a [parameterOptions](#) object.

Usage

```
maxSignal(object) <- value
```

Arguments

object	object is a parameterOptions object.
value	value is a numerical value to be assigned to the maxSignal slot.

Details

In the context of ChIPAnalyser, maxSignal represents the maximum normalised ChIP-Seq signal of a given Transcription factor (or DNA binding protein). Although, A default value of 1 has been assigned to this slot, we strongly recommend to tailor this value accordingly. We strongly encourage to set values when building a [parameterOptions](#) object.

Value

Returns a [parameterOptions](#) with an updated value for maxSignal.

Author(s)

Patrick C.N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. Nucleic Acids Res., 43, 84–94.

Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
#Setting new Value for maxSignal
maxSignal(OPP) <- 1.8
```

```
maxSignal<-methods      ~~ Methods for Function maxSignal<- ~~
```

Description

Setter method for maxSignal

Methods:

maxSignal(Object)<-value Maximum ChIP signal extracted from ChIP data (see processingChIP)

```
minPWMScore            Accessor method the minPWMScore slot in a genomicProfiles object
```

Description

Accessor method the minPWMScore slot in a [genomicProfiles](#) object

Usage

```
minPWMScore(object)
```

Arguments

object object is a [genomicProfiles](#) object.

Details

minPWMScore can be described as the lowest PWM score computed at a genome wide scale. Although it is possible to assign a value to minPWMScore, we strongly advise to use the value computed and assigned internally. This value is computed in the [computeGenomeWideScores](#) function.

Value

Returns the value assigned to the minPWMScore slot in a [genomicProfiles](#) object.

Author(s)

Patrick C. N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Examples

```
#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
#Building data objects
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR")

# Computing Genome Wide
GenomceWide <- computeGenomeWideScores(DNASequenceSet = DNASequenceSet,
  genomicProfiles = GPP)
minPWMScore(GenomceWide)

## If used before computeGenomeWidePWMScore, will return NULL
```

minPWMScore-methods *~~ Methods for Function minPWMScore ~~*

Description

Accessor for minPWMScore

Methods:

minPWMScore(object) Minimum PWM score computed during the computeGenomeWideScores step.

naturalLog *Accessor method the naturalLog slot in a [parameterOptions](#) object.*

Description

Accessor method the naturalLog slot in a [parameterOptions](#) object.

Usage

```
naturalLog(object)
```

Arguments

object object is [parameterOptions](#) object.

Details

During the computation of a Position Weight Matrix, the Position Probability Matrix (derived from a Position Frequency Matrix) is log transformed. This parameter provides which "log transform" will be used. If TRUE, the Natural Log will be used (ln). If FALSE, log2 will be used. We strongly encourage to set values when building a [parameterOptions](#) object.

Value

Returns the value assigned to the naturalLog slot in a [parameterOptions](#) object.

Author(s)

Patrick C.N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Loading data
data(ChIPAnalyserData)

#Building data objects
GPP <- parameterOptions(naturalLog=TRUE)
#Setting new Value for naturalLog
naturalLog(GPP)
```

naturalLog-methods *~~ Methods for Function naturalLog ~~*

Description

Accessor method for the naturalLog slot in a [parameterOptions](#) object.

Methods:

naturalLog(object)

naturalLog<- *Setter method for the naturalLog slot in a [parameterOptions](#) object.*

Description

Setter method for the naturalLog slot in a [parameterOptions](#) object.

Usage

```
naturalLog(object)<- value
```

Arguments

object	object is a parameterOptions object.
value	value is a logical value that will determine if the natural log or log2 should be used for the computation of the Position Weight Matrix.

Details

During the computation of a Position Weight Matrix, the Position Probability Matrix (derived from a Position Frequency Matrix) is log transformed. This parameter provides which "log transform" will be used. If TRUE, the Natural Log will be used (ln). If FALSE, log2 will be used. We strongly encourage to set values when building a [parameterOptions](#) object.

Value

Returns [parameterOptions](#) object with an updated value for the naturalLog slot.

Author(s)

Patrick C.N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Loading data
data(ChIPAnalyserData)

#Building data objects
OPP <- parameterOptions(naturalLog=TRUE)
#Setting new Value for naturalLog
naturalLog(OPP) <- FALSE
```

```
naturalLog<-methods      ~~ Methods for Function naturalLog<- ~~
```

Description

Setter method for the naturalLog slot in a [parameterOptions](#) object.

Methods:

```
naturalLog(object)<-value
```

noiseFilter	<i>Accessor Method for the noiseFilter slot in a parameterOptions object</i>
-------------	--

Description

Accessor Method for the noiseFilter slot in a [parameterOptions](#) object

Usage

```
noiseFilter(object)
```

Arguments

object object is [parameterOptions](#) object

Details

Noise filtering method that should be used on ChIP-seq data. Four methods are available: Zero, Mean, Median and Sigmoid. Zero removes all ChIP-seq scores below zero, mean under the mean score, median under median score and sigmoid assigns a weight to each score based on a logistic regression curve. Mid point is set at 95 95 quantile of ChIP-seq scores. Below midpoint will receive a score between 0 and 1 , everything above will receive a score between 1 and 2

Value

Returns the value assigned to the noiseFilter slot in a [parameterOptions](#) object.

Author(s)

Patrick C. N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Examples

```
# Loading data
data(ChIPAnalyserData)

#Building data objects
GPP <- parameterOptions(noiseFilter="sigmoid")
#Setting new Value for noiseFilter
noiseFilter(GPP)
```

noiseFilter-methods *~~ Methods for Function noiseFilter ~~*

Description

Accessor method for noiseFilter

Methods:

noiseFilter(object) Noise Filter that will be applied to ChIP scores

noiseFilter<- *Setter Method for the noiseFilter slot in a [parameterOptions](#) object*

Description

Setter Method for the noiseFilter slot in a [parameterOptions](#) object

Usage

```
noiseFilter(object) <- value
```

Arguments

object	object is parameterOptions object
value	value is the value to be assigned to the noiseFilter slot (zero - mean - median -sigmoid)

Details

Noise filtering method that should be used on ChIP-seq data. Four methods are available: Zero, Mean, Median and Sigmoid. Zero removes all ChIP-seq scores below zero, mean under the mean score, median under median score and sigmoid assigns a weight to each score based on a logistic regression curve. Mid point is set at 95 95 quantile of ChIP-seq scores. Below midpoint will receive a score between 0 and 1 , everything above will receive a score between 1 and 2

Value

Returns the value assigned to the noiseFilter slot in a [parameterOptions](#) object.

Author(s)

Patrick C. N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. Nucleic Acids Res., 43, 84–94.

Examples

```
# Loading data
data(ChIPAnalyserData)

#Building data objects
GPP <- parameterOptions(noiseFilter="sigmoid")
#Setting new Value for noiseFilter
noiseFilter(GPP) <-"zero"
```

noiseFilter<-methods	~~ <i>Methods for Function</i> noiseFilter<- ~~
----------------------	---

Description

Setter method for noiseFilter

Methods:

noiseFilter(object)<-value Noise Filter that will be applied to ChIP scores

noOfSites	<i>Accessor Method for the noOfSites slot in a parameterOptions object</i>
-----------	--

Description

Accessor Method for the noOfSites slot in a [parameterOptions](#) object

Usage

```
noOfSites(object)
```

Arguments

object object is [parameterOptions](#) object

Details

While computing Position Weight Matrices (PWM) from Position Frequency Matrices (PFM), it is possible to restrict the number of sites that will be used to compute the PWM. The default is set at "all". In this case, all sites will be used to compute the PWM.

Value

Returns the value assigned to the noOfSites slot in a [parameterOptions](#) object.

Author(s)

Patrick C. N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Examples

```
# Loading data
data(ChIPAnalyserData)

#Building data objects
GPP <- parameterOptions(noOfSites="all")
#Setting new Value for naturalLog
noOfSites(GPP)
```

noOfSites-methods *~~ Methods for Function noOfSites ~~*

Description

~~ Methods for function noOfSites ~~

Methods:

signature(object = "parameterOptions")

noOfSites<- *Setter Method for the noOfSites slot in a [parameterOptions](#) object.*

Description

Setter Method for the noOfSites slot in a [parameterOptions](#) object.

Usage

```
noOfSites(object) <- value
```

Arguments

object	object is a parameterOptions object.
value	value is a positive integer that will be assigned to the noOfSites slot.

Details

While computing Position Weight Matrices (PWM) from Position Frequency Matrices (PFM), it is possible to restrict the number of sites that will be used to compute the PWM. The default is set at "all". In this case, all sites will be used to compute the PWM.

Value

Returns a [parameterOptions](#) object with an updated value for the noOfSites slot.

Author(s)

Patrick C.N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Loading data
data(ChIPAnalyserData)

#Building data objects
GPP <- parameterOptions(noOfSites=0)
#Setting new Value for naturalLog
noOfSites(GPP) <- 8
```

```
noOfSites<-methods      ~~ Methods for Function noOfSites<- ~~
```

Description

Setter method for noOfSites

Methods:

```
noOfSites(object)<-"all"
noOfSites(object)<-value
```

```
nos-class              Class "nos"
```

Description

Virtual class to handle Number of Sites

Objects from the Class

A virtual Class: No objects may be created from it.

Methods

No methods defined with class "nos" in the signature.

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
showClass("nos")
```

parameterOptions	<i>parameter Options object</i>
------------------	---------------------------------

Description

parameterOptions is an object used to store and parse the various parameters needed throughout this analysis pipeline.

Usage

```
parameterOptions(ploidy = 2, boundMolecules = 1000, backgroundSignal = 0, maxSignal = 1, lociWidth = 20000)
```

Arguments

ploidy	ploidy : A numeric Value describing the ploidy of the organism. Default: 2
boundMolecules	boundMolecules : A vector (or single value) containing the number of bound Molecules (bound Transcription Factors): Default: 1000
backgroundSignal	backgroundSignal : A numeric value describing the ChIP-seq background Signal (average signal from real ChIP seq data). Default: 0
maxSignal	maxSignal : A numeric value describing the highest ChIP-seq signal (from real ChIP-seq data). Default: 1
lociWidth	lociWidth : A numeric value describing the width of the bins used to split ChIP profiles parsed to processingChIP . Default = 20000
chipMean	chipMean : A numeric value describing the mean width of a ChIP- seq peak: Default:200
chipSd	chipSd : A numeric value describing the standard deviation of ChIP-seq peaks. Default: 200
chipSmooth	chipSmooth : A numeric value describing the width of the window used to smooth Occupancy profiles into ChIP profiles. Default:250
stepSize	stepSize : A numeric value describing the step Size (in base pairs) between each ChIP-seq score. Default:10 (Scored every 10 base pairs)
removeBackground	removeBackground : A numeric value describing the value at which score should be removed. Default:0 (If negative scores then remove)
noiseFilter	noiseFilter : A character string of one of the following: Zero, Mean, Median, or Sigmoid. Noise filter that will be applied to the ChIP Score during the processingChIP step.
naturalLog	naturalLog : A logical value describing if natural Log will be used to compute the PWM (if FALSE then log2 will be used). Default: TRUE
noOfSites	noOfSites : A Positive integer describing number of sites (in base pair) should be used from the PFM to compute PWM. Default =0 (Full width of binding site will be used when set to 0)

PWMThreshold	PWMThreshold: Threshold at which PWM Score should be selected (only sites above threshold will be selected - between 0 and 1)
strandRule	strandRule: 'mean', 'max' or 'sum' will determine how strand should be handle for computing PWM Scores. Default : 'max'
whichstrand	whichstrand: '+', '-' or '+-' on which strand should PWM Score be computed. Default: '+-'
PWMpseudocount	PWMpseudocount: A numeric value describing a PWMpseudocount for PWM computation. Default:1
lambdaPWM	A vector (or single value) containing values for the ScalingFactorPWM (Also known as lambda).Default:1

Details

ChIPAnalyser requires a lot of parameters. parameterOptions was created with the intent of storing and parsing these numerous arguments to the different functions. All parameters in this object are optional although strongly recommend. Some parameters are extracted and updated from function along the pipeline e.g. maxSignal and backgroundSignal are extracted during the processingChIP step. These parameters will be automatically parsed. If you do not wish to use them (or any other parameter) simply parse a new parameterOptions object with your desired parameters.

Value

Returns a parameterOptions with updated values.

Author(s)

Patrick C. N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. Nucleic Acids Res., 43, 84–94.

See Also

[genomicProfiles](#)

Examples

```
#
parameterOptions(ploidy = 2, boundMolecules = 1000, backgroundSignal = 0,
  maxSignal = 1, lociWidth = 20000, chipMean = 200, chipSd = 200,
  chipSmooth = 250, stepSize = 10, removeBackground = 0, noiseFilter = "zero",
  naturalLog = TRUE, noOfSites = "all", PWMThreshold = 0.7,
  strandRule = "max", whichstrand = "+-", PWMpseudocount = 1,
  lambdaPWM = 1)
```

parameterOptions-class

Class "parameterOptions"

Description

parameterOptions is an object used to store and parse the various parameters needed throughout this analysis pipeline.

Objects from the Class

Objects can be created by calls of the form parameterOptions(ploidy, boundMolecules, backgroundSignal, maxSignal, lociWidth, chipMean, chipSd, chipSmooth, stepSize, noiseFilter, removeBackground, lambdaPWM, PWMpseudocount, naturalLog, noOfSites, PWMThreshold, strandRule, whichstrand).

Slots

ploidy: Object of class "numeric": A numeric Value describing the ploidy of the organism. Default: 2

boundMolecules: Object of class "vector": A vector (or single value) containing the number of bound Molecules (bound Transcription Factors): Default: 1000

backgroundSignal: Object of class "numeric": A numeric value describing the ChIP-seq background Signal (average signal from real ChIP seq data). Default: 0

maxSignal: Object of class "numeric": A numeric value describing the highest ChIP-seq signal (from real ChIP-seq data). Default: 1

lociWidth: Object of class "numeric": A numeric value describing bin size when splitting ChIP seq scores). Default: 20 000

chipMean: Object of class "numeric": A numeric value describing the mean width of a ChIP- seq peak. Default:150

chipSd: Object of class "numeric": A numeric value describing the standard deviation of ChIP-seq peaks. Default: 150

chipSmooth: Object of class "vector": A numeric value describing the width of the window used to smooth Occupancy profiles into ChIP profiles. Default:250

stepSize: Object of class "numeric": A numeric value describing the step Size (in base pairs) between each ChIP-seq score. Default:10 (Scored every 10 base pairs)

removeBackground: Object of class "numeric": A numeric value describing the value at which score should be removed. Default:0 (If negative scores then remove)

noiseFilter: Object of class "character" Describes noiseFilter method applied to ChIP scores

PWMThreshold: Object of class "numeric": Threshold at which PWM Score should be selected (only sites above threshold will be selected - between 0 and 1)

strandRule: Object of class "character": "mean", "max" or "sum" will determine how strand should be handle for computing PWM Scores. Default : "max"

whichstrand: Object of class "character": "+", "-" or "+-" on which strand should PWM Score be computed. Default: "+-"

lambdaPWM: Object of class "vector" A vector (or single value) containing values for lambdaPWM Default: 1

naturalLog: Object of class "logical": A logical value describing if natural Log will be used to compute the PWM (if FALSE then log2 will be used). Default: TRUE

noOfSites: Object of class "nos" A Positive integer describing number of sites (in base pair) should be used from the PFM to compute PWM. Default = 0 (Full width of binding site will be used when set to 0)

PWMpseudocount: Object of class "numeric": A numeric value describing a PWMpseudocount for PWM computation. Default: 1

paramTag: Object of class "character" ~Internal~

Methods

.paramTag signature(object = "parameterOptions"): ...

.paramTag<- signature(object = "parameterOptions", value = "character"): ...

.ZeroBackground signature(object = "parameterOptions"): ...

.ZeroBackground<- signature(object = "parameterOptions", value = "vector"): ...

backgroundSignal signature(object = "parameterOptions"): ...

backgroundSignal<- signature(object = "parameterOptions", value = "numeric"): ...

boundMolecules signature(object = "parameterOptions"): ...

boundMolecules<- signature(object = "parameterOptions", value = "vector"): ...

chipMean signature(object = "parameterOptions"): ...

chipMean<- signature(object = "parameterOptions", value = "numeric"): ...

chipSd signature(object = "parameterOptions"): ...

chipSd<- signature(object = "parameterOptions", value = "numeric"): ...

chipSmooth signature(object = "parameterOptions"): ...

chipSmooth<- signature(object = "parameterOptions", value = "vector"): ...

initialize signature(.Object = "parameterOptions"): ...

lambdaPWM signature(object = "parameterOptions"): ...

lambdaPWM<- signature(object = "parameterOptions", value = "vector"): ...

lociWidth signature(object = "parameterOptions"): ...

lociWidth<- signature(object = "parameterOptions", value = "numeric"): ...

maxSignal signature(object = "parameterOptions"): ...

maxSignal<- signature(object = "parameterOptions", value = "numeric"): ...

naturalLog signature(object = "parameterOptions"): ...

naturalLog<- signature(object = "parameterOptions", value = "logical"): ...

noiseFilter signature(object = "parameterOptions"): ...

noiseFilter<- signature(object = "parameterOptions", value = "character"): ...

```

noOfSites signature(object = "parameterOptions"): ...
noOfSites<- signature(object = "parameterOptions", value = "character"): ...
noOfSites<- signature(object = "parameterOptions", value = "numeric"): ...
ploidy signature(object = "parameterOptions"): ...
ploidy<- signature(object = "parameterOptions", value = "numeric"): ...
PWMPseudocount signature(object = "parameterOptions"): ...
PWMPseudocount<- signature(object = "parameterOptions", value = "numeric"): ...
PWMThreshold signature(object = "parameterOptions"): ...
PWMThreshold<- signature(object = "parameterOptions", value = "numeric"): ...
removeBackground signature(object = "parameterOptions"): ...
removeBackground<- signature(object = "parameterOptions", value = "vector"): ...
show signature(object = "parameterOptions"): ...
stepSize signature(object = "parameterOptions"): ...
stepSize<- signature(object = "parameterOptions", value = "numeric"): ...
strandRule signature(object = "parameterOptions"): ...
strandRule<- signature(object = "parameterOptions", value = "character"): ...
whichstrand signature(object = "parameterOptions"): ...
whichstrand<- signature(object = "parameterOptions", value = "character"): ...

```

Author(s)

Partick C. N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

See Also

[genomicProfiles](#)

Examples

```
showClass("parameterOptions")
```

PFMFormat	<i>Accessor method for the PFMFormat slot in a genomicProfiles object</i>
-----------	---

Description

Accessor method for the PFMFormat slot in a [genomicProfiles](#) object

Usage

```
PFMFormat(object)
```

Arguments

object object is a [genomicProfiles](#) object

Details

If loading a [PositionFrequencyMatrix](#) from a file, the format of the file should be specified. Default is raw. Please keep in mind that this argument is used when parsing the [PositionFrequencyMatrix](#) file. IF this argument is changed after building the [genomicProfiles](#) with a PositionFrequency-Matrix file, this will not influence the parsing of the file. PFMFormat can be one of the following: "raw", "transfac", "JASPAR" or "sequences"

Value

Returns the value assigned to the PFMFormat slot a [genomicProfiles](#)

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Loading data
data(ChIPAnalyserData)
#Loading PFM files
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#Building data objects
#### THIS IS THE PREFERRED METHOD FOR SETTING PFMFormat
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR")
```

```
#Setting New value for PFMFormat
PFMFormat(GPP)
```

PFMFormat-methods *~~ Methods for Function PFMFormat ~~*

Description

Accesor method for the PFMFormat slot in a [genomicProfiles](#) object

Methods:

PFMFormat(object)

PFMFormat<- *Setter method for the PFMFormat slot in a [genomicProfiles](#) object*

Description

Setter method for the PFMFormat slot in a [genomicProfiles](#) object

Usage

```
PFMFormat(object) <- value
```

Arguments

object	object is a genomicProfiles object
value	value is character string of one of the following: "raw", "transfac", "JASPAR" or "sequences". If loading a PositionFrequencyMatrix from a file, the format of the file should specified. Default is JASPAR.

Details

If loading a [PositionFrequencyMatrix](#) from a file, the format of the file should be specified. Default is JASPAR. Please keep in mind that this argument is used when parsing the [PositionFrequencyMatrix](#) file. IF this argument is changed after building the [genomicProfiles](#) with a PositionFrequency-Matrix file, this will not influence the parsing of the file.

Value

Returns a [genomicProfiles](#) object with an updated value for the PFMFormat slot.

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Loading data
data(ChIPanalyserData)
#Loading PFM files
PFM <- file.path(system.file("extdata",package="ChIPanalyser"),"BEAF-32.pfm")
#Building data objects
#### THIS IS THE PREFERRED METHOD FOR SETTING PFMFormat
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR")
#Setting New value for PFMFormat
PFMFormat(GPP) <- "JASPAR"
```

PFMFormat<-methods *~~ Methods for Function PFMFormat<- ~~*

Description

Setter method for the PFMFormat slot in a [genomicProfiles](#) object

Methods:

PFMFormat(object)<-value

ploidy *Accessor method for the ploidy slot in a [parameterOptions](#) object*

Description

Accessor method for the ploidy slot in a [parameterOptions](#) object

Usage

ploidy(object)

Arguments

object object is a [parameterOptions](#) object

Details

Default value for ploidy is set a 2. It should be mentioned that ChIPanalyser is based on a model that also considers the ploidy of the organism of interest however this only considers simple polyploidy (or haploidy). The model does not consider hybrids such as wheat.

Value

Returns the value assigned to the ploidy slot in a `parameterOptions` object

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
#Setting new Value for maxSignal
ploidy(OPP)
```

ploidy-methods

~~ *Methods for Function ploidy* ~~

Description

Accessor method for the ploidy slot in a `parameterOptions` object

Methods:

`ploidy(object)`

ploidy<- *Setter Method for the ploidy slot in an [parameterOptions](#) object*

Description

Setter Method for the ploidy slot in an [parameterOptions](#) object

Usage

```
ploidy(object)<- value
```

Arguments

object	object is a parameterOptions object
value	value is a positive integer that describes the ploidy of the organism of interest.

Details

Default value for ploidy is set a 2. It should be mentioned that ChIPanalyser is based on a model that also considers the ploidy of the organism however this only considers simple polyploidy (or haploidy). The model does not consider hybrids such as wheat.

Value

Returns a [parameterOptions](#) object with an updated value for the ploidy slot.

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
#Setting new Value for maxSignal
ploidy(OPP) <- 2
```

ploidy<-methods *~~ Methods for Function ploidy<- ~~*

Description

Setter Method for the ploidy slot in an [parameterOptions](#) object

Methods:

ploidy(object)<-value

plotOccupancyProfile *Plot Occupancy Profiles*

Description

plotOccupancyProfile plots the predicted profiles. If provided, this functions will also plot ChIP-seq profiles, PWMScores (or Occupancy), chromatin States, Goodness of Fit estimates and gene information.

Usage

```
plotOccupancyProfile(predictedProfile, ChIPScore = NULL, chromatinState = NULL,
  occupancy = NULL, goodnessOfFit = NULL, PWM=FALSE,
  geneRef = NULL, addLegend = TRUE, ...)
```

Arguments

predictedProfile	predictedProfile is a either GRanges containing the predicted profiles for one loci, all loci selected for one paramter, or all loci selected for all parameter combinations selected. (see searchSites)
ChIPScore	ChIPScore is a ChIPscore object containing ChIPscore (or a list of numeric values representing ChIP scores (Experimental ChIP))
chromatinState	chromatinState is a GRanges containing accesible DNA sites or chromatin States.
occupancy	occupancy is a GRanges or a genomicProfiles object contaning PWM scores and Occupancy (see computeOccupancy)
goodnessOfFit	goodnessOfFit results of the profileAccuracyEstimate function.
PWM	PWM is a logical value that in the case occupancy is provided which of occupancy scores of PWM scores hould be plotted. Default set at FALSE
geneRef	geneRef is a GRanges containing gene information on exons,introns, UTR's, enhancers or any other genetic element to be plotted.

addLegend	addLegend is a logical value defining if the legend should be added. The legend will add all elements provided. See details.
...	Any other graphical Parameter of the following : cex, cex.lab, cex.main, densityCS, densityGR, ylab, xlab, main, colPred, colChIP, colOccup, colCS, colGR, n_axis_ticks. See details.

Details

Once the predicted ChIP-seq like profiles have been computed, it is possible to plot these profiles.

This functions allows to control graphical parameters. In short:

* col = color values - exact number of colors or colors that will be used in a colorRampPalette.

* cex = font sizes - for text, axis labels and main

* Density = fill density for chromatin state and/or geneRef blocks

Pred = predictedProfile ChIP = ChIP score (Experimental ChIP data) CS = Chromatin States GR = Gene reference Occup = Occupancy locations

Value

Returns a profile plot with "Occupancy" on the y axis and DNA position on the the X- axis.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
#Building data objects
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR", BPFfrequency=DNASequenceSet)
```

```

# Computing Genome Wide
GenomeWide <- computeGenomeWideScores(DNASequenceSet = DNASequenceSet,
  genomicProfiles = GPP)

#Compute PWM Scores
PWMScores <- computePWMScore(DNASequenceSet = DNASequenceSet,
  genomicProfiles = GenomeWide,
  loci = top, chromatinState = Access)
#Compute Occupancy
Occupancy <- computeOccupancy(genomicProfiles = PWMScores)

#Compute ChIP profiles
chipProfile <- computeChipProfile(loci = top,
  genomicProfiles = Occupancy)

#Plotting Profile
plotOccupancyProfile(predictedProfile=chipProfile,
  ChIPScore = chip,
  chromatinState = Access,
  occupancy = Occupancy,
  geneRef =geneRef)

plotOccupancyProfile(predictedProfile=chipProfile,
  ChIPScore = chip,
  chromatinState = Access,
  occupancy = Occupancy,
  geneRef = geneRef,
  colCS = c("red","blue"),
  densityGR = 60)

```

plotOptimalHeatMaps *Heat Map of optimal Parameters*

Description

plotOptimalHeatMaps will plot heat maps of optimal Parameters and highlight the optimal combination of [lambdaPWM](#) and [boundMolecules](#)

Usage

```
plotOptimalHeatMaps(optimalParam, contour=TRUE, col=NULL, main=NULL, layout=TRUE, overlay=FALSE)
```

Arguments

optimalParam	optimalParam is a list containing containing optimal matrices (or only one if only one parameter was selected). These matrices are the result of the computeOptimal function
contour	parameter is logical. Should contour lines be plotted?
col	col vector of colours to be used for each heat map. If none are specified, rainbow colours will be used. NOTE: colour vector will be recycled if not enough colours are provided.
main	main title.
layout	layout is either TRUE or FALSE specifying if standard layout should be used or not. If TRUE, each heat map will be plotted on an individual page with a heat map scale of the right side.
overlay	overlay is either TRUE or FALSE specifying if an overlay plot should be produced. The overlay plot takes the top 10 percent of best performing parameters per scoring metric and overlays them in a single plot. The resulting plots shows the optimal set of parameters for all metrics combined.

Details

Once the optimal set of Parameters ([lambdaPWM](#) and [boundMolecules](#)), it is possible to plot the results in the form of a heat map. Each heat map will be plotted in a separate page if layout = TRUE, If layout= FALSE, it is up to the user to define how they wish to layout their heat maps.

Value

Returns a heat map of optimal combinations of [lambdaPWM](#) and [boundMolecules](#). The x axis represents the different value assigned to lambda ([lambdaPWM](#)) and the y axis represents the different values to boundMolecules ([boundMolecules](#)).

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. Nucleic Acids Res., 43, 84–94.

Examples

```
#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata", package="ChIPAnalyser"), "BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
```

```

        install.packages("BiocManager")
        BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
    }
    library(BSgenome.Dmelanogaster.UCSC.dm6)
    DNASequenceset <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)

    #Building data objects
    GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR",BPFrequency=DNASequenceset)

    #Computing Optimal set of Parameters
    optimalParam <- computeOptimal(genomicProfiles = GPP,
        DNASequenceset = DNASequenceset,
        ChIPScore = chip,
        chromatinState = Access,
        parameterOptions = OPP,
        parameter = "all",
        peakMethod="moving_kernel")
    plotOptimalHeatMaps(optimalParam)

```

PositionFrequencyMatrix

Accessor method for the PFM slot in a genomicProfiles object

Description

Accessor method for the PFM slot in a genomicProfiles object

Usage

```
PositionFrequencyMatrix(object)
```

Arguments

object object is a [genomicProfiles](#) object

Details

After creating a [genomicProfiles](#) object, it is possible to access the Position Frequency Matrix slot. However this slot will be empty if the [genomicProfiles](#) object was built using directly a Position Weight Matrix. See [genomicProfiles](#)

Value

Returns the Position Frequency Matrix (PFM slot) used to compute the [PositionWeightMatrix](#) in a [genomicProfiles](#) object

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
#Loading data
data(ChIPAnalyserData)
#Loading PFM files
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#Building genomicProfiles object
GPP<-genomicProfiles(PFM=PFM,PFMFormat="JASPAR")
# Accessing Slot
PositionFrequencyMatrix(GPP)
```

PositionFrequencyMatrix-methods

~~ *Methods for Function PositionFrequencyMatrix* ~~

Description

Accessor method for the PFM slot in a `genomicProfiles` object

Methods:

`PositionFrequencyMatrix(object)`

`PositionFrequencyMatrix<-`

Setter method for the PFM slot in a `genomicProfiles` object

Description

Setter method for the PFM slot in a `genomicProfiles` object

Usage

`PositionFrequencyMatrix(object)<- value`

Arguments

<code>object</code>	<code>object</code> is a <code>genomicProfiles</code> object
<code>value</code>	<code>value</code> can be of two forms. Either a matrix in the form of a Position Frequency Matrix or a path/to/file character string.

Details

The Position Frequency Matrix is one of the fundamental object that needs to be supplied to a [genomicProfiles](#). If after building a [genomicProfiles](#), only the Position Frequency Matrix needs to be modified then it is possible to manually update the value of this matrix using the function above. There are two options for the type of data that may be supplied to the PFM slot: a matrix in the form of a Position Frequency Matrix (matrix with four rows - one for each base pair (ACTG) and a number of columns equal to the number of sites in the binding site), or it is possible (also recommended) to provide a path to the file containing the Position Frequency Matrix. This Position Frequency Matrix file may come in multiple form such as RAW, Transfac or JASPAR. WARNING: if a genomicProfiles object has already been created and only the PFM is supplied/updated , then the Positon Weight Matrix will automatically updated as well.

Value

Returns a [genomicProfiles](#) with an updated PFM slot (as described above this will lead to an updated PositionWeightMatrix).

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. Nucleic Acids Res., 43, 84–94.

Examples

```
#Loading data
data(ChIPAnalyserData)
#Loading PFM files
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#Building genomicProfiles object
# NOT ADVISED!!!! PLEASE PARSE PFM AND PFMFormat together
GPP<-genomicProfiles(PFMFormat = "JASPAR")
#Setting PFM
PositionFrequencyMatrix(GPP) <- PFM
```

PositionFrequencyMatrix<-methods

~~ *Methods for Function* PositionFrequencyMatrix<- ~~

Description

Setter method for the PFM slot in a [genomicProfiles](#) object

Methods:

```
PositionFrequencyMatrix(object)<-"path/to/file/"
PositionFrequencyMatrix(object)<-value
```

PositionWeightMatrix *Accessor Method for the PWM slot in a [genomicProfiles](#) object*

Description

Accessor Method for the PWM slot in a [genomicProfiles](#) object

Usage

```
PositionWeightMatrix(object)
```

Arguments

object object is a [genomicProfiles](#)

Details

After creating a [genomicProfiles](#) object, it is possible to access the Position Weight Matrix stored in this slot. This slot should always contain something. This slot is either supplied by user or directly computed from a Position Frequency Matrix when supplied.

Value

Returns a matrix in the form of a Position Weight Matrix

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
#Loading data
data(ChIPAnalyserData)
#Loading PFM files
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#Building genomicProfiles object
GPP<-genomicProfiles(PFM=PFM,PFMFormat="JASPAR")
# Accessing Slot
PositionWeightMatrix(GPP)
```

PositionWeightMatrix-methods

~~ Methods for Function PositionWeightMatrix ~~

Description

Accessor Method for the PWM slot in a [genomicProfiles](#) object

Methods:

PositionWeightMatrix(object)

PositionWeightMatrix<-

*Setter Method for the PositionWeightMatrix slot in a
[genomicProfiles](#) object*

Description

Setter Method for the PositionWeightMatrix slot in a [genomicProfiles](#) object

Usage

PositionWeightMatrix(object) <- value

Arguments

object object is a [genomicProfiles](#) object

value value is a matrix in the form of a Position Weight Matrix.

Details

If a Position Weight Matrix is readily available, it is possible to directly assign this matrix to the PWM slot. However, this is only possible if a [genomicProfiles](#) object has already been created. In that case, we advise to first create a [genomicProfiles](#) object. It should be noted that this Position Weight Matrix will be automatically computed from a Position Frequency Matrix. If no Position Frequency Matrix are available, then a Position Weight Matrix can be directly assigned to this slot.

Value

Returns a [genomicProfiles](#) object with an updated value for the PWM slot

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
#Building genomicProfiles object
GPP <- genomicProfiles()
#Setting PWM to PositionWeightMatrix slot
PWM <- matrix(runif(32,-10,20), ncol=8)
rownames(PWM) <- c("A","C","T","G")
PositionWeightMatrix(GPP) <- PWM
```

PositionWeightMatrix<-methods

~~ *Methods for Function* PositionWeightMatrix<- ~~

Description

Setter Method for the PositionWeightMatrix slot in a [genomicProfiles](#) object

Methods:

PositionWeightMatrix(object)<-value

processingChIP

Pre-processing ChIP-seq data

Description

processingChIP will process and extract ChIP scores at a set of loci of interest.

Usage

```
processingChIP(profile, loci=NULL, reduce=NULL,
               peaks=NULL, chromatinState=NULL, parameterOptions=NULL,
               cores=1)
```

Arguments

profile	profile is a path to a UCSC format file, a GRanges or data frame. The input data should contain 4 columns: chromosome, start, end and score.
loci	loci is GRanges describing the loci at which ChIP scores should be extracted. If NULL, a set of Loci will be extracted from profile. The data provided will then be split into bins of width equal to lociWidth (Default 20kbp) Default=NULL
reduce	reduce is a the top regions to select based on the mean ChIP score. If peaks are provided, regions overlapping with known peaks will be selected based on highest ChIP score. If NULL, all regions will be considered. Default=NULL
parameterOptions	parameterOptions is an parameterOptions object containing chip Parameters to be parsed for ChIP score extraction. If NULL, parameterOptions will be built internally with default ChIP extraction parameters (see chipSmooth , chipSd and chipMean) Default=NULL
peaks	peaks is a path to UCSC format file or a GRanges object containing location of ChIP peaks. Default=NULL
chromatinState	chromatinState is a GRanges containing Accessible DNA or chromatin States. If provided, regions will be selected only if they contain accessible DNA. Default=NULL
cores	cores is the number of cores used to extract ChIP scores. Default = 1

Details

When using [computeOptimal](#), it is required to supply real ChIP data in order to have a point of comparison. The correlation and MSE Scores are computed based on how well the model fits biological data. processingChIP will extract this data from ChIP data at loci of interest. When using the reduce option, this function will only select the top regions based on peak height or mean ChIP score. processingChIP will also extract maxSignal and backgroundSignal from ChIP data and parse it to an [parameterOptions](#) object.

Value

Returns a ChIPScore object containing extracted (and normalised) ChIP scores, the loci of interest and newly extracted Parameters(e.g. maxSignal)

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
#Data extraction
data(ChIPAnalyserData)

## Extracting ChIP scores at loci of interest

ChIP<-processingChIP(profile=chip, loci=top)
```

profileAccuracyEstimate

Estimating Accuracy of predicted Profiles

Description

profileAccuracyEstimate will compare the predicted ChIP-seq-like profile to real ChIP-seq data and return a set of metrics describing how accurate the predicted model is compared to real data.

Usage

```
profileAccuracyEstimate(genomicProfiles,ChIPScore,
  parameterOptions=NULL,method="all",cores=1)
```

Arguments

genomicProfiles	genomicProfiles is the result of computeChIPProfile
ChIPScore	ChIPScore is the result of processingChIP. Extracted/Normalised experimental ChIP scores.
parameterOptions	parameterOptions is a parameterOptions object for parameter specification.
method	method is the method that will be used to assess model quality against ChIP-seq data. Method can be one of the following: pearson, spearman, kendall, ks, geometric,fscore, MSE,or all.Fscore contains f-score, precision,recall, MCC, Accuracy and AUC ROC.
cores	cores is the number of cores used to extract ChIP scores. Default = 1

Details

In order to assess the quality of the model against experimental ChIP-seq data, ChIPAnalyser offers a wide range of method to choose from. These methods are also used when computing optimal parameters.

Value

Returns list of goodness of fit metrics for each loci and each parameter selected.

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
# Building genomicProfiles object
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR", BPFrequency=DNASequenceSet)

# Computing Genome Wide
GenomeWide <- computeGenomeWideScore(genomicProfiles = GPP,
  DNASequenceSet = DNASequenceSet)

#Compute PWM Scores
PWMScores <- computePWMScore(genomicProfiles = GenomeWide,
  DNASequenceSet = DNASequenceSet, loci = top, chromatinState = Access)
#Compute Occupancy
Occupancy <- computeOccupancy(genomicProfiles = PWMScores)

#Compute ChIP profiles
chipProfile <- computeChIPProfile(genomicProfiles=Occupancy,loci=top)
#Estimating accuracy estimate
AccuracyEstimate <- profileAccuracyEstimate(genomicProfiles = chipProfile,
  ChIPScore = chip,
  occupancyProfileParameters = OPP)
```

profiles-methods	~~ <i>Methods for Function profiles</i> ~~
------------------	--

Description

Accessor method for profiles in a [genomicProfiles](#) object

Methods:

`profiles(object)` Computed PWM scores, Occupancy or ChIP-seq like profiles for loci of interest and parameter combination of interest.

PWMPseudocount	<i>Accessor Method for a PWMPseudocount slot in a</i> parameterOptions
----------------	--

Description

Accessor Method for a PWMPseudocount slot in a [parameterOptions](#)

Usage

`PWMPseudocount(object)`

Arguments

`object` `object` is a [parameterOptions](#) object.

Details

In the context of Position Weight Matrices, the pseudocount is used to avoid 0 probabilities during the transformation of Position Frequency Matrix to a Position Probability Matrix and finally to a Position Weight Matrix. It is essentially a sample correction that is added in the case of small sample size. The effect of the base pair to which a pseudocount was assigned will not influence the model nor will create mathematical issues such as infinities or zero division. Default is set at 1.

Value

Returns the value assigned to a PWMPseudocount slot in a [parameterOptions](#) object

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Loading data
data(ChIPAnalyserData)

#Building data objects
GPP <- parameterOptions(PWMpseudocount=0)
#Accessing slot value
PWMpseudocount(GPP)
```

PWMpseudocount-methods

~~ *Methods for Function PWMpseudocount* ~~

Description

Accessor Method for a PWMpseudocount slot in a [parameterOptions](#)

Methods:

PWMpseudocount(object)

PWMpseudocount<-

Setter Method for the pseudocount slot in a [parameterOptions](#) object

Description

Setter Method for the pseudocount slot in a [parameterOptions](#) object

Usage

```
PWMpseudocount(object) <- value
```

Arguments

object	object is a parameterOptions object
value	value is a numeric value that will be assigned to the pseudocount slot. Default is set at 1

Details

In the context of Position Weight Matrices, the pseudocount is used to avoid 0 probabilities during the transformation of Position Frequency Matrix to a Position Probability Matrix and finally to a Position Weight Matrix. It is essentially a sample correction that is added in the case of small sample size. The effect of the base pair to which a pseudocount was assigned will not influence the model nor will create mathematical issues such as infinities or zero division.

Value

Returns a `parameterOptions` object with an updated value for the pseudocount slot.

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Loading data
data(ChIPAnalyserData)

#Building data objects
GPP <- parameterOptions( PWMpseudocount=0)
#Setting Value for new PWMpseudocount
PWMpseudocount(GPP) <- 1
```

PWMpseudocount<-methods

~~ *Methods for Function PWMpseudocount<-* ~~

Description

Setter Method for the pseudocount slot in a `parameterOptions` object

Methods:

`PWMpseudocount(object)<-value`

PWMThreshold	<i>Accessor method for the PWMThreshold slot in a parameterOptions object</i>
--------------	---

Description

Accessor method for the PWMThreshold slot in a [parameterOptions](#) object

Usage

```
PWMThreshold(object)
```

Arguments

object object is a [parameterOptions](#) object

Details

The computePWMScore function requires a so-called PWM Threshold. This threshold represents the Threshold at which PWM Score should be selected. The PWMThreshold is a positive numeric value (between 0 and 1. If set at 0, all sites will be selected. If set at 0.7 (Default value), then 70 % of PWM Score (and by extension binding sites) will be IGNORED. The top 30 % will be selected.

Value

Returns the value assigned to the PWMThreshold slot in a [parameterOptions](#) object

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Loading data
data(ChIPAnalyserData)

#Building data objects
GPP <- parameterOptions(PWMThreshold=0.7)
#Accessing Value for PWMThreshold
PWMThreshold(GPP)
```

PWMThreshold-methods *~~ Methods for Function PWMThreshold ~~*

Description

Accessor method for the PWMThreshold slot in a [parameterOptions](#) object

Methods:

PWMThreshold(object)

PWMThreshold<- *Setter Method for the PWMThreshold slot in a [parameterOptions](#) object*

Description

Setter Method for the PWMThreshold slot in a [parameterOptions](#) object

Usage

```
PWMThreshold(object) <- value
```

Arguments

object	object is a parameterOptions object
value	value is a numeric value (between 0 and 1) to be assigned to the PWMThreshold slot in parameterOptions object. Default is set at 0.7

Details

The computePWMScore function requires a so-called PWM Threshold. This threshold represents the Threshold at which PWM Score should be selected. The PWMThreshold is a positive numeric value (between 0 and 1). If set at 0, all sites will be selected. If set at 0.7 (Default value), then 70 % of PWM Score (and by extension binding sites) will be IGNORED. The top 30 % will be selected.

Value

Returns [parameterOptions](#) object with an updated value for the PWMThreshold slot

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Loading data
data(ChIPAnalyserData)

#Building data objects
GPP <- parameterOptions(PWMThreshold=0.7)
#Setting Value for new PWMThreshold
PWMThreshold(GPP) <- 0.8
```

PWMThreshold<-methods *~~ Methods for Function PWMThreshold<- ~~*

Description

Setter Method for the PWMThreshold slot in a [parameterOptions](#) object

Methods:

PWMThreshold(object)<-value

removeBackground	<i>Accessor Method for the removeBackground slot in a parameterOptions object</i>
------------------	---

Description

Accessor Method for the removeBackground slot in a [parameterOptions](#) object

Usage

```
removeBackground(object)
```

Arguments

object object is a [parameterOptions](#) object

Details

A numeric value describing a threshold at which Occupancy signals must be removed (Default is set at 0). The removal of Occupancy signals will occur when computing [computeOccupancy](#) (see [computeOccupancy](#) function)

Value

Returns the value assigned to the removeBackground slot in a [parameterOptions](#) object

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
#Building parameterOptions object
OPP <- parameterOptions()
#Accessing Value for removeBackground
removeBackground(OPP)
```

removeBackground-methods

~~ *Methods for Function removeBackground* ~~

Description

Accessor Method for the removeBackground slot in a [parameterOptions](#) object

Methods:

removeBackground(object)

removeBackground<- *Setter Method for the removeBackground slot in a parameterOptions object*

Description

Setter Method for the removeBackground slot in a parameterOptions object

Usage

removeBackground(object) <-value

Arguments

object	object is an parameterOptions object
value	value is positive numerical value to be assigned to the removeBackground slot in a parameterOptions object. Default is set a 0.

Details

A numeric value describing a threshold at which Occupancy signals must be removed (Default is set at 0). The removal of Occupancy signals will occur when computing [computeOccupancy](#) (see [computeOccupancy](#) function)

Value

Returns an [parameterOptions](#) object with an updated value for the removeBackground slot

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
#Building parameterOptions object
OPP <- parameterOptions()
#Setting new Value for removeBackground
removeBackground(OPP) <- 0.1
```

```
removeBackground<-methods
```

```
~~ Methods for Function removeBackground<- ~~
```

Description

Setter Method for the removeBackground slot in a parameterOptions object

Methods:

```
removeBackground(object)<-value
```

scores

Accessor Method for the scores slot in a [ChIPScore](#) object

Description

Setter Method for the scores slot in a [ChIPScore](#) object

Usage

```
scores(object)
```

Arguments

object object is [ChIPScore](#) object

Details

When using the [processingChIP](#), this functions will return a name list of normalised ChIP scores at loci of interest. This functions enables you to extract those scores from the [ChIPScore](#) object.

Value

Returns the value assigned to the scores slot in a [ChIPScore](#) object.

Author(s)

Patrick C. N. Martin <p.martin@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94.

Examples

```
# Loading data
data(ChIPAnalyserData)

chip<-processingChIP(chip,top)
str(scores(chip))
```

scores-methods	~~ <i>Methods for Function scores</i> ~~
----------------	--

Description

Accessor method for scores slot in a ChIPScore object.

Methods:

scores(object) Extracted and normalised ChIP scores at loci of interest.

searchSites	<i>Searching function for Sites above threshold and predicted ChIP-seq Profiles</i>
-------------	---

Description

searchSites is function enabling quick extraction and search for parameter combinations and/or loci in any [genomicProfiles](#) object from computeOccupancy onwards.

Usage

```
searchSites(Sites,lambdaPWM="all",BoundMolecules="all", Locus="all")
```

Arguments

Sites	Sites is either a genomicProfiles or the result of computeOptimal
lambdaPWM	lambdaPWM is a numeric vector describing the ScalingFactors that should be searched within Sites.
BoundMolecules	BoundMolecules is a numeric vector describing the BoundMolecules that should be searched within Sites.
Locus	Locus is a character vector describing the Loci that should be searched within Sites.

Details

When testing numerous combinations of lambdaPWM and boundMolecules on top of many loci, it can become challenging to navigate the large data output searchSites will make searching in this slot a lot easier. If all arguments are left at their default value of "all", then all Parameters will be searched thus returning the full list of Sites above threshold. If a value for lambdaPWM is user provided then only this lambdaPWM will be selected (all boundMolecules and loci will also be selected). searchSites also works on the result of computeOptimal.

Value

Returns object of same time as parsed to this function with only the parameters and/or loci selected.

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
#Data extraction
data(ChIPAnalyserData)
# path to Position Frequency Matrix
PFM <- file.path(system.file("extdata",package="ChIPAnalyser"),"BEAF-32.pfm")
#As an example of genome, this example will run on the Drosophila genome

if(!require("BSgenome.Dmelanogaster.UCSC.dm6", character.only = TRUE)){
  if (!requireNamespace("BiocManager", quietly=TRUE))
    install.packages("BiocManager")
  BiocManager::install("BSgenome.Dmelanogaster.UCSC.dm6")
}
library(BSgenome.Dmelanogaster.UCSC.dm6)
DNASequenceSet <- getSeq(BSgenome.Dmelanogaster.UCSC.dm6)
# Building genomicProfiles object
GPP <- genomicProfiles(PFM=PFM,PFMFormat="JASPAR", BPFrequency=DNASequenceSet)

# Computing Genome Wide
GenomeWide <- computeGenomeWideScore(genomicProfiles = GPP,
  DNASequenceSet = DNASequenceSet)

#Compute PWM Scores
PWMScores <- computePWMScore(genomicProfiles = GenomeWide,
  DNASequenceSet = DNASequenceSet, loci = top, chromatinState = Access)
#Compute Occupancy
Occupancy <- computeOccupancy(genomicProfiles = PWMScores)
searchSites(Occupancy,ScalingFactor=c(1,4), BoundMolecules = c(1,100),
  Locus="eve")

#Compute ChIP profiles
chipProfile <- computeChIPProfile(genomicProfiles=Occupancy,loci=top)
searchSites(chipProfile,ScalingFactor=c(1,4), BoundMolecules = c(1,100),
  Locus="eve")

optimalParam <- computeOptimal(genomicProfiles = GPP,
  DNASequenceSet = DNASequenceSet,
  ChIPScore = chip,
  chromatinState = Access,
  parameterOptions = OPP,
  parameter = "all",
```

```

peakMethod="moving_kernel")

searchSites(optimalParam,ScalingFactor=c(1,4), BoundMolecules = c(1,100),
  Locus="eve")

```

setChromatinStates	<i>setChromatinStates</i>
--------------------	---------------------------

Description

setChromatinStates sets chromatin state affinity values to a GRanges object.

Usage

```
setChromatinStates(population,chromatinStates)
```

Arguments

population	Population list containing all individuals and associated parameter. Must contain chromatin state affinity values. See generateStartingPopulation.
chromatinStates	GRanges object containing chromatin state locations.

Details

Chromatin states can be loaded into R as a GRanges object. Each range represents the extent of a certain chromatin state and the chromatin state type should be assigned to a meta data column called "name". The affinity values names should be set accordingly.

Value

Returns a GRange object with affinity scores for each chromatin state range. Affinity scores are placed in the DNAAffinity meta data column.

Author(s)

Patrick C.N. Martin

Examples

```

library(ChIPAnalyser)
# Input data
data(ChIPAnalyserData)

pop <- 10
params <- c("N","lambda","PWMThreshold", paste0("CS",seq(1:11)))
start_pop <- generateStartingPopulation(pop, params)

```

```
cs <- setChromatinStates(start_pop,cs)
```

show-methods	~~ <i>Methods for Function show</i> ~~
--------------	--

Description

Show methods for various objects

Methods:

```
signature(object = "ChIPScore")
signature(object = "genomicProfiles")
signature(object = "parameterOptions")
```

singleRun	<i>singleRun</i>
-----------	------------------

Description

singleRun runs ChIPanalyser after optimal paramters have been found by the evolve function.

Usage

```
singleRun(indiv,DNAAffinity,
          genomicProfiles,DNASequenceSet,
          ChIPScore,fitness="all")
```

Arguments

- | | |
|-----------------|---|
| indiv | Population list containing the top scoring individual. Note that this should be a list of length 1 containing another list with all parameter values. |
| DNAAffinity | GRanges object as outputed by the setchromatinStates. |
| genomicProfiles | genomicProfiles object containing PWM scores and other desired metrics. Note that PWMThreshold, lambda and N will be overwritten using values from indiv. |
| DNASequenceSet | DNA string set object containing DNA sequence of interest. |
| ChIPScore | ChIPScore object as outputed by the processingChIP function. |
| fitness | character string describing which metric should be used to assess fitness and should be one of the following:"geometric","ks","MSE","pearson","spearman","kendall","recall","precesion","fscore","MCC","Accuracy" or "AUC". |

Details

Once the genetic algorithm has been optimised, the top individual may be run on its own to get predicted ChIP profiles. The use of this function requires a few extract steps in order to predict ChIP profiles.

First, the index of the top individual should be extracted (see `getHighestFitnessSolutions`). Second, using this index, subset top individual from GA population. Note this should be done using `[]` single bracket notation as, a list of length 1 containing another list with all parameter values is required for the next steps. Yes, this is might seem annoying but the functions were design for list structures... Third, `setchromatinStates` using the top individual list. This will add chromatin affinity values to your chromatinState GRanges. Use this new chromatinState object as your new chromatinState object. Fourth, parse your indiv list object to `singleRun`.

Value

Return a list with three elements. First element contains a `genomicProfiles` object with occupancy scores. Second element contains a `genomicProfiles` object with ChIP profile scores. Third element contains a goodness of fit metrics.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

Examples

```
library(ChIPAnalyser)
data(ChIPAnalyserData)
# See GA vignette for usage
```

splitData

Get Training and Testing data from ChIPscore objects

Description

`splitData` splits processed ChIP data into training and testing sets.

Usage

```
splitData(ChIPscore, dist = c(80, 20), as.proportion = TRUE)
```

Arguments

ChIPscore	ChIPscore object as returned by <code>processingChIP</code>
dist	If <code>as.proportion</code> is to <code>TRUE</code> , split the data into desired proportions. Default sets 80% training and 20% testing. If <code>as.proportion</code> is to <code>FALSE</code> , a vector of 4 numeric values describing start and end of training and testing respectively.
as.proportion	Logical describing if values provided to <code>dist</code> should be treated as % of training and testing or if <code>dist</code> should be considered as start and end of loci selected for training and testing respectively.

Value

Returns a named list of ChIPScore objects

* trainingSet = ChIPscore containing training set * testingSet = ChIPscore containing testing set.

Author(s)

Patrick C.N. Martin <pcnmartin@gmail.com>

Examples

```
library(ChIPAnalyser)
data(ChIPAnalyserData)
# See GA vignette for usage
test <- processingChIP(chip,top)
usingDist <- splitData(test, dist = c(50,50),as.proportion = TRUE )
usingIndex <- splitData(test, dist = c(1,2,3,4),as.proportion = FALSE )
```

stepSize

Accessor method of the stepSize slot in [parameterOptions](#) object

Description

Accessor method of the stepSize slot in [parameterOptions](#) object

Usage

```
stepSize(object)
```

Arguments

object object is a [parameterOptions](#) object.

Details

It possible to restrict the size of the ChIP-seq-like profile produced by [computeChIPProfile](#). Instead of returning ChIP-seq like score for each base pair, it is possible to skip base pairs and only return the predicted enrichment score for every "n" base pair (n is the value assigned to stepSize). This will reduce the size of the output data (unless step size is very large, this will not affect the accuracy of the model). Default is set at 10 base pairs.

Value

Returns the value assigned to the stepSize slot in a [parameterOptions](#)

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
#Setting new Value for maxSignal
stepSize(OPP)
```

stepSize-methods *~~ Methods for Function stepSize ~~*

Description

Accessor method of the stepSize slot in [parameterOptions](#) object

Methods:

stepSize(object)

stepSize<- *Setter Method for the stepSize slot in a [parameterOptions](#)*

Description

Setter Method for the stepSize slot in a [parameterOptions](#)

Usage

```
stepSize(object) <- value
```

Arguments

object	object is a parameterOptions object
value	value is a positive numeric value that will be assigned to the stepSize slot in a parameterOptions object. Default is set at 10 base pairs.

Details

It possible to restrict the size of the ChIP-seq-like profile produced by [computeChIPProfile](#). Instead of returning ChIP-seq like score for each base pair, it is possible to skip base pairs and only return the predicted enrichment score for every "n" base pair (n is the value assigned to stepSize). This will reduce the size of the output data (unless step size is very large, this will not affect the accuracy of the model). Default is set at 10 base pairs.

Value

Returns a parameterOptions object with an updated value for the stepSize slot.

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Building parameterOptions object
OPP <- parameterOptions()
#Setting new Value for maxSignal
stepSize(OPP) <- 20
```

```
stepSize<-methods      ~~ Methods for Function stepSize<- ~~
```

Description

Setter Method for the stepSize slot in a [parameterOptions](#)

Methods:

```
stepSize(object)<-value
```

```
strandRule      Accessor Method for the strandRule slot in a parameterOptions
                  object
```

Description

Accessor Method for the strandRule slot in a [parameterOptions](#) object

Usage

```
strandRule(object)
```

Arguments

object object is a `parameterOptions` object

Details

When computing the PWM Scores and if `whichstrand` is set to "+-", `strandRule` will determine how to handle both strands (one of three options : "mean", "max", "sum"). If set to "mean", the average PWM Score of both strand will be computed. If set to "max", the highest PWM score between each strand will be selected and finally "sum" will sum both score together. Default set at "max"

Value

Returns the value assigned to `strandRule` slot (one of three options : "mean", "max", "sum") in a `parameterOptions` object

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Loading data
data(ChIPAnalyserData)

#Building data objects
GPP <- parameterOptions( strandRule="max")
#Accesssing Value for strandRule
strandRule(GPP)
```

strandRule-methods *~~ Methods for Function strandRule ~~*

Description

Accessor Method for the `strandRule` slot in a `parameterOptions` object

Methods:

`strandRule(object)`

strandRule<- *Setter method for the strandRule slot in a [parameterOptions](#) object.*

Description

Setter method for the strandRule slot in a [parameterOptions](#) object.

Usage

```
strandRule(object) <- value
```

Arguments

object	object is a parameterOptions object
value	value is a character string and can be one of the following 'mean', 'max', 'sum'. This will only apply if whichstrand is '+-'. Default set at 'max'

Details

When computing the PWM Scores and if [whichstrand](#) is set to '+-', strandRule will determine how to handle both strands (one of three options : 'mean', 'max', 'sum'). If set to 'mean', the average PWM Score of both strand will be computed. If set to 'max', the highest PWM score between each strand will be selected and finally 'sum' will sum both score together. Default set at 'max'

Value

Returns a [parameterOptions](#) object with an updated value for the strandRule slot

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Loading data
data(ChIPanalyserData)

#Building data objects
GPP <- parameterOptions(strandRule="max")
#Setting New Value for strandRule
strandRule(GPP) <- "mean"
```

```
strandRule<-methods    ~~ Methods for Function strandRule<- ~~
```

Description

Setter method for the strandRule slot in a [parameterOptions](#) object.

Methods:

```
strandRule(object)<-value
```

whichstrand	<i>Accessor method for the whichstrand slot in a parameterOptions object</i>
-------------	--

Description

Accessor method for the whichstrand slot in a [parameterOptions](#) object

Usage

```
whichstrand(object)
```

Arguments

object object is a [parameterOptions](#) object

Details

PWM Score may be computed on either the positive strand ("+"), the negative strand ("-") or on both strands ("+-").

Value

Returns on which strand PWM Scores should be computed (whichstrand in a [parameterOptions](#) object)

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Loading data
data(ChIPAnalyserData)

#Building data objects
GPP <- parameterOptions( whichstrand="+-")
#Setting New Value for whichstrand
whichstrand(GPP)
```

whichstrand-methods *~~ Methods for Function whichstrand ~~*

Description

Accessor method for the whichstrand slot in a [parameterOptions](#) object

Methods:

whichstrand(object)

whichstrand<- *Setter method for the whichstrand slot in a [parameterOptions](#) object*

Description

Setter method for the whichstrand slot in a [parameterOptions](#) object

Usage

```
whichstrand(object) <- value
```

Arguments

object	object is a parameterOptions object
value	value is a character string specifying which strand should be used to compute PWM Scores. The three available options are the following: "+", "-" or "+-". Default is "+-"

Details

PWM Score may be computed on either the positive strand ("+"), the negative strand ("-") or on both strands ("+-").

Value

Returns a [parameterOptions](#) object with an updated value for the whichstrand slot

Author(s)

Patrick C. N. Martin <pm16057@essex.ac.uk>

References

Zabet NR, Adryan B (2015) Estimating binding properties of transcription factors from genome-wide binding profiles. *Nucleic Acids Res.*, 43, 84–94. Patrick C.N. Martin and Nicolae Radu Zabe (2020) Dissecting the binding mechanisms of transcription factors to DNA using a statistical thermodynamics framework. *CSBJ*, 18, 3590-3605.

Examples

```
# Loading data
data(ChIPanalyserData)

#Building data objects
GPP <- parameterOptions( whichstrand="+-")
#Setting New Value for whichstrand
whichstrand(GPP) <- "+"
```

whichstrand<-methods *~~ Methods for Function whichstrand<- ~~*

Description

Setter method for the whichstrand slot in a [parameterOptions](#) object

Methods:

whichstrand(object)<-value

Index

* classes

- ChIPScore-class, [20](#)
- genomicProfiles-class, [43](#)
- genomicProfilesInternal-class, [46](#)
- GRList-class, [50](#)
- loci-class, [54](#)
- nos-class, [71](#)
- parameterOptions-class, [74](#)

* methods

- averageExpPWMScore-methods, [7](#)
- backgroundSignal-methods, [9](#)
- backgroundSignal<-methods, [10](#)
- boundMolecules-methods, [12](#)
- boundMolecules<-methods, [13](#)
- BPFrequency-methods, [14](#)
- BPFrequency<-methods, [16](#)
- chipMean-methods, [18](#)
- chipMean<-methods, [20](#)
- chipSd-methods, [23](#)
- chipSd<-methods, [24](#)
- chipSmooth-methods, [25](#)
- chipSmooth<-methods, [26](#)
- DNASequenceLength-methods, [37](#)
- drop-methods, [39](#)
- initialize-methods, [51](#)
- lambdaPWM-methods, [52](#)
- lambdaPWM<-methods, [53](#)
- loci-methods, [55](#)
- lociWidth-methods, [56](#)
- lociWidth<-methods, [58](#)
- maxPWMScore-methods, [59](#)
- maxSignal-methods, [60](#)
- maxSignal<-methods, [62](#)
- minPWMScore-methods, [63](#)
- naturalLog-methods, [64](#)
- naturalLog<-methods, [66](#)
- noiseFilter-methods, [67](#)
- noiseFilter<-methods, [68](#)
- noOfSites-methods, [69](#)

- noOfSites<-methods, [71](#)
- PFMFormat-methods, [78](#)
- PFMFormat<-methods, [79](#)
- ploidy-methods, [80](#)
- ploidy<-methods, [82](#)
- PositionFrequencyMatrix-methods, [87](#)
- PositionFrequencyMatrix<-methods, [88](#)
- PositionWeightMatrix-methods, [90](#)
- PositionWeightMatrix<-methods, [91](#)
- profiles-methods, [95](#)
- PWMPseudocount-methods, [96](#)
- PWMPseudocount<-methods, [97](#)
- PWMThreshold-methods, [99](#)
- PWMThreshold<-methods, [100](#)
- removeBackground-methods, [101](#)
- removeBackground<-methods, [102](#)
- scores-methods, [104](#)
- show-methods, [107](#)
- stepSize-methods, [110](#)
- stepSize<-methods, [111](#)
- strandRule-methods, [112](#)
- strandRule<-methods, [114](#)
- whichstrand-methods, [115](#)
- whichstrand<-methods, [116](#)

* package

- ChIPAnalyser-package, [5](#)
- .DNASequenceLength<- , genomicProfilesInternal, vector-method (genomicProfilesInternal-class), [46](#)
- .ZeroBackground, parameterOptions-method (parameterOptions-class), [74](#)
- .ZeroBackground<- , parameterOptions, vector-method (parameterOptions-class), [74](#)
- .averageExpPWMScore<- , genomicProfilesInternal, numeric-method (genomicProfilesInternal-class), [46](#)
- .drop<- , genomicProfilesInternal, vector-method

- (genomicProfilesInternal-class), 46
- .generatePWM, genomicProfilesInternal-method (genomicProfilesInternal-class), 46
- .maxPWMscore<-, genomicProfilesInternal, vector-method (genomicProfilesInternal-class), 46
- .minPWMscore<-, genomicProfilesInternal, vector-method (genomicProfilesInternal-class), 46
- .paramTag, parameterOptions-method (parameterOptions-class), 74
- .paramTag<-, parameterOptions, character-method (parameterOptions-class), 74
- .profiles<-, genomicProfilesInternal, GRList-method (genomicProfilesInternal-class), 46
- .tags, genomicProfilesInternal-method (genomicProfilesInternal-class), 46
- .tags<-, genomicProfilesInternal, character-method (genomicProfilesInternal-class), 46
- Access (ChIPAnalyserData), 16
- averageExpPWMscore, 6
- averageExpPWMscore, genomicProfilesInternal-method (genomicProfilesInternal-class), 46
- averageExpPWMscore-methods, 7
- backgroundSignal, 8, 72
- backgroundSignal, parameterOptions-method (parameterOptions-class), 74
- backgroundSignal-methods, 9
- backgroundSignal<-, 9
- backgroundSignal<-methods, 10
- backgroundSignal<-, parameterOptions, numeric-method (parameterOptions-class), 74
- boundMolecules, 11, 28, 32, 33, 72, 84, 85
- boundMolecules, parameterOptions-method (parameterOptions-class), 74
- boundMolecules-methods, 12
- boundMolecules<-, 12
- boundMolecules<-methods, 13
- boundMolecules<-, parameterOptions, vector-method (parameterOptions-class), 74
- BPFrequency, 13
- BPFrequency, genomicProfilesInternal-method (genomicProfilesInternal-class), 46
- BPFrequency-methods, 14
- BPFrequency<-, 15
- BPFrequency<-methods, 16
- BPFrequency<-, genomicProfilesInternal, DNASTringSet-method (genomicProfilesInternal-class), 46
- BPFrequency<-, genomicProfilesInternal, vector-method (genomicProfilesInternal-class), 46
- chip (ChIPAnalyserData), 16
- ChIPAnalyser (ChIPAnalyser-package), 5
- ChIPAnalyser-package, 5
- ChIPAnalyserData, 16
- chipMean, 17, 17, 72, 92
- chipMean, parameterOptions-method (parameterOptions-class), 74
- chipMean-methods, 18
- chipMean<-, 19
- chipMean<-methods, 20
- chipMean<-, parameterOptions, numeric-method (parameterOptions-class), 74
- ChIPScore, 27, 28, 35, 53–56, 58, 103
- ChIPScore-class, 20
- chipSd, 22, 72, 92
- chipSd, parameterOptions-method (parameterOptions-class), 74
- chipSd-methods, 23
- chipSd<-, 23
- chipSd<-methods, 24
- chipSd<-, parameterOptions, numeric-method (parameterOptions-class), 74
- chipSmooth, 24, 72, 92
- chipSmooth, parameterOptions-method (parameterOptions-class), 74
- chipSmooth-methods, 25
- chipSmooth<-, 25
- chipSmooth<-methods, 26
- chipSmooth<-, parameterOptions, vector-method (parameterOptions-class), 74
- computeChIPProfile, 8, 9, 18, 19, 22–24, 26, 27, 93, 109, 110
- computeGenomeWideScores, 7, 29, 35, 58, 62
- computeOccupancy, 8, 9, 11, 12, 24, 26, 27, 30, 100, 102
- computeOptimal, 11, 12, 32, 92

- computePWMScore, [31](#), [34](#), [35](#)
- cs (ChIPAnalyserData), [16](#)
- DNASequenceLength, [7](#), [36](#)
- DNASequenceLength, genomicProfilesInternal-method (genomicProfilesInternal-class), [46](#)
- DNASequenceLength-methods, [37](#)
- DNAStringSet, [14](#), [15](#), [33](#), [35](#), [44](#)
- drop, [38](#)
- drop, genomicProfilesInternal-method (genomicProfilesInternal-class), [46](#)
- drop-methods, [39](#)
- evolve, [39](#)
- generateStartingPopulation, [41](#)
- geneRef (ChIPAnalyserData), [16](#)
- genomicProfiles, [6](#), [7](#), [13–15](#), [27](#), [28](#), [31](#), [32](#), [35](#), [36](#), [38](#), [42](#), [43](#), [45](#), [47](#), [58](#), [62](#), [73](#), [76–79](#), [86–91](#), [95](#), [104](#)
- genomicProfiles-class, [43](#)
- genomicProfilesInternal, [45](#)
- genomicProfilesInternal-class, [46](#)
- getHighestFitnessSolutions, [48](#)
- getTestingData, [49](#)
- getTrainingData, [49](#)
- GRanges, [17](#), [27](#), [31](#), [33](#), [35](#), [38](#), [82](#), [92](#)
- GRangesList, [31](#), [35](#)
- GRList-class, [50](#), [50](#)
- initialize, ChIPScore-method (initialize-methods), [51](#)
- initialize, genomicProfiles-method (initialize-methods), [51](#)
- initialize, parameterOptions-method (initialize-methods), [51](#)
- initialize-methods, [51](#)
- lambdaPWM, [28](#), [32](#), [33](#), [51](#), [84](#), [85](#)
- lambdaPWM, parameterOptions-method (parameterOptions-class), [74](#)
- lambdaPWM-methods, [52](#)
- lambdaPWM<-, [52](#)
- lambdaPWM<-methods, [53](#)
- lambdaPWM<-, parameterOptions, vector-method (parameterOptions-class), [74](#)
- loci, [53](#)
- loci, ChIPScore-method (ChIPScore-class), [20](#)
- loci-class, [54](#)
- loci-methods, [55](#)
- lociWidth, [55](#), [72](#)
- lociWidth, parameterOptions-method (parameterOptions-class), [74](#)
- lociWidth-methods, [56](#)
- lociWidth<-, [57](#)
- lociWidth<-methods, [58](#)
- lociWidth<-, parameterOptions, numeric-method (parameterOptions-class), [74](#)
- maxPWMScore, [58](#)
- maxPWMScore, genomicProfilesInternal-method (genomicProfilesInternal-class), [46](#)
- maxPWMScore-methods, [59](#)
- maxSignal, [8](#), [9](#), [59](#), [72](#)
- maxSignal, parameterOptions-method (parameterOptions-class), [74](#)
- maxSignal-methods, [60](#)
- maxSignal<-, [61](#)
- maxSignal<-methods, [62](#)
- maxSignal<-, parameterOptions, numeric-method (parameterOptions-class), [74](#)
- minPWMScore, [62](#)
- minPWMScore, genomicProfilesInternal-method (genomicProfilesInternal-class), [46](#)
- minPWMScore-methods, [63](#)
- naturalLog, [63](#), [72](#)
- naturalLog, parameterOptions-method (parameterOptions-class), [74](#)
- naturalLog-methods, [64](#)
- naturalLog<-, [65](#)
- naturalLog<-methods, [66](#)
- naturalLog<-, parameterOptions, logical-method (parameterOptions-class), [74](#)
- noiseFilter, [66](#), [72](#)
- noiseFilter, parameterOptions-method (parameterOptions-class), [74](#)
- noiseFilter-methods, [67](#)
- noiseFilter<-, [67](#)
- noiseFilter<-methods, [68](#)
- noiseFilter<-, parameterOptions, character-method (parameterOptions-class), [74](#)
- noOfSites, [68](#), [72](#)

- noOfSites,parameterOptions-method
(parameterOptions-class), 74
- noOfSites-methods, 69
- noOfSites<-, 70
- noOfSites<-methods, 71
- noOfSites<- ,parameterOptions,character-method
(parameterOptions-class), 74
- noOfSites<- ,parameterOptions,numeric-method
(parameterOptions-class), 74
- nos-class, 71
- parameterOptions, 8–12, 17–19, 21–28, 31,
33, 35, 43, 45, 47, 51–53, 55–57,
59–61, 63–70, 72, 79–82, 92, 93,
95–102, 109–116
- parameterOptions-class, 74
- PFMFormat, 77
- PFMFormat,genomicProfilesInternal-method
(genomicProfilesInternal-class),
46
- PFMFormat-methods, 78
- PFMFormat<-, 78
- PFMFormat<-methods, 79
- PFMFormat<- ,genomicProfilesInternal,character-method
(genomicProfilesInternal-class),
46
- ploidy, 72, 79
- ploidy,parameterOptions-method
(parameterOptions-class), 74
- ploidy-methods, 80
- ploidy<-, 81
- ploidy<-methods, 82
- ploidy<- ,parameterOptions,numeric-method
(parameterOptions-class), 74
- plotOccupancyProfile, 8, 9, 82
- plotOptimalHeatMaps, 84
- PositionFrequencyMatrix, 77, 78, 86
- PositionFrequencyMatrix,genomicProfilesInternal-method
(genomicProfilesInternal-class),
46
- PositionFrequencyMatrix-methods, 87
- PositionFrequencyMatrix<-, 87
- PositionFrequencyMatrix<-methods, 88
- PositionFrequencyMatrix<- ,genomicProfilesInternal,character-method
(genomicProfilesInternal-class),
46
- PositionFrequencyMatrix<- ,genomicProfilesInternal,numeric-method
(genomicProfilesInternal-class),
46
- PositionWeightMatrix, 86, 89
- PositionWeightMatrix,genomicProfilesInternal-method
(genomicProfilesInternal-class),
46
- PositionWeightMatrix-methods, 90
- PositionWeightMatrix<- , 90
- PositionWeightMatrix<-methods, 91
- PositionWeightMatrix<- ,genomicProfilesInternal,matrix-method
(genomicProfilesInternal-class),
46
- processingChIP, 20, 21, 27, 33, 35, 54, 56,
57, 72, 91, 103
- profileAccuracyEstimate, 8, 9, 82, 93
- profiles, 31, 35, 50
- profiles(profiles-methods), 95
- profiles,genomicProfilesInternal-method
(genomicProfilesInternal-class),
46
- profiles-methods, 95
- PWMpseudocount, 73, 95
- PWMpseudocount,parameterOptions-method
(parameterOptions-class), 74
- PWMpseudocount-methods, 96
- PWMpseudocount<- , 96
- PWMpseudocount<-methods, 97
- PWMpseudocount<- ,parameterOptions,numeric-method
(parameterOptions-class), 74
- PWMThreshold, 29, 34, 35, 73, 98
- PWMThreshold,parameterOptions-method
(parameterOptions-class), 74
- PWMThreshold-methods, 99
- PWMThreshold<- , 99
- PWMThreshold<-methods, 100
- PWMThreshold<- ,parameterOptions,numeric-method
(parameterOptions-class), 74
- removeBackground, 72, 100
- removeBackground,parameterOptions-method
(parameterOptions-class), 74
- removeBackground-methods, 101
- removeBackground<- , 101
- removeBackground<-methods, 102
- removeBackground<- ,parameterOptions,vector-method
(parameterOptions-class), 74
- scores, 103
- scores,ChIPScore-method
(ChIPScore-class), 20
- scores-methods, 104

searchSites, [82](#), [104](#)
setChromatinStates, [106](#)
show, ChIPScore-method (show-methods),
 [107](#)
show, genomicProfiles-method
 (show-methods), [107](#)
show, parameterOptions-method
 (show-methods), [107](#)
show-methods, [107](#)
singleRun, [107](#)
splitData, [108](#)
stepSize, [72](#), [109](#)
stepSize, parameterOptions-method
 (parameterOptions-class), [74](#)
stepSize-methods, [110](#)
stepSize<-, [110](#)
stepSize<-methods, [111](#)
stepSize<-, parameterOptions, numeric-method
 (parameterOptions-class), [74](#)
strandRule, [73](#), [111](#)
strandRule, parameterOptions-method
 (parameterOptions-class), [74](#)
strandRule-methods, [112](#)
strandRule<-, [113](#)
strandRule<-methods, [114](#)
strandRule<-, parameterOptions, character-method
 (parameterOptions-class), [74](#)

top (ChIPAnalyserData), [16](#)

whichstrand, [73](#), [112](#), [113](#), [114](#)
whichstrand, parameterOptions-method
 (parameterOptions-class), [74](#)
whichstrand-methods, [115](#)
whichstrand<-, [115](#)
whichstrand<-methods, [116](#)
whichstrand<-, parameterOptions, character-method
 (parameterOptions-class), [74](#)