

# Package ‘sosta’

July 24, 2025

**Title** A package for the analysis of anatomical tissue structures in spatial omics data

**Version** 1.0.1

**Description** sosta (Spatial Omics SStructure Analysis) is a package for analyzing spatial omics data to explore tissue organization at the anatomical structure level. It reconstructs anatomically relevant structures based on molecular features or cell types. It further calculates a range of metrics at the structure level to quantitatively describe tissue architecture. The package is designed to integrate with other packages for the analysis of spatial omics data.

**License** GPL (>= 3) + file LICENSE

**Encoding** UTF-8

**Depends** R (>= 4.4.0)

**Roxygen** list(markdown = TRUE)

**RoxygenNote** 7.3.2

**URL** <https://github.com/sgunz/sosta>, <https://sgunz.github.io/sosta/>

**BugReports** <https://github.com/sgunz/sosta/issues>

**biocViews** Software, Spatial, Transcriptomics, Visualization

**Imports** terra, sf, smoothr, spatstat.explore, spatstat.geom, SpatialExperiment, SingleCellExperiment, dplyr, ggplot2, patchwork, SummarizedExperiment, stats, rlang, parallel, EBImage, spatstat.random, S4Vectors

**Suggests** knitr, rmarkdown, BiocStyle, ExperimentHub, lme4, lmerTest, ggfortify, tidyverse, testthat (>= 3.0.0)

**VignetteBuilder** knitr

**Config/testthat.edition** 3

**LazyData** true

**git\_url** <https://git.bioconductor.org/packages/sosta>

**git\_branch** RELEASE\_3\_21

**git\_last\_commit** 0ab9772

**git\_last\_commit\_date** 2025-07-17

**Repository** Bioconductor 3.21

**Date/Publication** 2025-07-23

**Author** Samuel Gunz [aut, cre] (ORCID: <<https://orcid.org/0000-0002-8909-0932>>),  
Mark D. Robinson [aut, fnd]

**Maintainer** Samuel Gunz <[samuel.gunz@uzh.ch](mailto:samuel.gunz@uzh.ch)>

## Contents

sosta-package . . . . .	3
.df2ppp . . . . .	3
.intensityImage . . . . .	4
.intensityThreshold . . . . .	5
.SPE2df . . . . .	5
assingCellsToStructures . . . . .	6
binaryImageToSF . . . . .	7
cellTypeProportions . . . . .	8
createPointPatternTissue . . . . .	9
estimateReconstructionParametersSPE . . . . .	10
findIntensityThreshold . . . . .	11
getDimXY . . . . .	11
meanShapeMetrics . . . . .	12
minBoundaryDistances . . . . .	13
normalizeCoordinates . . . . .	14
reconstructShapeDensity . . . . .	15
reconstructShapeDensityImage . . . . .	15
reconstructShapeDensitySPE . . . . .	17
shapeIntensityImage . . . . .	18
shapeMetrics . . . . .	19
simulateTissueBlobs . . . . .	20
sostaSPE . . . . .	20
spatialCoords2SF . . . . .	21
SPE2ppp . . . . .	22
stCalculateCurvature . . . . .	22
stCalculateShapeCurl . . . . .	23
stFeatureAxes . . . . .	24
totalShapeMetrics . . . . .	25
xyCoordinates . . . . .	25

---

**sosta-package***sosta: A package for the analysis of anatomical tissue structures in spatial omics data*

---

## Description

sosta (Spatial Omics STructure Analysis) is a package for analyzing spatial omics data to explore tissue organization at the anatomical structure level. It reconstructs anatomically relevant structures based on molecular features or cell types. It further calculates a range of metrics at the structure level to quantitatively describe tissue architecture. The package is designed to integrate with other packages for the analysis of spatial omics data.

## Author(s)

**Maintainer:** Samuel Gunz <[samuel.gunz@uzh.ch](mailto:samuel.gunz@uzh.ch)> ([ORCID](#))

Authors:

- Mark D. Robinson <[mark.robinson@mls.uzh.ch](mailto:mark.robinson@mls.uzh.ch)> [funder]

## See Also

Useful links:

- <https://github.com/sgunz/sosta>
  - <https://sgunz.github.io/sosta/>
  - Report bugs at <https://github.com/sgunz/sosta/issues>
- 

**.df2ppp***Function to convert data.frame to ppp object*

---

## Description

Assumes that the `data.frame` is the output of `.SPE2df()`. Column order is important!

## Usage

```
.df2ppp(df, xName, yName, marks = NULL)
```

## Arguments

<code>df</code>	data.frame; with x, y coordinates, image, and categorical mark information.
<code>xName</code>	character; column name of x coordinate
<code>yName</code>	character; column name of y coordinate
<code>marks</code>	character; column name of the mark variable

**Value**

ppp; object of type ppp

**See Also**

[.SPE2df](#), [as.ppp](#)

**Examples**

```
data(sostaSPE)
df <- .SPE2df(sostaSPE, marks = "cellType", imageCol = "imageName")
ppp <- .df2ppp(df, xName = "x", yName = "y", marks = "cellType")
```

**.intensityImage**

*Function to estimate the intensity image of a point pattern*

**Description**

Function to estimate the intensity image of a point pattern

**Usage**

```
.intensityImage(ppp, markSelect = NULL, bndw = NULL, dim)
```

**Arguments**

ppp	point pattern object of class ppp
markSelect	character; name of mark that is to be selected for the reconstruction
bndw	bandwidth of kernel density estimator
dim	numeric; x dimension of the final reconstruction.

**Value**

list; list with the intensity image and the bandwidth and dimension parameters

---

.intensityThreshold     *Function to estimate the intensity threshold for the reconstruction of spatial structures*

---

### Description

Function to estimate the intensity threshold for the reconstruction of spatial structures

### Usage

```
.intensityThreshold(densityImage, steps = 250)
```

### Arguments

densityImage	real-valued pixel image; output from the function .intensityImage
steps	numeric; value used to filter the density estimates, where only densities greater than the maximum value divided by threshold are considered. Default is 250.

### Value

numeric; estimated threshold

---

.SPE2df                    *Function to convert SpatialExperiment object to a data frame*

---

### Description

Function to convert SpatialExperiment object to a data frame

### Usage

```
.SPE2df(spe, imageCol = NULL, marks = NULL, colNames = FALSE)
```

### Arguments

spe	SpatialExperiment; a object of class SpatialExperiment
imageCol	character; name of a column in colData that corresponds to the image
marks	character; name of column in colData with categorical marks
colNames	logical; extract colnames from SpatialExperiment

### Value

data.frame with x, y coordinates, image, and categorical mark information

## Examples

```
data(sostaSPE)
.SPE2df(sostaSPE, marks = "cellType", imageCol = "imageName") |> head()
```

### assingCellsToStructures

*Function to assign points / coordinates to structures*

## Description

This function assigns each spatial coordinate in a `SpatialExperiment` object (`spe`) to the first intersecting structure from a given set of spatial structures.

## Usage

```
assingCellsToStructures(
  spe,
  allStructs,
  imageCol,
  uniqueId = "structID",
  nCores = 1
)
```

## Arguments

<code>spe</code>	<code>SpatialExperiment</code> ; An object of class <code>SpatialExperiment</code> containing spatial point data. Must contain colnames for correct assignment.
<code>allStructs</code>	<code>sf</code> ; A simple feature collection ( <code>sf</code> object) representing spatial structures. Must contain a column which contains a unique identifier for each structure. Default = <code>structID</code> .
<code>imageCol</code>	character; The column name in <code>spe</code> and <code>allStructs</code> that identifies the corresponding image.
<code>uniqueId</code>	character; The column name in the simple feature collection for which to compute the assignment.
<code>nCores</code>	integer; The number of cores to use for parallel processing (default is 1).

## Value

A named list with structure assignments for each spatial point in `spe`. Points that do not overlap with any structure are assigned NA. Names correspond to colnames of the `SpatialExperiment` input object.

## Examples

```

library("SpatialExperiment")
data("sostaSPE")
allStructs <- reconstructShapeDensitySPE(sostaSPE,
  marks = "cellType", imageCol = "imageName",
  markSelect = "A", bndw = 3.5, thres = 0.045
)
# The function `assingCellsToStructures` needs colnames so we create them here
colnames(sostaSPE) <- paste0("cell_", c(1:dim(sostaSPE)[2]))

res <- assingCellsToStructures(
  spe = sostaSPE, allStructs = allStructs, imageCol = "imageName"
)
# Assign the structure assignment in the order of the columns in the `SpatialExperiment` object
colData(sostaSPE)$structAssign <- res[colnames(sostaSPE)]

if (require("ggplot2")) {
  cbind(
    colData(sostaSPE[, sostaSPE[["imageName"]] == "image1"]),
    spatialCoords(sostaSPE[, sostaSPE[["imageName"]] == "image1"])
  ) |>
    as.data.frame() |>
    ggplot(aes(x = x, y = y, color = structAssign)) +
    geom_point(size = 0.25) +
    coord_equal()
}

```

`binaryImageToSF`      *Converts a binary matrix to an sf polygon*

## Description

Converts a binary matrix to an sf polygon

## Usage

```
binaryImageToSF(binaryMatrix, xmin, xmax, ymin, ymax)
```

## Arguments

<code>binaryMatrix</code>	matrix; binary matrix
<code>xmin</code>	integer; minimum x coordinate of the coordinate system
<code>xmax</code>	integer; maximum x coordinate of the coordinate system
<code>ymin</code>	integer; minimum y coordinate of the coordinate system
<code>ymax</code>	integer; maximum y coordinate of the coordinate system

## Value

sf object

## Examples

```
matrixR <- matrix(c(
  0, 0, 0, 0, 0, 0, 0, 0, 0,
  0, 1, 1, 1, 1, 1, 0, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 1, 1, 1, 1, 1, 0, 0, 0,
  0, 1, 1, 0, 1, 1, 0, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 1, 1, 0, 0, 0, 1, 1, 0,
  0, 0, 0, 0, 0, 0, 0, 0, 0
), nrow = 9, byrow = TRUE)
polyR <- binaryImageToSF(matrixR, xmin = 0, xmax = 1, ymin = 0, ymax = 1)
plot(polyR)
```

**cellTypeProportions**     *Calculate the proportion of each cell type within spatial structures*

## Description

Calculate the proportion of each cell type within spatial structures

## Usage

```
cellTypeProportions(spe, structColumn, cellTypeColumn, nCores = 1)
```

## Arguments

spe	SpatialExperiment object
structColumn	character; name of the colData column specifying the structure assignments
cellTypeColumn	character; name of the colData column specifying cell types
nCores	integer; The number of cores to use for parallel processing (default is 1).

## Value

A data frame where rows correspond to unique structures and columns correspond to cell types, containing the proportion of each cell type within each structure.

## Examples

```
library("SpatialExperiment")
data("sostaSPE")
allStructs <- reconstructShapeDensitySPE(sostaSPE,
  marks = "cellType", imageCol = "imageName",
  markSelect = "A", bndw = 3.5, thres = 0.045
)
# The function `assingCellsToStructures` needs colnames so we create them here
colnames(sostaSPE) <- paste0("cell_", c(1:dim(sostaSPE)[2]))
```

```
# Assign the structure assignment in the order of the columns in the `SpatialExperiment` object
colData(sostaSPE)$structAssign <- assingCellsToStructures(
  spe = sostaSPE, allStructs = allStructs, imageCol = "imageName"
)[colnames(sostaSPE)]
cellTypeProportions(sostaSPE, "structAssign", "cellType")
```

---

**createPointPatternTissue**

*Create a Point Pattern on a Simulated Tissue Image*

---

**Description**

This function generates a spatial point pattern with different types of points (A, B, C) distributed over the simulated tissue structure.

**Usage**

```
createPointPatternTissue(tissueImage, intA, intB, intCInA, intCInB)
```

**Arguments**

tissueImage	Matrix; A binary matrix representing the simulated tissue.
intA	Numeric; Intensity of type "A" points (points per unit area) on tissue regions.
intB	Numeric; Intensity of type "B" points (points per unit area) on non-tissue regions.
intCInA	Numeric; Intensity of type "C" points placed in extended regions around tissue.
intCInB	Numeric; Intensity of type "C" points placed within tissue.

**Value**

A ppp object representing the spatial point pattern.

**Examples**

```
tissueImage <- simulateTissueBlobs(128, 100, 7)
createPointPatternTissue(tissueImage, 0.1, 0.1, 0.005, 0.005)
```

**estimateReconstructionParametersSPE***Estimate reconstruction parameters from a set of images***Description**

Estimate reconstruction parameters from a set of images

**Usage**

```
estimateReconstructionParametersSPE(
  spe,
  marks,
  imageCol,
  markSelect = NULL,
  nImages = NULL,
  fun = "bw.diggle",
  dim = 500,
  nCores = 1,
  plotHist = TRUE
)
```

**Arguments**

spe	SpatialExperiment; a object of class SpatialExperiment
marks	character; name of column in colData that will correspond to the ppp marks
imageCol	character; name of a column in colData that corresponds to the image
markSelect	character; name of mark that is to be selected for the reconstruction
nImages	integer; number of images for the estimation. Will be randomly sampled
fun	character; function to estimate the kernel density. Default bw.diggle.
dim	numeric; x dimension of the final reconstruction. A lower resolution speed up computation but lead to less exact reconstruction. Default = 500
nCores	numeric; number of cores for parallel processing using mclapply. Default = 1
plotHist	logical; if histogram of estimated densities and thresholds should be plotted. Default = TRUE

**Value**

tibble; tibble with estimated intensities and thresholds

**Examples**

```
data("sostaSPE")
estimateReconstructionParametersSPE(sostaSPE,
  marks = "cellType", imageCol = "imageName",
  markSelect = "A", plotHist = TRUE
)
```

---

**findIntensityThreshold**

*Estimate the intensity threshold for the reconstruction of spatial structures*

---

**Description**

Estimate the intensity threshold for the reconstruction of spatial structures

**Usage**

```
findIntensityThreshold(ppp, markSelect = NULL, bndw = NULL, dim, steps = 250)
```

**Arguments**

ppp	point pattern object of class ppp
markSelect	character; name of mark that is to be selected for the reconstruction
bndw	numeric; bandwith of the sigma parameter in the density estimation, if no value is given the bandwith is estimated using cross validation with the bw.diggle function.
dim	numeric; x dimension of the final reconstruction.
steps	numeric; value used to filter the density estimates, where only densities greater than the maximum value divided by threshold are considered. Default is 250.

**Value**

numeric; estimated intensity threshold

**Examples**

```
data(sostaSPE)
ppp <- SPE2ppp(sostaSPE, marks = "cellType", imageCol = "imageName", imageId = "image1")
findIntensityThreshold(ppp, markSelect = "A", dim = 250)
```

---

**getDimXY**

*Function to get the dimension based on dim of y axis*

---

**Description**

Function to get the dimension based on dim of y axis

**Usage**

```
getDimXY(ppp, ydim)
```

**Arguments**

<code>ppp</code>	point pattern object of class ppp
<code>ydim</code>	dimension of y axis

**Value**

vector; vector with x and y dimension

**Examples**

```
data(sostaSPE)
pp <- SPE2ppp(sostaSPE,
  marks = "cellType", imageCol = "imageName",
  imageId = "image1"
)
getDimXY(pp, 500)
```

**meanShapeMetrics**      *Calculate mean shape metrics of a set of polygons*

**Description**

Calculate mean shape metrics of a set of polygons

**Usage**

```
meanShapeMetrics(totalShapeMetricMatrix)
```

**Arguments**

<code>totalShapeMetricMatrix</code>	
	matrix of shape metrics

**Value**

matrix; matrix of mean shape metrics

**Examples**

```
data(sostaSPE)
struct <- reconstructShapeDensityImage(sostaSPE,
  marks = "cellType", imageCol = "imageName",
  imageId = "image1", markSelect = "A", dim = 500
)
shapeMetrics <- totalShapeMetrics(struct)
meanShapeMetrics(shapeMetrics)
```

---

<code>minBoundaryDistances</code>	<i>Compute minimum boundary distances for each cell within its corresponding image structures</i>
-----------------------------------	---

---

## Description

Compute minimum boundary distances for each cell within its corresponding image structures

## Usage

```
minBoundaryDistances(spe, imageCol, structColumn, allStructs, nCores = 1)
```

## Arguments

<code>spe</code>	SpatialExperiment object
<code>imageCol</code>	character; name of the colData column specifying the image name
<code>structColumn</code>	character; name of the colData column specifying structure assignments
<code>allStructs</code>	sf object; contains spatial structures with corresponding image names
<code>nCores</code>	integer; The number of cores to use for parallel processing (default is 1).

## Value

A named list containing the minimum distances between cells and structure boundaries, values within structures have negative values. Names correspond to colnames of the SpatialExperiment input object.

## Examples

```
library("SpatialExperiment")
data("sostaSPE")

allStructs <- reconstructShapeDensitySPE(sostaSPE,
  marks = "cellType", imageCol = "imageName",
  markSelect = "A", bndw = 3.5, thres = 0.045
)
# The function `assingCellsToStructures` needs colnames so we create them here
colnames(sostaSPE) <- paste0("cell_", c(1:dim(sostaSPE)[2]))
# Assign the structure assignment in the order of the columns in the `SpatialExperiment` object
colData(sostaSPE)$structAssign <- assingCellsToStructures(
  spe = sostaSPE, allStructs = allStructs, imageCol = "imageName"
)[colnames(sostaSPE)]

res <- minBoundaryDistances(
  spe = sostaSPE, imageCol = "imageName", structColumn = "structAssign",
  allStructs = allStructs
)
colData(sostaSPE)$minDist <- res[colnames(sostaSPE)]
```

```

if (require("ggplot2")) {
  cbind(colData(sostaSPE), spatialCoords(sostaSPE)) |>
    as.data.frame() |>
    ggplot(aes(x = x, y = y, color = minDist)) +
    geom_point(size = 0.25) +
    scale_colour_gradient2() +
    geom_sf(data = allStructs, fill = NA, inherit.aes = FALSE) +
    facet_wrap(~imageName)
}

```

**normalizeCoordinates** *Function to normalize coordinates between zero and one while keep scaling*

## Description

Function to normalize coordinates between zero and one while keep scaling

## Usage

```
normalizeCoordinates(coords)
```

## Arguments

coords	matrix; matrix with coordinates
--------	---------------------------------

## Value

matrix; coordinates scaled between 0 and 1

## Examples

```

matrixR <- matrix(c(
  0, 0, 0, 0, 0, 0, 0, 0, 0,
  0, 1, 1, 1, 1, 1, 0, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 1, 1, 1, 1, 1, 0, 0, 0,
  0, 1, 1, 0, 1, 1, 0, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 0, 0, 0, 0, 0, 0, 0, 0
), nrow = 9, byrow = TRUE)
coords <- xyCoordinates(matrixR)
normalizeCoordinates(coords)

```

**reconstructShapeDensity***Reconstruct polygon from point pattern density***Description**

This function estimates the density of a spatial point pattern (ppp), thresholds the density to create a binary image, and then converts it to a valid sf object (polygons).

**Usage**

```
reconstructShapeDensity(ppp, markSelect = NULL, bndw = NULL, thres = NULL, dim)
```

**Arguments**

ppp	point pattern object of class ppp
markSelect	character; name of mark that is to be selected for the reconstruction
bndw	bandwidth of kernel density estimator
thres	intensity threshold for the reconstruction
dim	numeric; x dimension of the final reconstruction.

**Value**

sf object of class POLYGON

**Examples**

```
data("sostaSPE")
ppp <- SPE2ppp(sostaSPE, marks = "cellType", imageCol = "imageName", imageId = "image1")
thres <- findIntensityThreshold(ppp, markSelect = "A", dim = 500)
struct <- reconstructShapeDensity(ppp, markSelect = "A", thres = thres, dim = 500)
plot(struct)
```

**reconstructShapeDensityImage***Reconstruct structure from spe object with given image id***Description**

Reconstruct structure from spe object with given image id

**Usage**

```
reconstructShapeDensityImage(
  spe,
  marks,
  imageCol,
  imageId,
  markSelect,
  dim = 500,
  bndw = NULL,
  thres = NULL
)
```

**Arguments**

<code>spe</code>	SpatialExperiment; a object of class SpatialExperiment
<code>marks</code>	character; name of column in colData that will correspond to the ppp marks
<code>imageCol</code>	character; name of a column in colData that corresponds to the image
<code>imageId</code>	character; image id, must be present in imageCol
<code>markSelect</code>	character; name of mark that is to be selected for the reconstruction
<code>dim</code>	numeric; x dimension of the final reconstruction. A lower resolution speed up computation but lead to less exact reconstruction. Default = 500
<code>bndw</code>	numeric; smoothing bandwidth in the density estimation, corresponds to the sigma parameter in the density.ppp function, if no value is given the bandwidth is estimated using cross validation with the bw.diggle function.
<code>thres</code>	numeric; intensity threshold for the reconstruction; if NULL the threshold is set as the mean between the mode of the pixel intensity distributions

**Value**

sf object of class POLYGON

**Examples**

```
data("sostaSPE")
struct <- reconstructShapeDensityImage(sostaSPE,
  marks = "cellType", imageCol = "imageName", imageId = "image1",
  markSelect = "A", dim = 500
)
plot(struct)
```

---

**reconstructShapeDensitySPE**

*Reconstruct structure from spatial experiment object per image id*

---

**Description**

Reconstruct structure from spatial experiment object per image id

**Usage**

```
reconstructShapeDensitySPE(  
  spe,  
  marks,  
  imageCol,  
  markSelect,  
  dim = 500,  
  bndw = NULL,  
  thres = NULL,  
  nCores = 1  
)
```

**Arguments**

spe	SpatialExperiment; a object of class SpatialExperiment
marks	character; name of column in colData that will correspond to the ppp marks
imageCol	character; name of a column in colData that corresponds to the image
markSelect	character; name of mark that is to be selected for the reconstruction
dim	numeric; x dimension of the final reconstruction. A lower resolution speed up computation but lead to less exact reconstruction. Default = 500
bndw	numeric; bandwidth of the sigma parameter in the density estimation, if no value is given the bandwidth is estimated using cross validation with the bw.diggle function for each image individually.
thres	numeric; intensity threshold for the reconstruction; if NULL the threshold is set as the mean between the mode of the pixel intensity distributions estimated for each image individual
nCores	numeric; number of cores for parallel processing using mclapply. Default = 1

**Value**

simple feature collection

## Examples

```
data("sostaSPE")
allStructs <- reconstructShapeDensitySPE(sostaSPE,
  marks = "cellType", imageCol = "imageName",
  markSelect = "A", bndw = 3.5, thres = 0.005
)
allStructs
```

`shapeIntensityImage`    *Intensity plot*

## Description

This function plots the intensity of a point pattern image and displays a histogram of the intensity values. Note that intensities less than largest intensity value divided by 250 are not displayed in the histogram.

## Usage

```
shapeIntensityImage(
  spe,
  marks,
  imageCol,
  imageId,
  markSelect,
  bndw = NULL,
  dim = 500
)
```

## Arguments

<code>spe</code>	SpatialExperiment; a object of class SpatialExperiment
<code>marks</code>	character; name of column in colData that will correspond to the ppp marks
<code>imageCol</code>	character; name of a column in colData that corresponds to the image
<code>imageId</code>	character; image id, must be present in imageCol
<code>markSelect</code>	character; name of mark that is to be selected for the reconstruction
<code>bndw</code>	numeric; smoothing bandwidth in the density estimation, corresponds to the sigma parameter in the density.ppp function, if no value is given the bandwidth is estimated using cross validation with the bw.diggle function.
<code>dim</code>	numeric; x dimension of the final reconstruction. A lower resolution speeds up computation but lead to less exact reconstruction. Default = 500

## Value

ggplot object with intensity image and histogram

## Examples

```
data("sostaSPE")
shapeIntensityImage(sostaSPE,
  marks = "cellType", imageCol = "imageName",
  imageId = "image1", markSelect = "A"
)
```

**shapeMetrics**

*Calculate a set of shape metrics of a polygon*

## Description

Calculate a set of shape metrics of a polygon

## Usage

```
shapeMetrics(sfPoly)
```

## Arguments

sfPoly	POLYGON of class sfc
--------	----------------------

## Value

list; list of shape metrics

## Examples

```
matrix_R <- matrix(c(
  0, 0, 0, 0, 0, 0, 0, 0, 0,
  0, 1, 1, 1, 1, 1, 0, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 1, 1, 1, 1, 1, 0, 0, 0,
  0, 1, 1, 0, 1, 1, 0, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 0, 0, 0, 0, 0, 0, 0, 0
), nrow = 9, byrow = TRUE)
polyR <- binaryImageToSF(matrix_R, xmin = 0, xmax = 1, ymin = 0, ymax = 1)
shapeMetrics(polyR)
```

`simulateTissueBlobs`    *Simulate Tissue Blobs*

### Description

This function generates a simulated tissue-like structure using a Gaussian blur technique.

### Usage

```
simulateTissueBlobs(size, seedNumber, clumpSize)
```

### Arguments

<code>size</code>	Integer; The size (width and height) of the simulated tissue image.
<code>seedNumber</code>	Integer; The number of random seed points used to generate tissue blobs.
<code>clumpSize</code>	Numeric; The standard deviation (sigma) of the Gaussian blur applied to generate tissue clumps.

### Value

A binary matrix representing the simulated tissue structure.

### Examples

```
tissueImage <- simulateTissueBlobs(128, 100, 7)
image(tissueImage)
```

`sostaSPE`

*Example SpatialExperiment Object with Simulated Tissue Images and Point Patterns*

### Description

This dataset contains a simulated `SpatialExperiment` object (`sostaSPE`) representing three tissue images, each with a corresponding spatial point pattern. The point patterns contain different cell types (A, B, and C), distributed according to simulated tissue structures.

### Usage

```
sostaSPE
```

## Format

A SpatialExperiment object with the following structure:

- x** Numeric; x-coordinate of each point (cell location).
- y** Numeric; y-coordinate of each point (cell location).
- cell\_type** Factor; Cell type assigned to each point (A, B, or C).
- image\_name** Factor; Identifier for the tissue image (image1, image2, or image3).

## Details

The dataset was generated as follows:

- Three tissue images were simulated using `simulateTissueBlobs()`.
- Spatial point patterns were created for each tissue using `createPointPatternTissue()`.
- The point pattern data was converted into a SpatialExperiment object with spatial coordinates.

`spatialCoords2SF`

*Function to convert spatialCoords to an sf object*

## Description

Function to convert spatialCoords to an sf object

## Usage

`spatialCoords2SF(spe)`

## Arguments

`spe`      SpatialExperiment; a object of class SpatialExperiment

## Value

sf; Simple feature collection of geometry type POINT

## Examples

```
data(sostaSPE)
speSel <- sostaSPE[, sostaSPE[["imageName"]] == "image1"]
spatialCoords2SF(speSel)
```

**SPE2ppp***Function to convert spatial coordinates of a SpatialExperiment object to a ppp object***Description**

Function to convert spatial coordinates of a SpatialExperiment object to a ppp object

**Usage**

```
SPE2ppp(spe, marks = NULL, imageCol = NULL, imageId = NULL)
```

**Arguments**

<code>spe</code>	SpatialExperiment; a object of class SpatialExperiment
<code>marks</code>	character; name of column in colData that will correspond to the ppp marks
<code>imageCol</code>	character; name of a column in colData that corresponds to the image
<code>imageId</code>	character; image id, must be present in imageCol

**Value**

ppp; object of type ppp

**Examples**

```
data(sostaSPE)
SPE2ppp(sostaSPE,
         marks = "cellType", imageCol = "imageName",
         imageId = "image1"
     )
```

**stCalculateCurvature**    *Calculate curvature of sf object***Description**

Calculate curvature of sf object

**Usage**

```
stCalculateCurvature(sfPoly, smoothness = 5)
```

**Arguments**

<code>sfPoly</code>	POLYGON of class sf
<code>smoothness</code>	list; curvature measures

**Value**

list; list of curvatures values

**References**

<https://stackoverflow.com/questions/62250151/calculate-curvature-of-a-closed-object-in-r>

**Examples**

```
matrixR <- matrix(c(
  0, 0, 0, 0, 0, 0, 0, 0, 0,
  0, 1, 1, 1, 1, 1, 0, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 1, 1, 1, 1, 1, 0, 0, 0,
  0, 1, 1, 0, 1, 1, 0, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 0, 0, 0, 0, 0, 0, 0, 0
), nrow = 9, byrow = TRUE)
polyR <- binaryImageToSF(matrixR, xmin = 0, xmax = 1, ymin = 0, ymax = 1)
stCalculateCurvature(polyR)
```

stCalculateShapeCurl    *Calculate curl of a polygon*

**Description**

Calculate curl of a polygon

**Usage**

```
stCalculateShapeCurl(sfPoly)
```

**Arguments**

sfPoly	POLYGON of class sf
--------	---------------------

**Value**

numeric; the curl of the polygon

## Examples

```
matrixR <- matrix(c(
  1, 1, 1, 1, 1, 0,
  1, 1, 0, 0, 1, 1,
  1, 1, 0, 0, 1, 1,
  1, 1, 1, 1, 1, 0,
  1, 1, 0, 1, 1, 0,
  1, 1, 0, 0, 1, 1,
  1, 1, 0, 0, 1, 1
), nrow = 7, byrow = TRUE)
polyR <- binaryImageToSF(matrixR, xmin = 0, xmax = 1, ymin = 0, ymax = 1)
stCalculateShapeCurl(polyR)
```

**stFeatureAxes**

*Calculate the length of feature axes of an sf polygon*

## Description

Calculate the length of feature axes of an sf polygon

## Usage

```
stFeatureAxes(sfPoly)
```

## Arguments

sfPoly	POLYGON of class sf
--------	---------------------

## Value

list; list containing the major and minor axis lengths

## Examples

```
matrixR <- matrix(c(
  0, 0, 0, 0, 0, 0, 0, 0, 0,
  0, 1, 1, 1, 1, 1, 0, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 1, 1, 1, 1, 1, 0, 0, 0,
  0, 1, 1, 0, 1, 1, 0, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 0, 0, 0, 0, 0, 0, 0, 0
), nrow = 9, byrow = TRUE)
polyR <- binaryImageToSF(matrixR, xmin = 0, xmax = 1, ymin = 0, ymax = 1)
stFeatureAxes(polyR)
```

---

totalShapeMetrics      *Calculate a set of shape metrics of a set of polygons*

---

### Description

Calculate a set of shape metrics of a set of polygons

### Usage

```
totalShapeMetrics(sfInput)
```

### Arguments

sfInput      MULTIPOLYGON of class sf

### Details

Calculate a set of shape metrics of a set of polygons. The function calculates all metrics that are implemented in the function `shapeMetrics()`

### Value

matrix; matrix of shape metrics

### Examples

```
data(sostaSPE)
struct <- reconstructShapeDensityImage(sostaSPE,
  marks = "cellType", imageCol = "imageName",
  imageId = "image1", markSelect = "A", dim = 500
)
totalShapeMetrics(struct)
```

---

xyCoordinates      *Function to extract x y coordinates from binary image*

---

### Description

Function to extract x y coordinates from binary image

### Usage

```
xyCoordinates(inputMatrix)
```

### Arguments

inputMatrix      a binary matrix

**Value**

matrix; matrix with x,y coordinates of the cell of the input matrix

**Examples**

```
matrixR <- matrix(c(
  0, 0, 0, 0, 0, 0, 0, 0, 0,
  0, 1, 1, 1, 1, 1, 0, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 1, 1, 1, 1, 1, 0, 0, 0,
  0, 1, 1, 0, 1, 1, 0, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 1, 1, 0, 0, 1, 1, 0, 0,
  0, 0, 0, 0, 0, 0, 0, 0, 0
), nrow = 9, byrow = TRUE)
xyCoordinates(matrixR)
```

# Index

- \* **datasets**
  - sostaSPE, 20
- \* **internal**
  - sosta-package, 3
  - .SPE2df, 4, 5
  - .df2ppp, 3
  - .intensityImage, 4
  - .intensityThreshold, 5
- as.ppp, 4
- assingCellsToStructures, 6
- binaryImageToSF, 7
- cellTypeProportions, 8
- createPointPatternTissue, 9
- estimateReconstructionParametersSPE,  
10
- findIntensityThreshold, 11
- getDimXY, 11
- meanShapeMetrics, 12
- minBoundaryDistances, 13
- normalizeCoordinates, 14
- reconstructShapeDensity, 15
- reconstructShapeDensityImage, 15
- reconstructShapeDensitySPE, 17
- shapeIntensityImage, 18
- shapeMetrics, 19
- simulateTissueBlobs, 20
- sosta (sosta-package), 3
- sosta-package, 3
- sostasSPE, 20
- spatialCoords2SF, 21
- SPE2ppp, 22
- stCalculateCurvature, 22
- stCalculateShapeCurl, 23
- stFeatureAxes, 24
- totalShapeMetrics, 25
- xyCoordinates, 25