

# PICS

April 20, 2011

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makeRangedDataOutput

*Create a RangedData object from a PICS output*

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## Description

Create a list of 'RangedData' objects from a 'pics' object. The resulting RangedData object can then be analyzed with the 'IRanges' packages and/or exported to bed/wig files with the 'rtracklayer' package.

## Usage

```
makeRangedDataOutput(obj, type="fixed", filter=list(delta=c(0, Inf), se=c(0, Inf), s
```

## Arguments

obj	An object of class 'picsList' as returned by 'PICS' when running it on the IP/Control data.
type	The type of intervals to be created. The different types are 'bed', 'wig', 'ci' and 'fixed'. See details for more info.
filter	A list of filters to be used before computing the FDR. By default all regions are included, see details for more info on how to specify the filters.
length	The length to be used for the fixed type 'RangedData', see details.

## Details

'bed' will generate intervals from the forward peak max to the reverse peak max. 'wig' will generate a density profile for the forward and reverse reads. 'bed' and 'wig' types should be used to be exported to wig/bed files to be used with the UCSC genome browser. 'ci' corresponds to the binding site estimates  $\pm 3 \cdot se$ , while 'fixed' corresponds to the binding site estimates  $\pm 3 \cdot length$ . 'bed' and 'wig' files can be exported using the 'export' function fo the 'rtracklayer' package.

## Value

An object of type 'RangedData'.

## Author(s)

Xuekui Zhang, Arnaud Droit <<arnaud.droit@ircm.qc.ca>> and Raphael Gottardo <<raphael.gottardo@

## References

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq" arXiv, 0903.3206, 2009. To appear in Biometrics.

## See Also

export

## Examples

```
## Not run:
rdBed<-makeRangedDataOutput(pics,type="bed",filter=list(delta=c(50,Inf),se=c(0,50),sigm
export(rbBed,"myfile.bed")
rdBed<-makeRangedDataOutput(pics,type="wig",filter=list(delta=c(50,Inf),se=c(0,50),sigm
export(rbBed,"myfile.wig")
## End(Not run)
```

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pics-class

*The pics class*

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## Description

This object is used to gather all parameters from fitting PICS to a single candidate region. The object contains the following slots: 'estimates', 'infMat', 'Nmerged', 'converge', 'chr'. 'estimates' is a list containing all parameters estimates as well as standard errors. 'infMat' is the Cholesky decomposition of the information matrix, 'converge' is a logical value indicating whether the EM algorithm has converged, while 'chr' is a character string corresponding to a candidate region's chromosome. 'Nmerged' gives the number of binding events that were merged; binding events that overlap are merged (see the cited paper below for details).

## Accessors

The PICS package provide accessors to directly access to most of the parameters/standard errors and chromosome. In the code snippets below, 'x' is a 'pics' object.

**'chromosome(x)'** Gets the chromosome name of the candidate region.

**'mu(x)'** Gets the position estimates of all binding sites identified in the region.

**'delta(x)'** Gets the average fragment lengths of all binding sites identified in the region.

**'sigmaSqF(x)'** Gets the F peak variances of all binding sites identified in the region.

**'sigmaSqR(x)'** Gets the R peak variances of all binding sites identified in the region.

**'seF(x)'** Gets the standard errors of all binding site position estimates identified in the region.

**'seF(x)'** Gets the standard errors of all F peak modes identified in the region.

**'seR(x)'** Gets the standard errors of all R peak modes identified in the region.

**score** signature(x = "pics"): return the score for each binding event.

**scoreF** signature(x = "pics"): return the score of the forward (F) for each binding event.

**scoreR** signature(x = "pics"): return the score of the forward (R) for each binding event.

**Constructor**

`newPics(w,mu,delta,sigmaSqF,sigmaSqR,seMu,seMuF,seMuR,score,Nmerged,converge,infMat,chr)`  
 construct a new 'pics' object with the following arguments:

**w** The mixture weights (a vector)  
**mu** The binding site positions (a vector)  
**delta** The DNA fragment lengths (a vector)  
**sigmaSqF** The variance parameters for the forward distribution (vector)  
**sigmaSqR** The variance parameters for the forward distribution (vector)  
**seMu** The standard errors for mu (vector)  
**seMuF** The standard errors for muF (vector)  
**seMuR** The standard errors for muR (vector)  
**seMuR** The standard errors for muR (vector)  
**score** The scores for each binding event (vector)  
**Nmerged** The number of peaks that got merged (integer)  
**converge** A logical value, TRUE, if the EM as converged  
**infMat** The information matrix  
**chr** The chromosome for the region

**Author(s)**

Xuekui Zhang <<xzhang@stat.ubc.ca>> and Raphael Gottardo <<raphael.gottardo@ircm.qc.ca>>

**References**

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq" arXiv, 0903.3206, 2009. To appear in Biometrics.

**See Also**

[pics](#) [picsError](#)

**Examples**

```
# Here is an example of how to construct such a region.
# Typically, you would not do this manually, you would use the pics function to return a
w<-1
mu<-10000
delta<-150
sigmaSqF<-5000
sigmaSqR<-5000
seMu<-10
seMuF<-10
seMuR<-10
score<-5
Nmerged<-0
converge<-TRUE
chr<-"chr1"
range<-c(1000,2000)
# Constructor
#myPICS<-newPics(w,mu,delta,sigmaSqF,sigmaSqR,seMu,seMuF,seMuR,score,Nmerged,as.integer(r
```

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`picsError-class`     *The pics class*

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### Description

This object is used to return an error code when the PICS function failed to return a valid set of estimates for a candidate regions. This could be due to non-convergence of the EM algorithm, a singular information matrix, or a number of reads below the limit specified by the user. All of these are typically due to too few reads in the region and do not affect the rest of the analysis, as such regions would most likely be labelled as false positives.

### Accessors

All of the accessors defined for a ‘pics’ object still work for a ‘picsError’ object but will simply return a NULL pointer.

### Constructor

`newPicsError(string)` where ‘string’ is the error code.

### Constructor

```
newPicsError<-function(string)
  string The mixture weights (a vector)
```

### Author(s)

Xuekui Zhang <<xzhang@stat.ubc.ca>> and Raphael Gottardo <<raphael.gottardo@ircm.qc.ca>>

### References

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, “PICS: Probabilistic Inference for ChIP-seq” arXiv, 0903.3206, 2009. To appear in Biometrics.

### See Also

[pics](#)

### Examples

```
# Here is an example on how to construct such a picsError object
# Typically, you would not do this manually, you would use the pics function to return a
# Contructor
myPicsError<-newPicsError("Singular information matrix")
# Accessors
# Get the standard error of Mu
se(myPicsError)
# Get the standard error of MuF
seF(myPicsError)
# Get the scores
score(myPicsError)
```

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`picsFDR`*Compute the global FDR*

---

**Description**

Calculate an estimate of the FDR for PICS. This calculation requires control data (e.g. from an input DNA sample).

**Usage**

```
picsFDR(picsIP, picsCont, filter=list(delta=c(0, Inf), se=c(0, Inf), sigmaSqF=c(0, Inf))
```

**Arguments**

<code>picsIP</code>	An object of class 'picsList' as returned by 'PICS' when run on IP/Control data.
<code>picsCont</code>	An object of class 'picsList' as returned by 'PICS' when run on Control/IP data.
<code>filter</code>	A list of filters to be used before computing the FDR. By default all regions are included. See details for more info on how to specify the filters.

**Value**

A dataframe with three columns corresponding to the estimated FDR, the score, and the number of regions.

**Author(s)**

Xuekui Zhang and Raphael Gottardo <<raphael.gottardo@ircm.qc.ca>>

**References**

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq" arXiv, 0903.3206, 2009. To appear in Biometrics.

**See Also**

[pics](#)

**Examples**

```
## Not run:
# Segment the reads in order to identify candidate regions
segC<-segmentReads(RD, minReads=3, invert=TRUE)
# Use the serial version of PICS
picsC<-PICS(segC, dataType='TF')
plot(pics, picsC, xlim=c(0, 50), ylim=c(0, .2), filter=list(delta=c(50, 300), se=c(0, 50), sigmaSqF=
## End(Not run)
```

---

`picsList-class`      *The pics class*

---

### Description

This object is used to gather all parameters from fitting PICS to multiple candidate regions (as returned by the 'segmentReads' function). The object contains the following slots: 'List', 'paraPrior', 'paraEM', 'minReads', 'N', 'Nc'. 'List' is a list of 'pics' or 'picsError' objects. 'paraPrior' is a list containing the hyperparameters used for the prior, 'paraEM' is a list of convergence parameters for the EM, 'minReads' is a list containing the minimum number of reads used to fit a region with 'PICS', 'N' is the total number of reads in the ChIP samples while 'Nc' is the total number of reads in the control sample.

### Arguments

`object`      An object of class `pics`.

### Accessors

The PICS package provide accessors to directly access to most of the parameters/standard errors and chromosomes. In the code snippets below, 'x' is a 'picsList' object. For all accessors, the 'picsError' objects are omitted, so that the accessors only return values for the 'pics' objects (i.e. all valid binding events).

'**chromosome(x)**' Gets the chromosome names of all candidate regions.

'**mu(x)**' Gets the position estimates of all binding sites identified in all candidate regions.

'**delta(x)**' Gets the average fragment lengths of all binding sites identified in all candidate regions.

'**sigmaSqF(x)**' Gets the F peak variances of all binding sites identified in all candidate regions.

'**sigmaSqR(x)**' Gets the R peak variances of all binding sites identified in all candidate regions.

'**seF(x)**' Gets the standard errors of all binding site position estimates identified in all candidate regions.

'**seF(x)**' Gets the standard errors of all F peak modes identified in all candidate regions.

'**seR(x)**' Gets the standard errors of all R peak modes identified in all candidate regions.

'**score(x)**' Gets the scores of all binding events identified in all candidate regions.

### Constructor

`newPicsList(List, paraEM, paraPrior, minReads, N, Nc)`

**List** The mixture weights (a vector)

**paraEM** The binding site positions (a vector)

**paraPrior** The DNA fragment lengths (a vector)

**N** The variance parameters for the forward distribution (vector)

**Nc** The variance parameters for the forward distribution (vector)

### Methods

[ `signature(x = "pics")` ]: subset PICS object.

**Methods**

**length** signature (x = "pics"): subset PICS object.

**Constructor**

`newPicsList`<-function(List, paraEM, paraPrior, minReads, N, Nc) constructs a new 'picsList' object with the following arguments.

**newPicsList**

**w** The mixture weights (a vector)

**mu** The binding site positions (a vector)

**delta** The DNA fragment lengths (a vector)

**sigmaSqF** The variance parameters for the forward distribution (vector)

**sigmaSqR** The variance parameters for the reverse distribution (vector)

**seMu** The standard errors for mu (vector)

**seMuF** The standard errors for muF (vector)

**seMuR** The standard errors for muR (vector)

**seMuR** The standard errors for muR (vector)

**score** The scores for each binding event (vector)

**Nmerged** The number of peaks that were merged (integer)

**converge** A logical value, TRUE, if the EM as converged

**infMat** The information matrix

**chr** The chromosome for the region

**Author(s)**

Xuekui Zhang <<xzhang@stat.ubc.ca>> and Raphael Gottardo <<raphael.gottardo@ircm.qc.ca>>

**References**

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq" arXiv, 0903.3206, 2009. To appear in Biometrics.

**See Also**

[pics](#)

**Examples**

```
# Here is an example of how to construct such a region
# Typically, you would not do this manually, you would use the pics function to return a
w<-1
mu<-10000
delta<-150
sigmaSqF<-5000
sigmaSqR<-5000
seMu<-10
seMuF<-10
seMuR<-10
score<-5
```

```

Nmerged<-0
converge<-TRUE
infMat<-matrix(0)
chr<-"chr1"
range<-c(1000,2000)
# Constructor
#myPICS1<-newPics(w,mu,delta,sigmaSqF,sigmaSqR,seMu,seMuF,seMuR,score,Nmerged,converge,infMat)
#myPICS2<-newPics(w,mu+1000,delta,sigmaSqF,sigmaSqR,seMu,seMuF,seMuR,score,Nmerged,converge,infMat)

#minReads<-list(perPeak=2,perRegion=5)
#paraPrior<-list(xi=200,rho=1,alpha=20,beta=40000)
#paraEM<-list(minK=1,maxK=15,tol=10e-6,B=100)
#N<-100
#Nc<-200

#mynewPicsList<-newPicsList(list(myPICS1,myPICS2), paraEM, paraPrior, minReads, as.integer(N), as.integer(Nc))
# Accessors
# Get the standard error of Mu
#se(mynewPicsList)
# Get the standard error of MuF
#seF(mynewPicsList)
# Get the scores
#score(mynewPicsList)

```

---

pics

*Estimation of binding site positions*

---

## Description

This object contains Estimation of binding site positions and has the following slots: `segReadsList`, `dataType`.

## Usage

```
PICS<-pics(segReadsList, dataType="TF")
```

## Arguments

`segReadsList` This object contains segmentation of Genome  
`dataType` The type of data you are processing: specified 'TF' for transcription factor.

## Methods

**code** signature(`x = "pics"`): return the error code for each list element (i.e. candidate region) of a PICS object. If the string is empty, there were no errors.

**plot** signature(`x = "pics"`): Plot all regions in the PICS object. This might be long, and should only be used to plot a few regions, so subset the object before plotting.

**sigmaSqR** signature(`x = "pics"`): return the variance parameter of the reverse (R) distribution for each binding event.

**sigmaSqF** signature(`x = "pics"`): return the variance parameter of the forward (F) distribution for each binding event.

**score** signature(`x = "pics"`): return the score for each binding event.

**scoreF** signature(x = "pics"): return the score of the forward (F) for each binding event.

**scoreR** signature(x = "pics"): return the score of the forward (R) for each binding event.

**maxRange** signature(x = "pics"): return the range maximum.

**minRange** signature(x = "pics"): return the range minimal.

**K** signature(x = "pics"): subset PICS object.

**density** signature(x = "pics"): return the density for each binding event.

### Author(s)

Xuekui Zhang, <ubcxzhang@gmail.com> and Raphael Gottardo, <raphael.gottardo@ircm.qc.ca>  
Arnaud Droit, <arnaud.droit@ircm.qc.ca>

### References

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq" arXiv, 0903.3206, 2009. To appear in Biometrics.

### See Also

[pics](#)

---

plot-FDR

*FDR plot for PICS*

---

### Description

This method plots an FDR curve showing the FDR as a function of the PICS scores.

### Usage

```
## S4 method for signature 'picsList,picsList':
plot(x, y, filter=NULL, h=.1, ...)
```

### Arguments

x	A <code>picsList</code> object as returned by the function <code>PICS</code> run on the treatment data.
y	A <code>picsList</code> object as returned by the function <code>PICS</code> run on the control data.
filter	A list of ranges for filtering regions based on PICS parameters. By default filter is set to 'NULL' and all regions are used.
h	A value between 0 and 1, representing the desired FDR. This simply draws a horizontal line at the given value.
...	Further graphical parameters passed to the generic function <code>plot</code> .

### Author(s)

Raphael Gottardo <<raph@rglab.org>>

**References**

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq," *Biometrics*, iss. In press, 2010.

**See Also**

[PICS](#)

---

segmentReads

*Segment the genome into candidate regions*

---

**Description**

Pre-process bidirectional aligned reads data from a single ChIP-Seq experiment to detect candidate regions with a minimum number of forward and reverse reads. These candidate regions will then be processed by PICS.

**Usage**

```
segmentReads(data, dataC=NULL, map=NULL, minReads=2, minReadsInRegion=3,
             jitter=FALSE, dataType="TF", maxLregion=0, minLregion=100)
```

**Arguments**

data	A ‘GenomeData’ object containing the IP reads. See details for more information on how to set up the data.
dataC	A ‘GenomeData’ object containing the control reads. Set to NULL by default, i.e. no control.
map	A ‘RangedData’ object containing the mappability profiles. Set to NULL by default, i.e. no profiles.
minReads	The minimum number of F/R reads to be present in the sliding window.
minReadsInRegion	The minimum number of F/R reads to be present in the region.
jitter	A logical value stating whether some noise should be added to the read locations. This is recommended if the read positions have lots of duplicates.
dataType	Type of experiment.
maxLregion	The maximum length.
minLregion	The minimum length.

**Value**

An object of class ‘segmentReadsList’ containing the results for all regions pre-processed.

**Author(s)**

Xuekui Zhang and Raphael Gottardo <<raphael.gottardo@ircm.qc.ca>>

## References

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq" arXiv, 0903.3206, 2009.

## See Also

segmentReads, picsFDR

## Examples

```
# Read data
path<-system.file("extdata",package="PICS")
## Note that the col name for the chromosome needs to be space and not chr
dataIP<-read.table(file.path(path,"Treatment_tags_chr21_sort.bed"),header=TRUE,colClasses=
dataIP<-as(dataIP,"RangedData")
dataIP<-as(dataIP,"GenomeData")

dataCont<-read.table(file.path(path,"Input_tags_chr21_sort.bed"),header=TRUE,colClasses=
dataCont<-as(dataCont,"RangedData")
dataCont<-as(dataCont,"GenomeData")

map<-read.table(file.path(path,"mapProfileShort"),header=TRUE,colClasses=c("factor","inte
map<-as(map,"RangedData")
## Remove the chrM
map<-map[-23]
seg<-segmentReads(dataIP, dataC=dataCont, map=map, minReads=1)
```

---

segReadsList

*Segment the genome into candidate regions*

---

## Description

Pre-process bidirectional aligned reads data from a single ChIP-Seq experiment to detect candidate regions with a minimum number of forward and reverse reads. These candidate regions will then be processed by PICS.

## Usage

```
segReadsList<-new(List, paraSW, N, Nc)
```

## Arguments

List	This object contains an ExpressionSet
paraSW	String containing the genome name used (vector).
N	String containing the name of chromosome used (vector).
Nc	String containing the Position of the sequences (vector).

## Methods

```
[ signature(x = "pics"): subset gadem object.
[ signature(x = "pics"): subset gadem object.
```

**Methods**

**length** signature(x = "pics"): subset PICS object.

**Author(s)**

Charles Cheung, <cykc@interchange.ubc.ca> and Raphael Gottardo, <raphael.gottardo@ircm.qc.ca>  
 Arnaud Droit, <arnaud.droit@ircm.qc.ca>

**References**

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq" arXiv, 0903.3206, 2009. To appear in Biometrics.

**See Also**

[pics](#)

---

 segReads

---

*Segment the genome into candidate regions*


---

**Description**

Pre-process bidirectional aligned reads data from a single ChIP-Seq experiment to detect candidate regions with a minimum number of forward and reverse reads. These candidate regions will then be processed by PICS.

**Usage**

```
segReads<-new(yF, yR, cF, cR, map, chr)
```

**Arguments**

yF	This object contains an ExpressionSet
yR	String containing the genome name used (vector).
cF	String containing the name of chromosome used (vector).
cR	String containing the Position of the sequences (vector).
map	String containing the copy number of sequence (vector).
chr	String containing the expression data of enriched region (matrix with n column).

**Methods**

**map** signature(x = "pics"): subset PICS object.

**Author(s)**

Charles Cheung, <cykc@interchange.ubc.ca> and Raphael Gottardo, <raphael.gottardo@ircm.qc.ca>  
 Arnaud Droit, <arnaud.droit@ircm.qc.ca>

## References

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq" arXiv, 0903.3206, 2009. To appear in Biometrics.

## See Also

[pics](#)

---

setParaEM

*Set convergence parameters of the EM algorithm*

---

## Description

This function can be used to change the internal PICS parameters for the EM algorithm. This function should only be called if you really now what you are doing!.

## Usage

```
setParaEM(minK=1, maxK=15, tol=1e-4, B=100, mSelect="BIC", mergePeaks=TRUE, mapCorrect
```

## Arguments

minK	An integer value. The minimum number of binding events per region.
maxK	An integer value. The maximum number of binding events per region.
tol	The tolerance for the EM algorithm
B	An integer value. The maximum number of iterations to be used.
mSelect	A character string specifying the information criteria to be used when selecting the number of binding events.
mergePeaks	A logical value stating whether overlapping binding events should be picked.
mapCorrect	Should mappability profiles be incorporated in the estimation, that is missing reads estimated.
dataType	A character string equal to either 'H' or 'TF', 'H' for histone and 'TF' for transcription factors.

## Value

No value returned. The function simply modifies the internal variables 'paraEMTF' if dataType='TF' and 'paraEMH' if dataType='H'.

## Author(s)

Xuekui Zhang and Raphael Gottardo <<raphael.gottardo@ircm.qc.ca>>

## References

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, "PICS: Probabilistic Inference for ChIP-seq" arXiv, 0903.3206, 2009. To appear in Biometrics.

**See Also**

setParaPrior

**Examples**

```
# Using mSelect="BIC"
setParaEM(minK=1,maxK=15,tol=1e-4,B=100,mSelect="BIC",mergePeaks=TRUE,mapCorrect=TRUE,dat
# Using mSelect="AIC"
setParaEM(minK=1,maxK=15,tol=1e-4,B=100,mSelect="AIC",mergePeaks=TRUE,mapCorrect=TRUE,dat
```

---

setParaPrior                      *Set convergence parameters of the EM algorithm*

---

**Description**

This function can be used to change the internal PICS parameters for the prior distribution. This function should only be called if you really now what you are doing! In particular, you may want to specify the average DNA fragment size for your sample by changing the ‘xi’ parameter.

**Usage**

```
setParaPrior(xi=200,rho=1,alpha=20,beta=40000,lambda=0,dMu=200,dataType="TF")
```

**Arguments**

xi	Our best guest for the average DNA fragment size.
rho	A variance parameter for the average DNA fragment size distribution.
alpha	First hyperparameter of the inverse Gamma distribution for $\sigma^2$ in the PICS model.
beta	First hyperparameter of the inverse Gamma distribution for $\sigma^2$ in the PICS model.
lambda	The precision of the prior for $\mu$ used for histone data.
dMu	Our best guess for the distance between two neighboring nucleosomes.
dataType	A character string equal to either ‘H’ or ‘TF’, ‘H’ for histone and ‘TF’ for transcription factors. ‘H’ is not yet supported

**Value**

No value returned. The function simply modifies the internal variables ‘paraPriorTF’ if dataType=‘TF’ and ‘paraPriorH’ if dataType=‘H’.

**Author(s)**

Xuekui Zhang and Raphael Gottardo <<raphael.gottardo@ircm.qc.ca>>

**References**

X. Zhang, G. Robertson, M. Krzywinski, K. Ning, A. Droit, S. Jones, and R. Gottardo, “PICS: Probabilistic Inference for ChIP-seq” arXiv, 0903.3206, 2009. To appear in Biometrics.

**See Also**

`setParaEM`

**Examples**

```
# Using xi=200 for the average DNA fragment size
setParaPrior(xi=200, rho=1, alpha=20, beta=40000, lambda=0, dMu=200, dataType="TF")
# Using xi=150 for the average DNA fragment size
setParaPrior(xi=150, rho=1, alpha=20, beta=40000, lambda=0, dMu=200, dataType="TF")
```

---

show

*show PICS*

---

**Description**

This methods show the objects of PICS

**Usage**

```
## S4 method for signature 'pics':
show(object)
## S4 method for signature 'picsError':
show(object)
## S4 method for signature 'picsList':
show(object)
## S4 method for signature 'segReads':
show(object)
## S4 method for signature 'segReadsList':
show(object)
```

**Arguments**

object            Object returned from `pics` .

**Details**

List of the slots include in the object

**Author(s)**

Xuekui Zhang <<xzhang@stat.ubc.ca> Arnaud Droit, <arnaud.droit@ircm.qc.ca>  
Raphael Gottardo <<raphael.gottardo@ircm.qc.ca>

**See Also**

[summary](#)

---

 summary

*summary PICS*


---

**Description**

This methods summarized ‘pics’, ‘picsList’, ‘seg’ or ‘segList’ objects.

**Usage**

```
## S4 method for signature 'pics':
summary(object)
## S4 method for signature 'picsList':
summary(object)
## S4 method for signature 'segReads':
summary(object)
## S4 method for signature 'segReadsList':
summary(object)
```

**Arguments**

object            Object returned from [pics](#) .

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**See Also**

[show](#)

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unique

*GenomeData Unique Reads*


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**Description**

This methods select the unique elements in a GenomeData object

**Arguments**

object            A ‘GenomeData’ object.

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**See Also**

[segmentReads](#)

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