

# Rolexa: Probabilistic Base Calling of Solexa Sequencing Data

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## 1 Introduction

This package provides an alternative base calling algorithm using model-based clustering (*mclust*) and probability theory to identify ambiguous bases and code them with IUPAC symbols. We also select optimal sub-tags using a score based on information content to remove uncertain bases towards the ends of the reads. There are also a few diagnostic plots functionalities. Details of the algorithms were published in [1].

## 2 Environment variables

The *Rolexa* package uses a `RolexaRun` object to store the various parameters of the run, and uses the *ShortRead* for manipulating data, in particular many *Rolexa* functions take a `SolexaPath` object as argument.

We load the library and create a configuration with default parameters except for the `idsep` variable:

```
> library(Rolexa)
```

by using `mclust`, invoked on its own or through another package, you accept the license agreement in the `mclust LICENSE` file and at <http://www.stat.washington.edu/mclust/license.txt>

```
> rolenv = SetModel(idsep = "_")
```

```
> GetModel(rolenv)
```

```
$MinimumTagLength
```

```
[1] 15
```

```
$SequencingLength
```

```
[1] 36
```

```
$Barcode
```

```
[1] 0
```

```
$HThresholds
```

```
[1] 0.5849625 1.3219281 1.8073549
```

```
$IThresholds
```

```
[1] 2.058894 2.115477 2.169925 2.222392 2.273018 2.321928 2.369234 2.415037  
[9] 2.459432 2.502500 2.544321 2.584963 2.624491 2.662965 2.700440 2.736966  
[17] 2.772590 2.807355 2.841302 2.874469 2.906891 2.938599 2.969626 3.000000  
[25] 3.029747 3.058894 3.087463 3.115477 3.142958 3.169925 3.196397 3.222392  
[33] 3.247928 3.273018 3.297681 3.321928
```

```
$PET
```

```
[1] FALSE
```

```
$fit
```

```
[1] FALSE
```

```
$normal
```

```
[1] TRUE
```

```
$decorrelate
```

```
[1] "both"
```

```
$verbose
```

```
[1] 0
```

```
$colors
```

```
[1] "black"      "green"      "blue"      "chocolate3" "red"
```

```
[6] "#007F7F"      "#66B20E"      "#7F7F00"      "#66338E"      "#7F007F"
[11] "#E6330E"      "#7F464E"      "#7F6035"      "#6C5649"      "#685F4C"
[16] "gray"
```

```
$idsep
[1] "_"
```

The meaning of these parameters is as follows:

**MinimumTagLength** tags shorter than this will not be saved

**SequencingLength** number of sequencing cycles, used to calculate the number of columns in files

**Barcode** number of bases used as barcode at the beginning of the tag

**HThresholds** entropy thresholds between 1 and 2-base ambiguities, 2 and 3-base ambiguities and 3-base ambiguity or undecided (the default is  $\log_2(c(1.5, 2.5, 3.5))$ )

**IThresholds** total entropy thresholds, as a function of tag length (the default is  $\log_2(4 + 1 : 36/6)$ )

**PET** paired-end sequencing run

**fit** use full EM convergence instead of only one-step optimization if TRUE

**normal** use tile-level normalization before base-calling if TRUE

**decorrelate** use 'cycle'-level decorrelation procedure, 'channel'-level, 'both' or 'none'

**idsep** character separating coordinate fields in sequence headers (default is ":",")

**verbose** print debug information if > 0

### 3 Loading data

Loading data is done using the *ShortRead* utilities (in particular the `SolexaPath` class) with two additional wrappers `CombineReads` and `CombineFastQ`:

```
> path = SolexaPath(system.file("extdata", package = "ShortRead"))
```

Then use the loading functions to read a selection of those files:

```
> (int = readIntensities(path, pattern = "s_1_0001", withVariability = FALSE))
```

```
class: SolexaIntensity
dim: 256 4 36
readInfo: SolexaIntensityInfo
intensity: ArrayIntensity
measurementError: not available
```

```
> (seq = CombineReads(run = rolenv, path = path, pattern = "s_1_0001_seq*"))
```

```
class: ShortRead
length: 256 reads; width: 36 cycles
```

```
> (seq_fastq = readFastq(path))
```

```
class: ShortReadQ
length: 256 reads; width: 36 cycles
```

## 4 Data transforms

Before going into the base calling itself, we can perform several data transformations to remove some of the systematic biases:

1. Reduce cross-talk between color channels

```
> (theta = OptimizeAngle(int = int))[1:10, ]
      [,1]      [,2]      [,3]      [,4]
[1,] 0.7767119 1.375080 0.4721182 1.557188
[2,] 0.7653824 1.377907 0.5618510 1.570796
[3,] 0.7276859 1.367992 0.5290140 1.570796
[4,] 0.7551378 1.384266 0.6453509 1.570796
[5,] 0.7349694 1.377229 0.6220983 1.570796
[6,] 0.7377151 1.383378 0.6556697 1.564773
[7,] 0.7213154 1.377866 0.6412864 1.570796
[8,] 0.7685749 1.384597 0.6472642 1.570796
[9,] 0.7681729 1.387350 0.5537521 1.570796
[10,] 0.7710965 1.379977 0.6961033 1.570796

> int = DeCorrelateChannels(int = int, theta = theta)
```

2. Reduce dephasing along cycles

```
> (rate = OptimizeRate(int = int))
[1] 0.01760222

> int = DeCorrelateCycles(int = int, rate = rate)
```

3. Reduce position-dependent bias within each tile

```
> int2 = TileNormalize(run = rolenv, int = int)
```

## 5 Base calling

The base calling algorithm fits a gaussian mixture model to the four-dimensional intensity values from each cycle. Sequences from a previous base calling, if available, are used to seed the algorithm:

```
> (res = SeqScore(run = rolenv, int = int, seqInit = seq, cycles = 1:36))$sread
```

```
A DNASTringSet instance of length 256
  width seq
[1] 36 TTGTTTTTCATGTGATTTTAAAAATGTATTTGTTTGT
[2] 36 TCCAAACTGGTAGACAATACAAACATTCTCAAATCT
[3] 36 TGCACCTGATAGGGTCTCTGCTCTGAGAGAGDAAGK
[4] 36 TATGAGAGTAGCYAATGCCACAAAGWSGRKGTGKBY
[5] 36 TAGTAGGTGTCCTATTCTGATGCYCAGCACGCCAAG
[6] 36 GAGAGAACTGAAAATCACAGAATATGAGAAATAGAC
[7] 36 GCAGAGACCCACAASCCAGCCAAGCGGCTCCWGACC
[8] 36 GAGATATTTATTGAACACTAACACTCTGTCATGCAA
[9] 36 GGTGAAAAGWAGGAAGCAYCCCSYTYTCYGCTTAYAT
... ..
[248] 36 TGGGGAGMYGKGGGMYMTGGCKGGMRYRTHRWVVDK
[249] 36 GTGGAGGCTAGCACCTGTTTGTGGCBTTGTGARGBA
[250] 36 GATTTTCAAAGTTAAGGGTAAAAATGTTATCACCCG
[251] 36 GAAAATGAGAAACATACAATTGACGACTTGAAAAAT
[252] 36 GGYATTTTCCTTTTGTTTTATTTMRCTTTGKWGBDH
[253] 36 GGTAGGRAGAGCTCGTGKCCGTCTTCTGCTTRRAW
[254] 36 GAAAAACGWGAAAAATGAGAAWTGCACACTGTAGRA
[255] 36 GATTCCTTATGTGGTAATGGAATAATATTTTCATC
[256] 36 GGATGAGAAGAATAGTATATTACATCTCTAGCCACA
```

## 6 Filtering and saving

The base calling results consist of a full-length tag with base quality entropy scores, which can then be filtered to extract the best sequence tag for each colony. This is where the parameters `IThresholds` comes into play:

```
> rolenv@MinimumTagLength = as.integer(1)
> (res2 = FilterResults(run = rolenv, results = res))$sread
```

```
A DNASTringSet instance of length 256
  width seq
[1] 36 TTGTTTTTCATGTGATTTTAAAAATGTATTTGTTTGT
[2] 36 TCCAAACTGGTAGACAATACAAACATTCTCAAATCT
[3] 36 TGCACCTGATAGGGTCTCTGCTCTGAGAGAGDAAGK
[4] 28 TATGAGAGTAGCYAATGCCACAAAGWSG
[5] 36 TAGTAGGTGTCCTATTCTGATGCYCAGCACGCCAAG
[6] 36 GAGAGAACTGAAAATCACAGAATATGAGAAATAGAC
[7] 36 GCAGAGACCCACAASCCAGCCAAGCGGCTCCWGACC
[8] 36 GAGATATTTATTGAACACTAACACTCTGTCATGCAA
[9] 21 GGTGAAAAGWAGGAAGCAYCCC
... ..
[248] 10 TGGGGAGMYG
[249] 34 GTGGAGGCTAGCACCTGTTTGTGGCBTTGTGARG
```

```

[250] 36 GATTTTCAAAGTTAAGGGTAAAAATGTTATCACCCG
[251] 36 GAAAATGAGAAACATACAATTGACGACTTGAAAAAT
[252] 30 GGYATTTTCCTTTTGTTTATTTMRCTTTG
[253] 33 GGTAGGRAGAGCTCGTGKCCGTCTTCTGCTTR
[254] 36 GAAAAACGWGAAAAATGAGAAWTGCACACTGTAGRA
[255] 36 GATTCCTTATGTGGTAATGAAAAATAATATTTTCATC
[256] 36 GGATGAGAAGAATAGTATATTACATCTCTAGCCACA

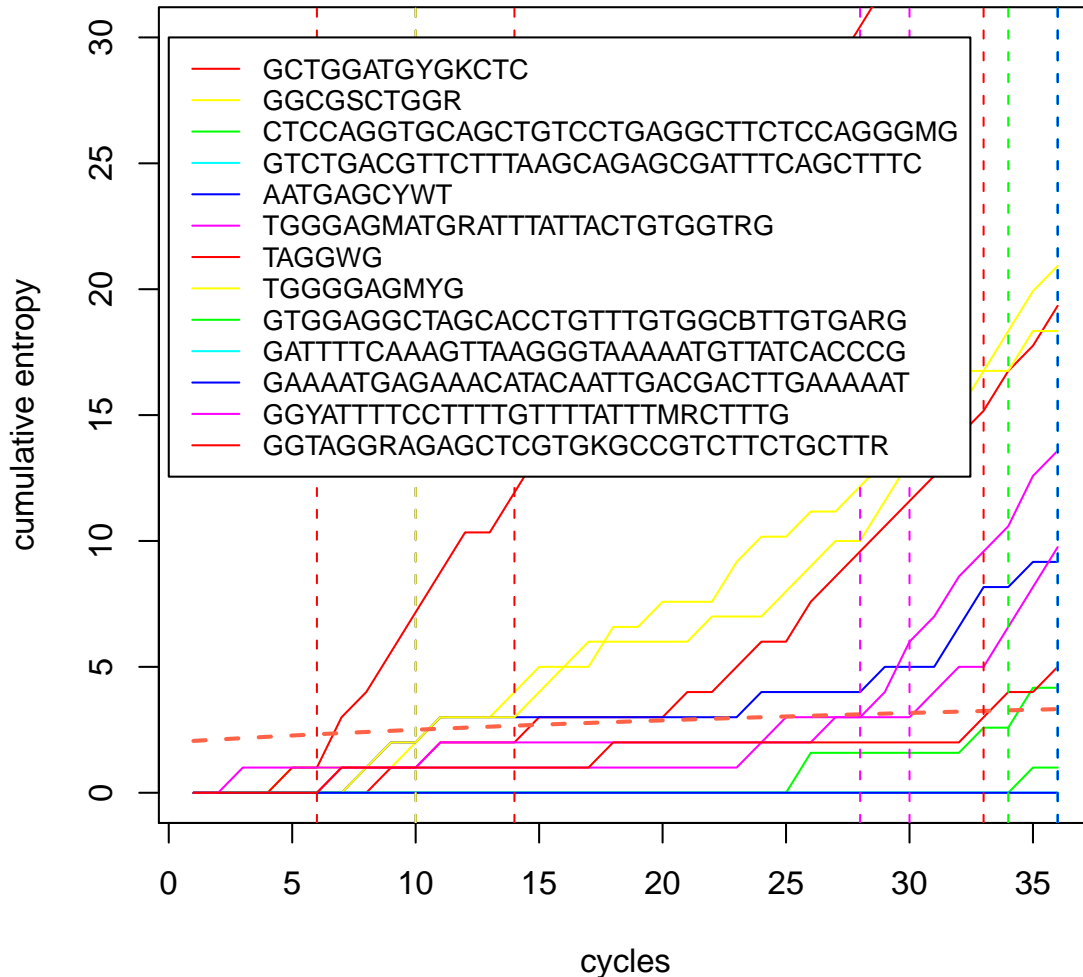
```

```

> str = as.matrix(res$sread[241:253])
> nt = DNA_ALPHABET
> post.entropy = matrix(0, nrow = nrow(str), ncol = 36)
> post.entropy[which(str %in% nt[5:10])] = 1
> post.entropy[which(str %in% nt[11:14])] = log2(3)
> post.entropy[which(str == "N")] = 2
> matplot(1:36, y = apply(post.entropy, 1, cumsum), t = "l", lty = 1,
+       col = rainbow(6), ylim = c(0, 30), xlim = c(1, 36), xlab = "cycles",
+       ylab = "cumulative entropy", main = "Tag length cutoff")
> lines(1:36, rolenv@IThresholds, t = "l", lty = 2, lwd = 2, col = "tomato")
> abline(v = nchar(res2$sread[241:253]), col = rainbow(6), lty = 2)
> legend(x = 0, y = 30, res2$sread[241:253], col = rainbow(6),
+       lty = 1, bg = "white", cex = 0.8)

```

## Tag length cutoff



The final step is to save results:

```
> SaveResults(run = rolenv, results = res2, outpath = "./")
```

## 7 Batch execution

The whole sequence of operations needed to load, analyse, filter and save a sequencing run can be performed in parallel (using calls to the *fork* package) via the function `ForkBatch`:

```
> library(fork)
> ForkBatch(run = rolenv, path = path, outpath = "./", prefix = "rs_",
+   nthreads = 2, nfiles = 5, lane = 1, tiles = 1, idsep = "_")
```

Each of the `nthreads` threads will execute a call to

```
> OneBatch(run = rolenv, path = path, lane = 1, tiles = tiles[n:m],  
+         outpath = "./", prefix = "rs_")
```

This function can be used in a loop on single-processor systems or in independent jobs distributed on a computing cluster.

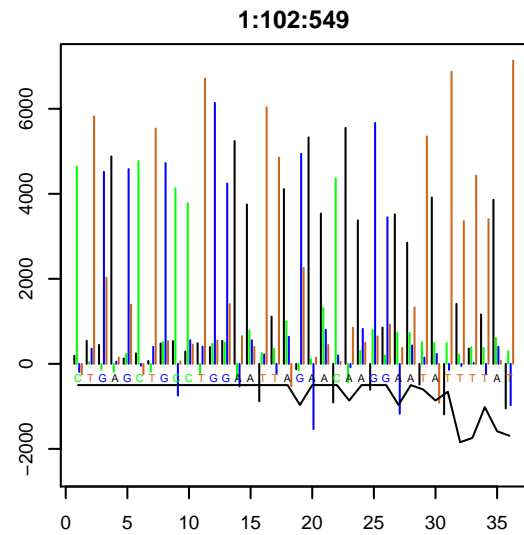
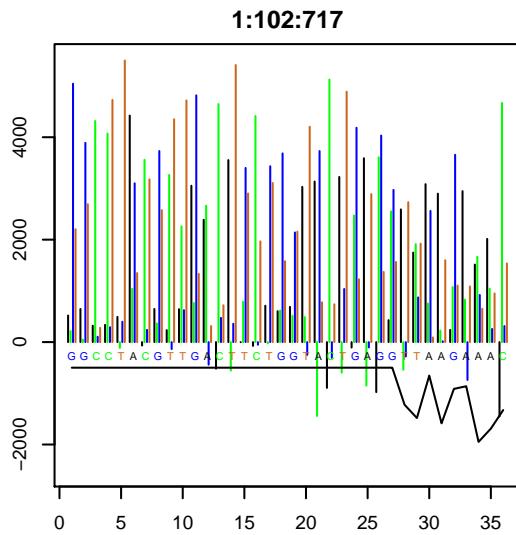
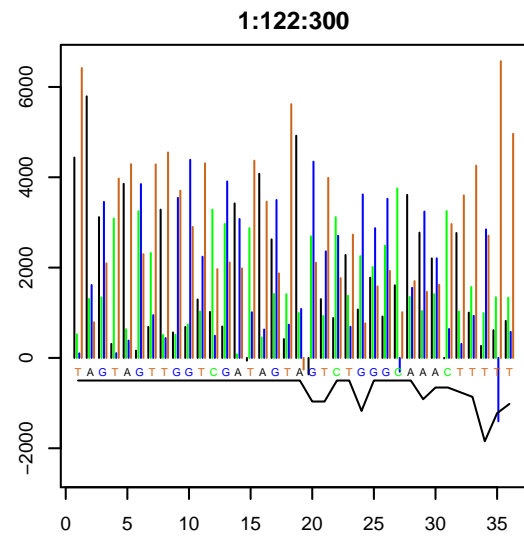
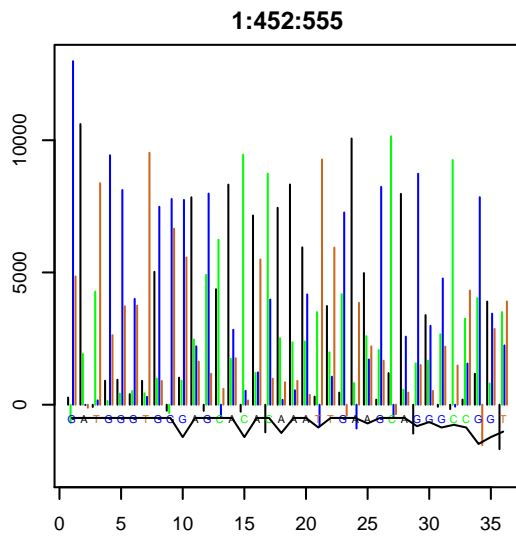
## 8 Diagnostic plots

There are multiple possibilities for evaluating the quality of the base calling, at the level of each sequence, tile or lane.

Given a sequence tag, the corresponding raw intensities and a base quality score, we can use `CombinedPlot`:

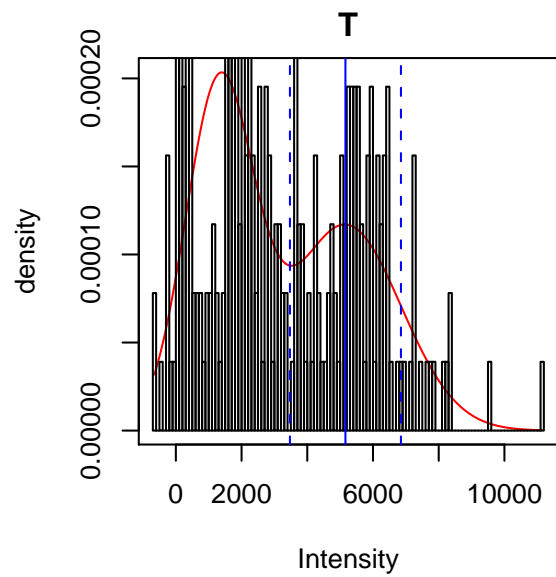
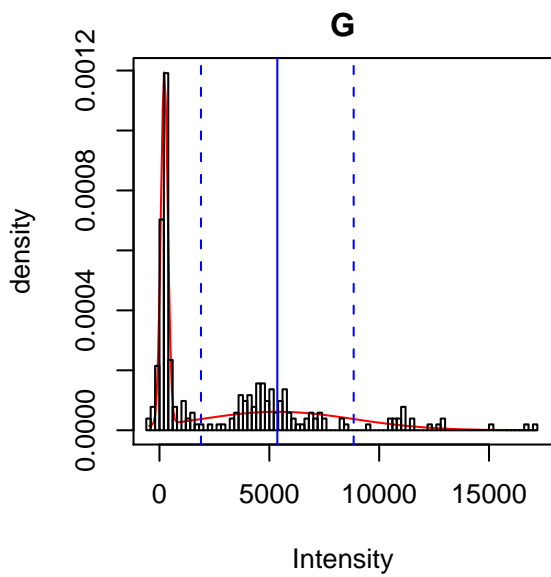
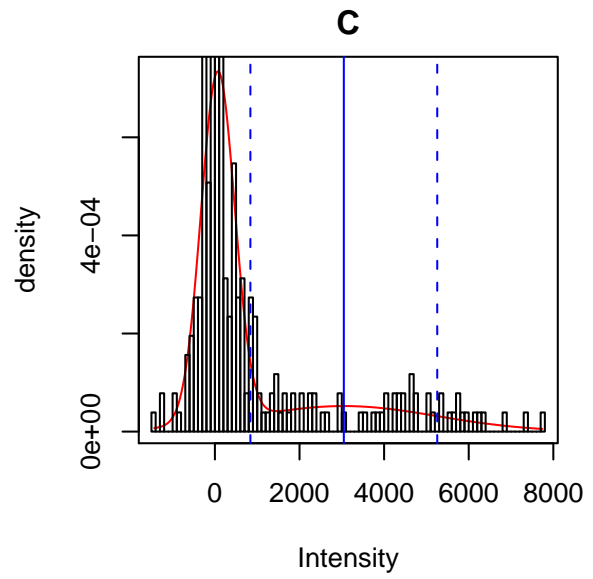
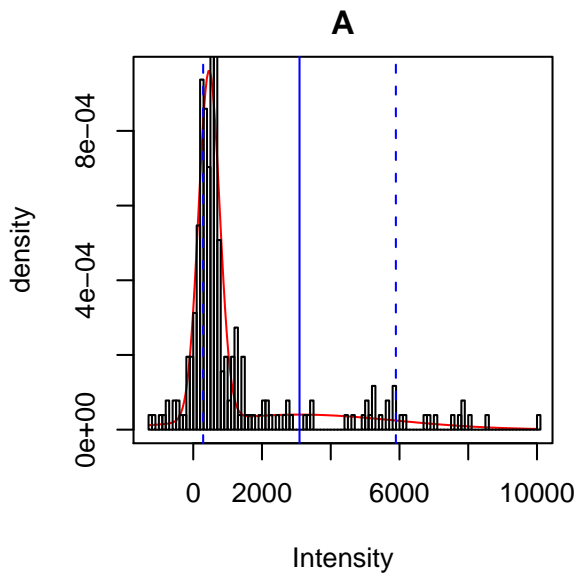
```
> CombinedPlot(run = rolenv, int = int, seq = seq, scores = as(quality(seq_fastq),  
+         "matrix"), colonies = sample(1:nrow(int), 4), par = list(mfrow = c(2,  
+         2), cex = 0.6, mar = c(4, 4, 2, 1) + 0.1))
```



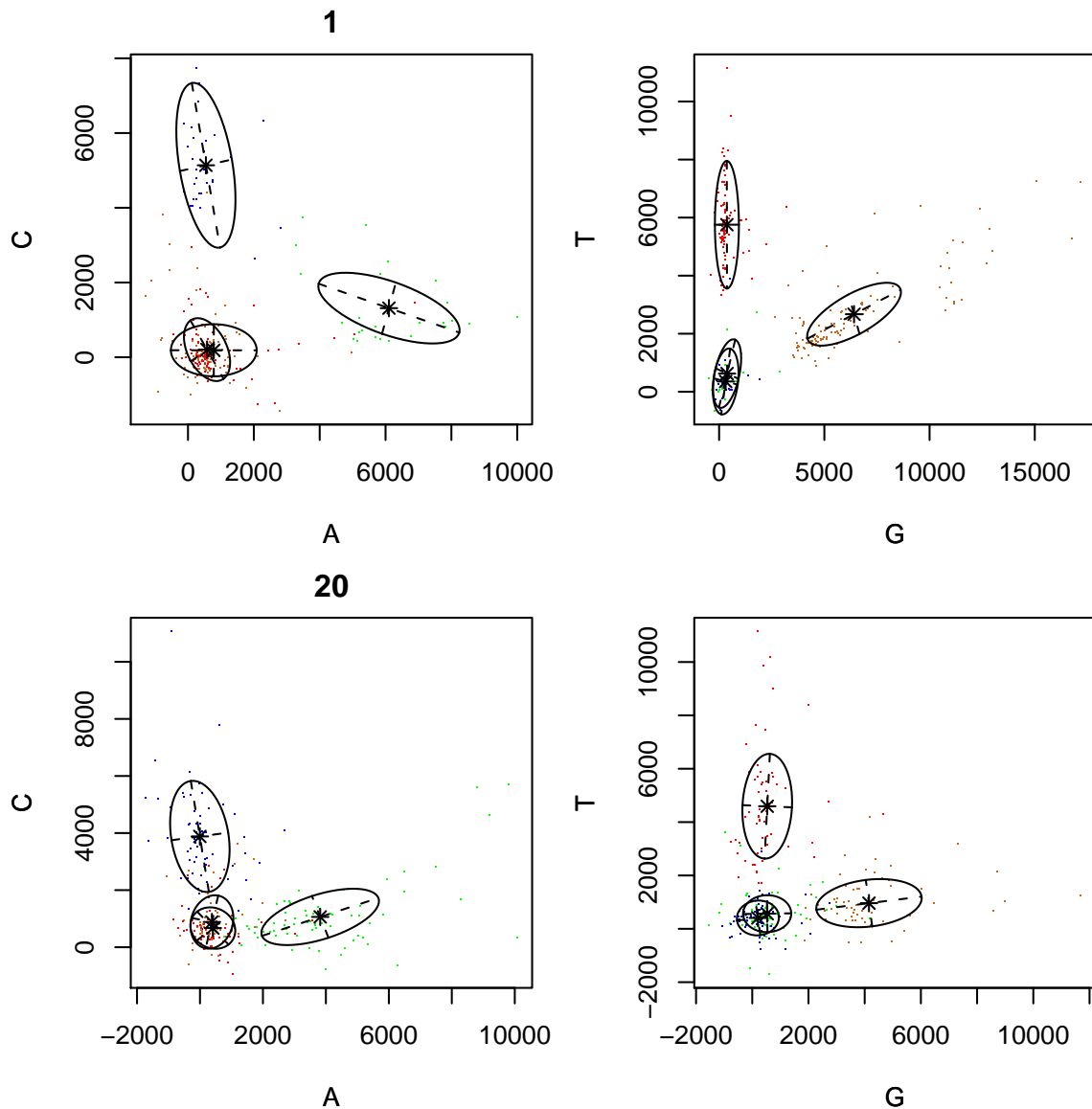


we can also evaluate the distribution of intensity values at selected cycles via 1- and 2-dimensional projections:

```
> ChannelHistogram(int = int, cycles = 1, par = list(mfrow = c(2,
+ 2), mar = c(4, 4, 2, 1) + 0.1))
```



```
> par(mfrow = c(2, 2), mar = c(4, 4, 2, 1) + 0.1)
> PlotCycles(run = rolenv, int = int, seq = seq, cycles = c(1,
+ 20))
```

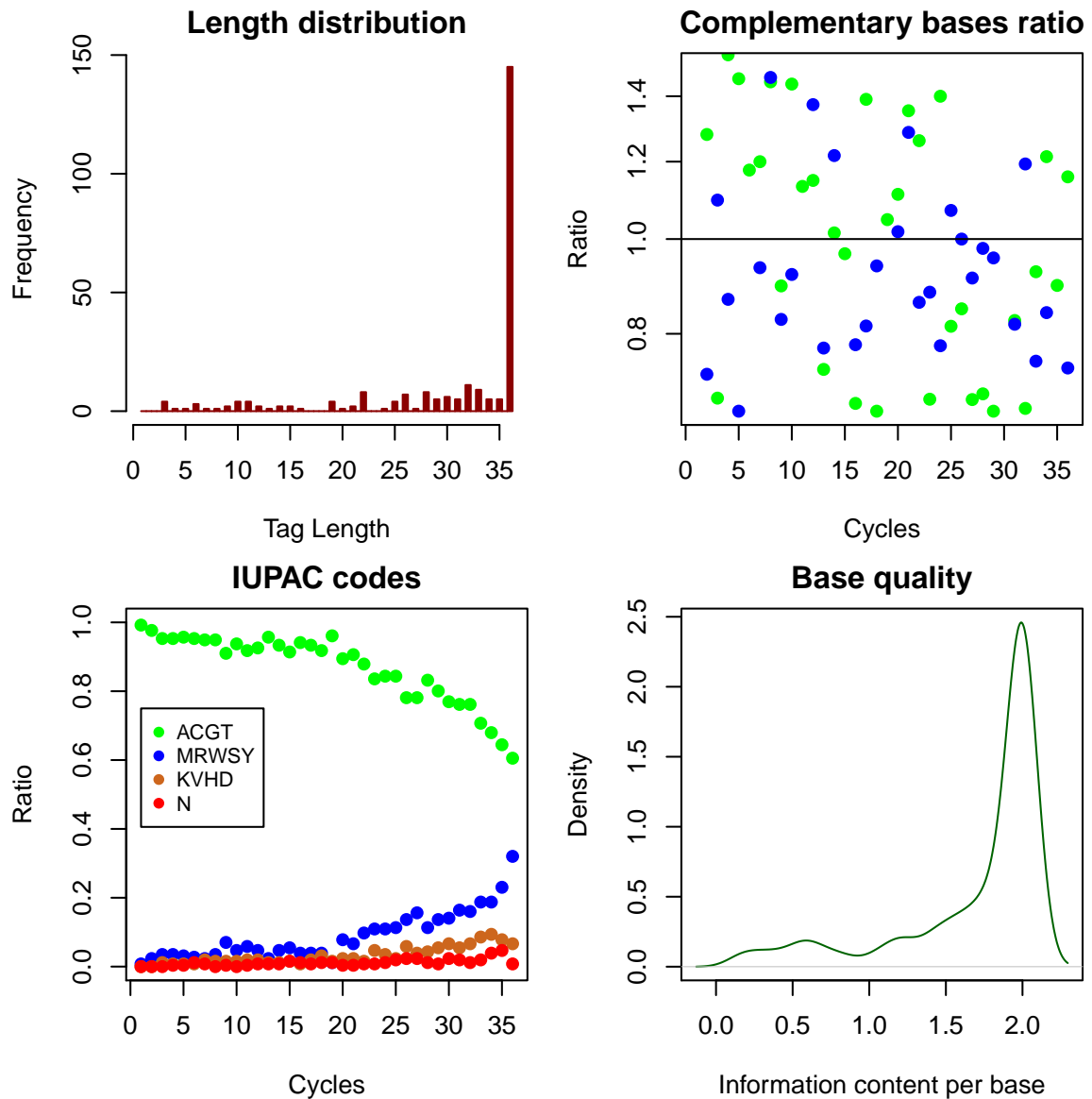


and look at global statistics of a base-calling:

```

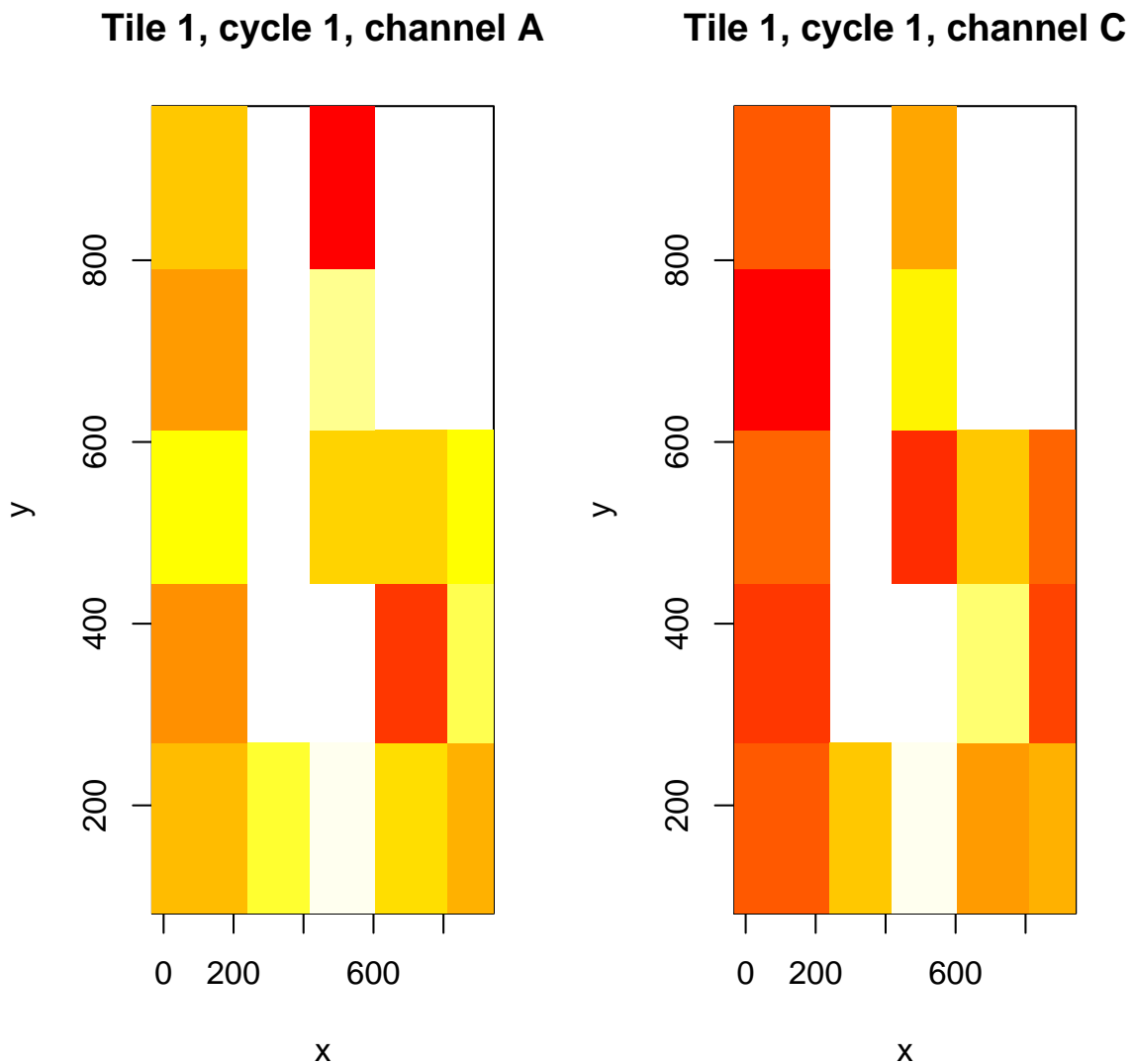
> par(mfrow = c(2, 2), cex = 0.8, mar = c(4, 4, 2, 1) + 0.1)
> BatchAnalysis(run = rolenv, seq = res2$sread, scores = res2$entropy,
+   what = "length", main = "Length distribution")
> BatchAnalysis(run = rolenv, seq = res$sread, scores = res$entropy,
+   what = "ratio", main = "Complementary bases ratio")
> BatchAnalysis(run = rolenv, seq = res$sread, scores = res$entropy,
+   what = "iupac", main = "IUPAC codes")
> BatchAnalysis(run = rolenv, seq = res2$sread, scores = res2$entropy,
+   what = "information", main = "Base quality")

```



and visualize the positional bias over a tile by

```
> par(mfrow = c(1, 2))
> TileImage(int = int, cycle = 1, tile = readInfo(int)$tile[1],
+   ncell = 5, channel = "A")
> TileImage(int = int, cycle = 1, tile = readInfo(int)$tile[1],
+   ncell = 5, channel = "C")
```



## 9 Session Information

The version number of R and packages loaded for generating the vignette were:

```
> toLatex(sessionInfo())
```

- R version 2.11.0 (2010-04-22), x86\_64-unknown-linux-gnu
- Locale: LC\_CTYPE=en\_US, LC\_NUMERIC=C, LC\_TIME=en\_US, LC\_COLLATE=en\_US, LC\_MONETARY=C, LC\_MESSAGES=en\_US, LC\_PAPER=en\_US, LC\_NAME=C, LC\_ADDRESS=C, LC\_TELEPHONE=C, LC\_MEASUREMENT=en\_US, LC\_IDENTIFICATION=C
- Base packages: base, datasets, graphics, grDevices, methods, stats, tools, utils

- Other packages: Biostrings 2.16.0, GenomicRanges 1.0.0, IRanges 1.6.0, lattice 0.18-5, mclust 3.4.4, Rolexa 1.4.0, Rsamtools 1.0.0, ShortRead 1.6.0
- Loaded via a namespace (and not attached): Biobase 2.8.0, grid 2.11.0, hwriter 1.2

## References

- [1] Jacques Rougemont, Arnaud Amzallag, Christian Iseli, Laurent Farinelli, Ioannis Xenarios, and Felix Naef. Probabilistic base calling of Solexa sequencing data. *BMC Bioinformatics*, **9**:431, 2008.