# Package 'ADImpute'

July 27, 2025

Type Package

**Title** Adaptive Dropout Imputer (ADImpute)

**Version** 1.18.0

Description Single-cell RNA sequencing (scRNA-seq) methods are typically unable to quantify the expression levels of all genes in a cell, creating a need for the computational prediction of missing values ('dropout imputation'). Most existing dropout imputation methods are limited in the sense that they exclusively use the scRNA-seq dataset at hand and do not exploit external gene-gene relationship information. Here we propose two novel methods: a gene regulatory network-based approach using gene-gene relationships learnt from external data and a baseline approach corresponding to a sample-wide average. ADImpute can implement these novel methods and also combine them with existing imputation methods (currently supported: DrImpute, SAVER). ADImpute can learn the best performing method per gene and combine the results from different methods into an ensemble.

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**Encoding UTF-8** 

LazyData true

**Depends** R (>= 4.0)

**Imports** checkmate, BiocParallel, data.table, DrImpute, kernlab, MASS, Matrix, methods, rsvd, S4Vectors, SAVER, SingleCellExperiment, stats, SummarizedExperiment, utils

Suggests BiocStyle, knitr, rmarkdown, testthat

**biocViews** GeneExpression, Network, Preprocessing, Sequencing, SingleCell, Transcriptomics

RoxygenNote 7.1.1 VignetteBuilder knitr

BugReports https://github.com/anacarolinaleote/ADImpute/issues

git\_url https://git.bioconductor.org/packages/ADImpute

git\_branch RELEASE\_3\_21
git\_last\_commit e66afe6

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git_last_commit_date 2025-04-15
Repository Bioconductor 3.21
Date/Publication 2025-07-27
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# Description

ArrangeData finds common genes to the network and provided data and limits both datasets to these

#### Usage

```
ArrangeData(data, net.coef = NULL)
```

# **Arguments**

data matrix with entries equal to zero to be imputed (genes as rows and samples as

columns)

net.coef matrix; object containing network coefficients

#### Value

list; data matrix, network coefficients matrix and intercept for genes common between the data matrix and the network

CenterData Data centering

#### **Description**

CenterData centers expression of each gene at 0

# Usage

CenterData(data)

# **Arguments**

data matrix of gene expression to be centered row-wise (genes as rows and samples

as columns)

# Value

list; row-wise centers and centered data

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CheckArguments\_Impute Argument check to Impute()

# **Description**

CheckArguments\_Impute checks whether the arguments passed to Impute are correct.

#### Usage

```
CheckArguments_Impute(data, method.choice, do, tr.length, labels, cell.clusters, true.zero.thr, drop_thre)
```

#### **Arguments**

data matrix; raw counts (genes as rows and samples as columns)
method.choice character; best performing method in training data for each gene

do character; choice of methods to be used for imputation. Currently supported

methods are 'Baseline', 'DrImpute', 'Network', and 'Ensemble'. Defaults to 'Ensemble'. Not case-sensitive. Can include one or more methods. Non-

supported methods will be ignored.

tr.length matrix with at least 2 columns: 'hgnc\_symbol' and 'transcript\_length' labels character; vector specifying the cell type of each column of data

cell.clusters integer; number of cell subpopulations

true.zero.thr if set to NULL (default), no true zero estimation is performed. Set to numeric

value between 0 and 1 for estimation. Value corresponds to the threshold used to determine true zeros: if the probability of dropout is lower than true.zero.thr,

the imputed entries are set to zero.

drop\_thre numeric; between 0 and 1 specifying the threshold to determine dropout values

#### Value

NULL object

ChooseMethod	Method choice per gene

#### **Description**

ChooseMethod determines the method for dropout imputation based on performance on each gene in training data

#### Usage

```
ChooseMethod(real, masked, imputed, write.to.file = TRUE)
```

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#### **Arguments**

real matrix; original gene expression data, i.e. before masking (genes as rows and

samples as columns)

masked matrix, logical indicating which entries were masked (genes as rows and sam-

ples as columns)

imputed list; list of matrices with imputation results for all considered methods

write.to.file logical; should the output be written to a file?

#### **Details**

The imputed values are compared to the real ones for every masked entry in real. The Mean Squared Error is computed for all masked entries per gene and the method with the best performance is chosen for each gene.

#### Value

character; best performing method in the training set for each gene

#### See Also

ComputeMSEGenewise

Combine	Combine imputation methods	
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#### **Description**

Combine imputation methods

#### Usage

```
Combine(data, imputed, method.choice, write = FALSE)
```

#### **Arguments**

data matrix with entries equal to zero to be imputed, already normalized (genes as

rows and samples as columns)

imputed list; list of matrices with imputation results for all considered methods method.choice named character; vector with the best performing method per gene write logical; should a file with the imputation results be written?

#### **Details**

Combines imputation results from all methods according to training results provided in method.choice

#### Value

matrix; imputation results combining the best performing method per gene

6 CreateArgCheck

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# **Description**

ComputeMSEGenewise computes the MSE of dropout imputation for a given gene.

# Usage

```
ComputeMSEGenewise(real, masked, imputed, baseline)
```

# Arguments

real numeric; vector of original expression of a given gene (before masking)
masked logical; vector indicating which entries were masked for a given gene

imputed matrix; imputation results for a given imputation method

baseline logical; is this baseline imputation?

#### Value

MSE of all imputations indicated by masked

t check
t ch

# **Description**

CreateArgCheck creates tests for argument correctness.

# Usage

```
CreateArgCheck(missing = NULL, match = NULL, acceptable = NULL,
null = NULL)
```

# **Arguments**

missing	named list; logical. Name corresponds to variable name, and corresponding entry to whether it was missing from the function call.
match	named list. Name corresponds to variable name, and corresponding entry to its value.
acceptable	named list. Name corresponds to variable name, and corresponding entry to its acceptable values.
null	named list; logical. Name corresponds to variable name, and corresponding entry to whether it was NULL in the function call.

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#### Value

argument check object.

CreateTrainData

Preparation of training data for method evaluation

#### **Description**

CreateTrainingData selects a subset of cells to use as training set and sets a portion (mask) of the non-zero entries in each row of the subset to zero

# Usage

```
CreateTrainData(data, train.ratio = .7, train.only = TRUE, mask = .1,
write = FALSE)
```

# **Arguments**

data matrix; raw counts (genes as rows and samples as columns)

train.ratio numeric; ratio of the samples to be used for training

train.only logical; if TRUE define only a training dataset, if FALSE writes both training

and validation sets (defaults to TRUE)

mask numeric; ratio of total non-zero samples to be masked per gene (defaults to .1)

write logical; should the output be written to a file?

#### Value

list with resulting matrix after subsetting and after masking

DataCheck\_Matrix Data check (matrix)

# Description

DataCheck\_Matrix tests for potential format and storage issues with matrices. Helper function to ADImpute.

#### Usage

```
DataCheck_Matrix(data)
```

#### **Arguments**

data object to check

#### Value

data object with needed adjustments

DataCheck\_Network

Data check (network)

# **Description**

DataCheck\_Network tests for potential format and storage issues with the network coefficient matrix. Helper function to ADImpute.

# Usage

DataCheck\_Network(network)

# **Arguments**

network

data object containing matrix coefficients

#### Value

network data object with needed adjustments

DataCheck\_SingleCellExperiment

Data check (SingleCellExperiment)

# Description

DataCheck\_SingleCellExperiment tests for existence of the appropriate assays in sce. Helper function to ADImpute.

# Usage

DataCheck\_SingleCellExperiment(sce, normalized = TRUE)

#### **Arguments**

sce

SingleCellExperiment; data for normalization or imputation

normalized

logical; is the data expected to be normalized?

#### Value

NULL object.

DataCheck\_TrLength 9

DataCheck\_TrLength

Data check (transcript length)

# **Description**

DataCheck\_TrLength tests for potential format and storage issues with the object encoding transcript length, for e.g. TPM normalization. Helper function to ADImpute.

#### Usage

```
DataCheck_TrLength(trlength)
```

# **Arguments**

trlength

data object containing transcript length information

#### Value

transcript length object with needed adjustments

demo\_data

Small dataset for example purposes

# Description

A small dataset to use on vignettes and examples (50 cells).

# Usage

demo\_data

#### **Format**

matrix; a subset of the Grun pancreas dataset, obtained with the scRNAseq R package, to use in the vignette and examples.

# References

Grun D et al. (2016). De novo prediction of stem cell identity using single-cell transcriptome data. Cell Stem Cell 19(2), 266-277.

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demo\_net

Small regulatory network for example purposes

# Description

Subset of the Gene Regulatory Network used by ADImpute's Network imputation method.

# Usage

demo\_net

# **Format**

matrix; subset of the Gene Regulatory Network installed along with ADImpute.

demo\_sce

Small dataset for example purposes

# Description

A small dataset to use on vignettes and examples (50 cells).

# Usage

demo\_sce

# **Format**

SingleCellExperiment; a subset of the Grun pancreas dataset, obtained with the scRNAseq R package, to use in the vignette and examples.

# References

Grun D et al. (2016). De novo prediction of stem cell identity using single-cell transcriptome data. Cell Stem Cell 19(2), 266-277.

EvaluateMethods 11

EvaluateMethods Imputation method evaluation on training set	EvaluateMethods	Imputation method evaluation on training set	
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# **Description**

EvaluateMethods returns the best-performing imputation method for each gene in the dataset

# Usage

```
EvaluateMethods(data, sce = NULL, do = c('Baseline', 'DrImpute',
'Network'), write = FALSE, train.ratio = .7, train.only = TRUE,
mask.ratio = .1, outdir = getwd(), scale = 1, pseudo.count = 1,
labels = NULL, cell.clusters = 2, drop_thre = NULL, type = 'count',
cores = BiocParallel::bpworkers(BPPARAM),
BPPARAM = BiocParallel::SnowParam(type = "SOCK"),
net.coef = ADImpute::network.coefficients, net.implementation = 'iteration',
tr.length = ADImpute::transcript_length, bulk = NULL, ...)
```

# **Arguments**

data	matrix; normalized counts, not logged (genes as rows and samples as columns)
sce	SingleCellExperiment; normalized counts and associated metadata.
do	character; choice of methods to be used for imputation. Currently supported methods are 'Baseline', 'DrImpute' and 'Network'. Not case-sensitive. Can include one or more methods. Non- supported methods will be ignored.
write	logical; write intermediary and imputed objects to files?
train.ratio	numeric; ratio of samples to be used for training
train.only	logical; if TRUE define only a training dataset, if FALSE writes and returns both training and validation sets (defaults to TRUE)
mask.ratio	numeric; ratio of samples to be masked per gene
outdir	character; path to directory where output files are written. Defaults to working directory
scale	integer; scaling factor to divide all expression levels by (defaults to 1)
pseudo.count	integer; pseudo-count to be added to expression levels to avoid $\log(0)$ (defaults to 1)
labels	character; vector specifying the cell type of each column of data
cell.clusters	integer; number of cell subpopulations
drop_thre	numeric; between 0 and 1 specifying the threshold to determine dropout values
type	A character specifying the type of values in the expression matrix. Can be 'count' or 'TPM'
cores	integer; number of cores used for paralell computation
BPPARAM	parallel back-end to be used during parallel computation. See BiocParallelParam-class.

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net.coef

matrix; network coefficients. Please provide if you don't want to use ADImpute's network model. Must contain one first column 'O' acconting for the intercept of the model and otherwise be an adjacency matrix with hgnc\_symbols in rows and columns. Doesn't have to be squared. See ADImpute::demo\_net for a small example.

net.implementation

character; either 'iteration', for an iterative solution, or 'pseudoinv', to use Moore-Penrose pseudo-inversion as a solution. 'pseudoinv' is not advised for

big data.

tr.length matrix with at least 2 columns: 'hgnc\_symbol' and 'transcript\_length'

bulk vector of reference bulk RNA-seq, if available (average across samples)

... additional parameters to pass to network-based imputation

#### **Details**

For each gene, a fraction (mask.ratio) of the quantified expression values are set to zero and imputed according to 3 different methods: scImpute, baseline (average gene expression across all cells) or a network-based method. The imputation error is computed for each of the values in the original dataset that was set to 0, for each method. The method resulting in a lowest imputation error for each gene is chosen.

#### Value

- if sce is provided: returns a SingleCellExperiment with the best performing method per gene stored as row-features. Access via SingleCellExperiment::int\_elementMetadata(sce)\$ADImpute\$methods.
- if see is not provided: returns a character with the best performing method in the training set for each gene

# See Also

ImputeBaseline, ImputeDrImpute, ImputeNetwork

#### **Examples**

```
# Normalize demo data
norm_data <- NormalizeRPM(ADImpute::demo_data)
method_choice <- EvaluateMethods(norm_data, do = c('Baseline','DrImpute'),
cores = 2)</pre>
```

GetDropoutProbabilities

Get dropout probabilities

# Description

GetDropoutProbabilities computes dropout probabilities (probability of being a dropout that should be imputed rather than a true biological zero) using an adaptation of scImpute's approach

# Usage

```
GetDropoutProbabilities(data, thre, cell.clusters, labels = NULL,
type = 'count', cores, BPPARAM, genelen = ADImpute::transcript_length)
```

# Arguments

data	matrix; original data before imputation
thre	numeric; probability threshold to classify entries as biological zeros
cell.clusters	integer; number of cell subpopulations
labels	character; vector specifying the cell type of each column of data
type	A character specifying the type of values in the expression matrix. Can be 'count' or 'TPM'
cores	integer; number of cores used for paralell computation
BPPARAM	parallel back-end to be used during parallel computation. See BiocParallelParam-class.
genelen	matrix with at least 2 columns: 'hgnc_symbol' and 'transcript_length'

# **Details**

This function follows scImpute's model to distinguish between true biological zeros and dropouts, and is based on adapted code from the scImpute R package.

#### Value

matrix with same dimensions as data containing the dropout probabilities for the corresponding entries

HandleBiologicalZeros Get dropout probabilities

#### Description

GetDropoutProbabilities computes dropout probabilities (probability of being a dropout that should be imputed rather than a true biological zero) using an adaptation of scImpute's approach

#### Usage

```
HandleBiologicalZeros(data, imputed, thre = 0.5, cell.clusters,
labels = NULL, type = 'count', cores = BiocParallel::bpworkers(BPPARAM),
BPPARAM = BiocParallel::SnowParam(type = "SOCK"),
genelen = ADImpute::transcript_length, prob.mat = NULL)
```

#### **Arguments**

data matrix; original data before imputation

imputed list; imputation results for considered methods

thre numeric; between 0 and 1 specifying the threshold to determine dropout values

cell.clusters integer; number of cell subpopulations

labels character; vector specifying the cell type of each column of data

type A character specifying the type of values in the expression matrix. Can be

'count' or 'TPM'

cores integer; number of cores used for paralell computation

BPPARAM parallel back-end to be used during parallel computation. See BiocParallelParam-class.

genelen matrix with at least 2 columns: 'hgnc\_symbol' and 'transcript\_length'

prob.mat matrix with same dimensions as data containing the dropout probabilities for

the corresponding entries

#### **Details**

This function follows scImpute's model to distinguish between true biological zeros and dropouts, and is based on adapted code from the scImpute R package.

#### Value

list with 2 components: zerofiltered, a list equivalent to imputed but with entries of imputed likely biological zeros set back to zero, and dropoutprobabilities matrix with same dimensions as data containing the dropout probabilities for the corresponding entries

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Impute Dropout imputation using different methods	Impute	Dropout imputation using different methods
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# Description

Impute performs dropout imputation on normalized data, based on the choice of imputation methods.

# Usage

```
Impute(data, sce = NULL, do = 'Ensemble', write = FALSE,
outdir = getwd(), method.choice = NULL, scale = 1, pseudo.count = 1,
labels = NULL, cell.clusters = 2, drop_thre = NULL, type = 'count',
tr.length = ADImpute::transcript_length,
cores = BiocParallel::bpworkers(BPPARAM),
BPPARAM = BiocParallel::SnowParam(type = "SOCK"),
net.coef = ADImpute::network.coefficients, net.implementation = 'iteration',
bulk = NULL, true.zero.thr = NULL, prob.mat = NULL, ...)
```

#### **Arguments**

data	matrix; raw counts (genes as rows and samples as columns)
sce	SingleCellExperiment; normalized counts and associated metadata.
do	character; choice of methods to be used for imputation. Currently supported methods are 'Baseline', 'DrImpute', 'Network', and 'Ensemble'. Defaults to 'Ensemble'. Not case-sensitive. Can include one or more methods. Non-supported methods will be ignored.
write	logical; write intermediary and imputed objects to files?
outdir	character; path to directory where output files are written. Defaults to working directory
method.choice	character; best performing method in training data for each gene
scale	integer; scaling factor to divide all expression levels by (defaults to 1)
pseudo.count	integer; pseudo-count to be added to expression levels to avoid $log(0)$ (defaults to 1)
labels	character; vector specifying the cell type of each column of data
cell.clusters	integer; number of cell subpopulations
drop_thre	numeric; between 0 and 1 specifying the threshold to determine dropout values
type	A character specifying the type of values in the expression matrix. Can be 'count' or 'TPM'
tr.length	matrix with at least 2 columns: 'hgnc_symbol' and 'transcript_length'
cores	integer; number of cores used for paralell computation
BPPARAM	parallel back-end to be used during parallel computation. See BiocParallelParam-class.

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net.coef

matrix; network coefficients. Please provide if you don't want to use ADImpute's network model. Must contain one first column 'O' acconting for the intercept of the model and otherwise be an adjacency matrix with hgnc\_symbols in rows and columns. Doesn't have to be squared. See ADImpute::demo\_net for a small example.

net.implementation

character; either 'iteration', for an iterative solution, or 'pseudoinv', to use Moore-Penrose pseudo-inversion as a solution. 'pseudoinv' is not advised for big data.

bulk vector of reference bulk RNA-seq, if available (average across samples)

true.zero.thr if set to NULL (default), no true zero estimation is performed. Set to numeric

value between 0 and 1 for estimation. Value corresponds to the threshold used to determine true zeros: if the probability of dropout is lower than true.zero.thr,

the imputed entries are set to zero.

prob.mat matrix of the same size as data, filled with the dropout probabilities for each

gene in each cell

... additional parameters to pass to network-based imputation

#### **Details**

Values that are 0 in data are imputed according to the best-performing methods indicated in method.choice. Currently supported methods are:

- Baseline: imputation with average expression across all cells in the dataset. See ImputeBaseline.
- Previously published approaches: DrImpute and SAVER.
- Network: leverages information from a gene regulatory network to predicted expression of genes that are not quantified based on quantified interacting genes, in the same cell. See ImputeNetwork.
- Ensemble: is based on results on a training subset of the data at hand, indicating which method best predicts the expression of each gene. These results are supplied via method.choice. Applies the imputation results of the best performing method to the zero entries of each gene.

If 'Ensemble' is included in do, method.choice has to be provided (use output from EvaluateMethods()). Impute can create a directory imputation containing the imputation results of all methods in do. If true.zero.thr is set, dropout probabilities are computed using scImpute's framework. Expression values with dropout probabilities below true.zero.thr will be set back to 0 if imputed, as they likely correspond to true biological zeros (genes not expressed in cell) rather than technical dropouts (genes expressed but not captured). If sce is set, imputed values by the different methods are added as new assays to sce. Each assay corresponds to one imputation method. If true.zero.thr is set, only the values after filtering for biological zeros will be added. This is different from the output if sce is not set, where the original values before filtering and the dropout probability matrix are returned.

#### Value

• if sce is not set: returns a list of imputation results (normalized, log-transformed) for all selected methods in do. If true.zero.thr is defined, returns a list of 3 elements: 1) a list,

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imputations, containing the direct imputation results from each method; 2) a list, zerofiltered, containing the results of imputation in imputations after setting biological zeros back to zero; 3) a matrix, dropoutprobabilities, containing the dropout probability matrix used to set biological zeros.

• if sce is set: returns a SingleCellExperiment with new assays, each corresponding to one of the imputation methods applied. If true.zero.thr is defined, the assays will contain the results after imputation and setting biological zeros back to zero.

#### See Also

EvaluateMethods, ImputeBaseline, ImputeDrImpute, ImputeNetwork, ImputeSAVER

#### **Examples**

```
# Normalize demo data
norm_data <- NormalizeRPM(demo_data)
# Impute with particular method(s)
imputed_data <- Impute(do = 'Network', data = norm_data[,1:10],
net.coef = ADImpute::demo_net)
imputed_data <- Impute(do = 'Network', data = norm_data[,1:10],
net.implementation = 'pseudoinv', net.coef = ADImpute::demo_net)</pre>
```

ImputeBaseline

Impute using average expression across all cells

#### **Description**

ImputeBaseline imputes dropouts using gene averages across cells. Zero values are excluded from the mean computation.

#### **Usage**

```
ImputeBaseline(data, write = FALSE, ...)
```

#### **Arguments**

data	matrix with entries equal to zero to be imputed, normalized and log2-transformed
	(genes as rows and samples as columns)
write	logical; should a file with the imputation results be written?
	additional arguments to saveRDS

# Value

matrix; imputation results considering the average expression values of genes

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ImputeDrImpute

Use DrImpute

# **Description**

ImputeDrImpute uses the DrImpute package for dropout imputation

# Usage

```
ImputeDrImpute(data, write = FALSE)
```

# Arguments

data

matrix with entries equal to zero to be imputed, normalized and log2-transformed

(genes as rows and samples as columns)

write

logical; should a file with the imputation results be written?

#### Value

matrix; imputation results from DrImpute

# See Also

DrImpute

ImputeNetParallel

Network-based parallel imputation

# **Description**

ImputeNetParallel implements network-based imputation in parallel

# Usage

```
ImputeNetParallel(drop.mat, arranged, cores =
BiocParallel::bpworkers(BPPARAM), type = 'iteration', max.iter = 50,
BPPARAM = BiocParallel::SnowParam(type = "SOCK"))
```

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# Arguments

drop.mat	matrix, logical; dropout entries in the data matrix (genes as rows and samples as columns)
arranged	list; output of ArrangeData
cores	integer; number of cores used for paralell computation
type	character; either 'iteration', for an iterative solution, or 'pseudoinv', to use Moore-Penrose pseudo-inversion as a solution.
max.iter	numeric; maximum number of iterations for network imputation. Set to -1 to remove limit (not recommended)
BPPARAM	parallel back-end to be used during parallel computation. See BiocParallelParam-class.

# Value

matrix; imputation results incorporating network information

ImputeNetwork	Network-based imputation	

# Description

Network-based imputation

# Usage

```
ImputeNetwork(data, net.coef = NULL,
cores = BiocParallel::bpworkers(BPPARAM),
BPPARAM = BiocParallel::SnowParam(type = "SOCK"),
type = 'iteration', write = FALSE, ...)
```

# Arguments

data	matrix with entries equal to zero to be imputed, normalized and log2-transformed (genes as rows and samples as columns)
net.coef	matrix; network coefficients.
cores	integer; number of cores to use
BPPARAM	$parallel\ back-end\ to\ be\ used\ during\ parallel\ computation.\ See\ {\tt BiocParallelParam-class}.$
type	character; either 'iteration', for an iterative solution, or 'pseudoinv', to use Moore-Penrose pseudo-inversion as a solution.
write	logical; should a file with the imputation results be written?
	additional arguments to ImputeNetParallel

20 ImputeNPDropouts

# **Details**

Imputes dropouts using a gene regulatory network trained on external data, as provided in net.coef. Dropout expression values are estimated from the expression of their predictor genes and the network coefficients.

# Value

matrix; imputation results incorporating network information

#### See Also

ImputeNetParallel

ImputeNPDropouts

Helper function to PseudoInverseSolution\_percell

# **Description**

ImputeNPDropouts computes the non-dropout- dependent solution of network imputation for each cell

# Usage

```
ImputeNPDropouts(net, expr)
```

# Arguments

net matrix, logical; network coefficients for all dropout (to be imputed) genes that

are predictive of the expression of other dropout genes

expr numeric; vector of gene expression for all genes in the cell at hand

# Value

vector; imputation results for the non-dropout-dependent genes

 ${\tt ImputePredictiveDropouts}$ 

Helper function to PseudoInverseSolution\_percell

# **Description**

ImputePredictiveDropouts applies Moore-Penrose pseudo-inversion to compute the dropout-dependent solution of network imputation for each cell

# Usage

ImputePredictiveDropouts(net, thr = 0.01, expr)

#### **Arguments**

net matrix, logical; network coefficients for all dropout (to be imputed) genes that

are predictive of the expression of other dropout genes

thr numeric; tolerance threshold to detect zero singular values

expr numeric; vector of gene expression for all genes in the cell at hand

#### Value

vector; imputation results for the dropout-dependent genes

ImputeSAVER	Use SAVER

# Description

ImputeSAVER uses the SAVER package for dropout imputation

# Usage

```
ImputeSAVER(data, cores, try.mean = FALSE, write = FALSE)
```

#### **Arguments**

data	matrix with entries ed	rual to zero to be imputed.	normalized (genes as rows and

samples as columns)

cores integer; number of cores to use

try.mean logical; whether to additionally use mean gene expression as prediction

write logical; should a file with the imputation results be written?

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#### Value

matrix; imputation results from SAVER

#### See Also

saver

MaskData

Masking of entries for performance evaluation

# Description

MaskData sets a portion (mask) of the non-zero entries of each row of data to zero

# Usage

```
MaskData(data, write.to.file = FALSE, mask = .1)
```

# **Arguments**

data matrix; raw counts (genes as rows and samples as columns)

write.to.file logical; should the output be written to a file?

mask numeric; ratio of total non-zero samples to be masked per gene (defaults to .1)

# **Details**

Sets a portion (mask) of the non-zero entries of each row of data to zero. Result is written to filename.

# Value

matrix containing masked raw counts (genes as rows and samples as columns)

MaskerPerGene

Helper mask function

# Description

Helper mask function, per feature.

# Usage

```
MaskerPerGene(x, rowmask)
```

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#### **Arguments**

x logical; data to mask

rowmask numeric; number of samples to be masked per gene

#### Value

logical containing positions to mask

# **Description**

Gene Regulatory Network used by ADImpute's Network imputation method. First column, 0, corresponds to the intercept of a gene--specific prediction model. The remaining rows and columns correspond to the adjacency matrix of the inferred network, where rows are target genes and columns are predictors. Genes are identified by their hgnc\_symbol.

#### Usage

```
network.coefficients
```

#### **Format**

dgCMatrix

NormalizeRPM

RPM normalization

# **Description**

NormalizeRPM performs RPM normalization, with possibility to log the result

# Usage

```
NormalizeRPM(data, sce = NULL, log = FALSE, scale = 1, pseudo.count = 1)
```

#### **Arguments**

data matrix; raw data (genes as rows and samples as columns)

sce SingleCellExperiment; raw data

log logical; log RPMs?

scale integer; scale factor to divide RPMs by

pseudo.count numeric; if log = TRUE, value to add to RPMs in order to avoid taking log(0)

24 NormalizeTPM

#### Value

```
matrix; library size normalized data
```

# **Examples**

```
demo <- NormalizeRPM(ADImpute::demo_data)</pre>
```

NormalizeTPM

TPM normalization

# **Description**

NormalizeTPM performs TPM normalization, with possibility to log the result

#### Usage

```
NormalizeTPM(data, sce = NULL, tr_length = NULL, log = FALSE, scale = 1, pseudo.count = 1)
```

#### **Arguments**

data matrix; raw data (genes as rows and samples as columns)

sce SingleCellExperiment; raw data

tr\_length data.frame with at least 2 columns: 'hgnc\_symbol' and 'transcript\_length'

log logical; log TPMs?

scale integer; scale factor to divide TPMs by

pseudo.count numeric; if log = T, value to add to TPMs in order to avoid taking log(0)

#### **Details**

Gene length is estimated as the median of the lengths of all transcripts for each gene, as obtained from biomaRt. Genes for which length information cannot be found in biomaRt are dropped.

#### Value

matrix; normalized data (for transcript length and library size)

#### **Examples**

```
demo <- NormalizeTPM(ADImpute::demo_data)</pre>
```

PseudoInverseSolution\_percell

Network-based parallel imputation - Moore-Penrose pseudoinversion

# **Description**

PseudoInverseSolution\_percell applies Moore-Penrose pseudo-inversion to compute the solution of network imputation for each cell

# Usage

```
PseudoInverseSolution_percell(expr, net, drop_ind, thr = 0.01)
```

# Arguments

expr numeric; expression vector for cell at hand

net matrix; network coefficients

drop\_ind logical; dropout entries in the cell at hand

thr numeric; tolerance threshold to detect zero singular values

#### Value

matrix; imputation results incorporating network information

ReadData Data read

# **Description**

ReadData reads data from raw input file (.txt or .csv)

# Usage

```
ReadData(path, ...)
```

# Arguments

```
path character; path to input file
... additional arguments to data.table::fread()
```

#### Value

matrix; raw counts (genes as rows and samples as columns)

26 ReturnOut

ReturnChoice	Wrapper for return of EvaluateMethods()

# Description

ReturnChoice Adjusts the output of EvaluateMethods to a character vector or a SingleCellExperiment object. Helper function to ADImpute.

#### Usage

ReturnChoice(sce, choice)

#### **Arguments**

sce SingleCellExperiment; a SingleCellExperiment object if available; NULL oth-

erwise

choice character; best performing method in the training set for each gene

#### Value

• if sce is provided: returns a SingleCellExperiment with the best performing method per gene stored as row-features. Access via SingleCellExperiment::int\_elementMetadata(sce)\$ADImpute\$methods.

• if sce is not provided: returns a character with the best performing method in the training set for each gene

ReturnOut Wrapper for return of Impute()

# Description

ReturnOut Adjusts the output of Impute to a list of matrices or a SingleCellExperiment object. Helper function to ADImpute.

# Usage

```
ReturnOut(result, sce)
```

#### **Arguments**

result list; imputation result

sce SingleCellExperiment; a SingleCellExperiment object if available; NULL oth-

erwise

#### Value

imputation results. A SingleCellExperiment if !is.null(sce), or a list with imputed results in matrix format otherwise.

SetBiologicalZeros 27

# Description

SetBiologicalZeros sets some of the entries back to zero after dropout imputation, as they likely correspond to true biological zeros (genes not expressed in given cell)

# Usage

```
SetBiologicalZeros(imputation, drop_probs, thre = .2, was_zero)
```

# **Arguments**

imputation	matrix; imputed values
drop_probs	matrix; dropout probabilities for each entry in imputation. 0 means certain biological zero, while 1 means certain dropout to be imputed
thre	numeric; probability threshold to classify entries as biological zeros
was_zero	matrix; logical matrix: was the corresponding entry of imputation originally a zero?

# **Details**

Entries which were originally zero and have dropout probability below thre are considered biological zeros and, if they were imputed, are set back to 0.

#### Value

matrix containing likely biological zeros set back to 0.

SplitData	Selection of samples for training	

#### **Description**

SplitData selects a portion (ratio) of samples (columns in data) to be used as training set

# Usage

```
SplitData(data, ratio = .7, write.to.file = FALSE, train.only = TRUE)
```

28 transcript\_length

# **Arguments**

data matrix; raw counts (genes as rows and samples as columns)

ratio numeric; ratio of the samples to be used for training

write.to.file logical; should the output be written to a file?

train.only logical; if TRUE define only a training dataset, if FALSE writes both training

and validation sets (defaults to TRUE)

# **Details**

Selects a portion (ratio) of samples (columns in data) to be used as training set and writes to file 'training\_raw.txt'.

#### Value

matrix containing raw counts (genes as rows and samples as columns)

# **Description**

A data.frame to be used for transcript length computations. May be necessary upon TPM normalization, or as input to scImpute. All data was retrieved from biomaRt.

# Usage

transcript\_length

#### **Format**

A data.frame with 2 columns:

hgnc\_symbol Gene symbol identifier

transcript length Length of transcript

WriteCSV 29

WriteCSV

Write csv file

# Description

WriteCSV writes data to a comma-delimited output file

# Usage

```
WriteCSV(object, file)
```

# Arguments

object

R object to write

file

character; path to output file

#### Value

Returns NULL

# **Examples**

```
file <- tempfile()
WriteCSV(iris, file = file)</pre>
```

WriteTXT

Write txt file

# Description

WriteTXT writes data to a tab-delimited output file

# Usage

```
WriteTXT(object, file)
```

# Arguments

object

R object to write

file

character; path to output file

# Value

Returns NULL

WriteTXT

# Examples

```
file <- tempfile()
WriteTXT(iris, file = file)</pre>
```

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